

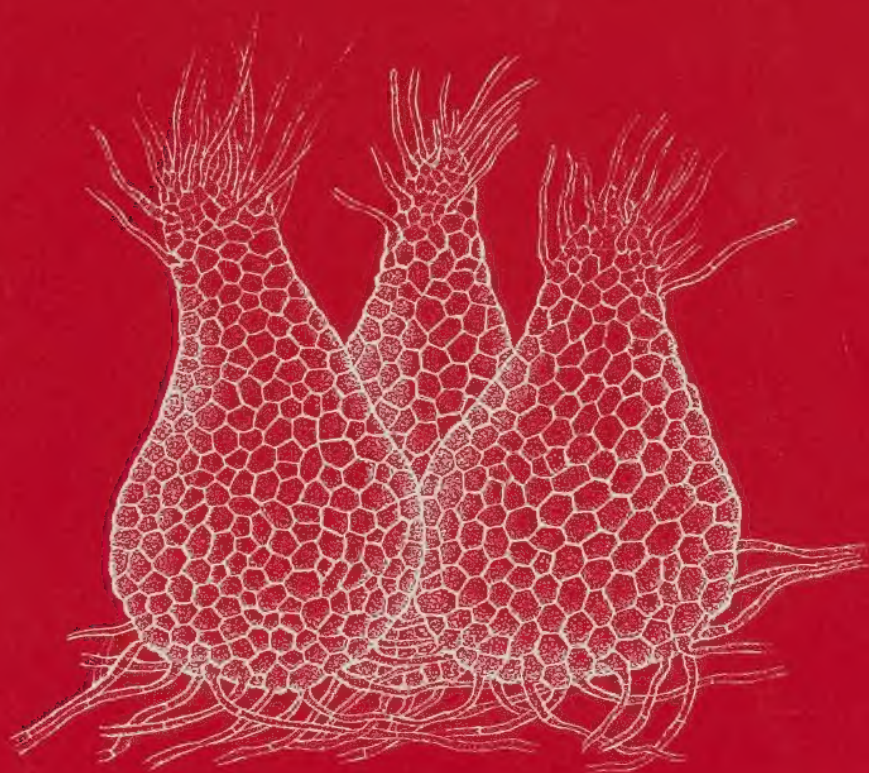
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Athanase Michel SACCAS
1911-1985

A. COUTÉ¹ et D. LAMY²

¹ Muséum National d'Histoire Naturelle, Laboratoire de Cryptogamie,
12 rue Buffon, F-75005 Paris

² UMS-CNRS 826, Lab. Cryptogamie, 12 rue Buffon,
F-75005 Paris — e-mail lamy@mnhn.fr



Athanase Michel SACCAS est né à Janina, en Grèce, le 10 mai 1911. À la fin de ses études secondaires, il s'inscrit à l'Ecole Nationale d'Horticulture de Patras (Grèce) d'où il sort diplômé en 1931. De 1931 à 1933, il occupe un poste de chef de travaux dans ce même établissement et y exerce comme professeur d'horticulture de 1933 à 1938.

Abandonnant alors son pays natal, Athanase SACCAS vient en France où il intègre l'Ecole Nationale d'Agriculture de Grignon. Il obtient son diplôme d'ingénieur agricole en 1941 et est alors recruté, comme attaché de recherches, par le CNRS, détaché au laboratoire de Cryptogamie du Muséum National d'Histoire Naturelle (jusqu'en 1948).

En 1943, il soutient, à l'Université de Paris, une thèse d'ingénieur-docteur en sciences naturelles dont le sujet porte sur la morphologie et la biologie des *Fusicladium* des Rosacées, champignons responsables des tavelures des Aubépines, Poiriers et Pommiers.

Fort de ses compétences tant en Biologie et Cytologie végétales qu'en Mycologie et Pathologie végétale, il est nommé chef de travaux de Pathologie Végétale, Anatomie et Biologie des champignons à l'Office de la Recherche Scientifique d'Outre-mer, rôle qu'il assume de 1946 à 1948.

Il est alors envoyé en République Centrafricaine, au Centre de Recherches Agronomiques de Boukoko, avec le titre de Maître de Recherche de 3^e classe. Il y devient successivement Maître de Recherche de 2^{ème} classe en 1956 et, enfin, Directeur de Laboratoire de 2^e classe en 1957 et Directeur de Recherche, en 1960, dans le cadre de l'ORS-TOM. Durant cette période, il assume, à la station de Boukoko, de 1948 à 1952, les fonctions de chef du Service des recherches et des laboratoires et de chef de la division Phytopathologie-Entomologie, de 1957 à 1958 celles de directeur par interim et, à partir de 1958, il devient directeur, poste qu'il occupe jusqu'en 1983. Il se retire en Savoie, région d'où est originaire sa femme. Il y meurt le 20 juillet 1985.

Auteur de plus de quatre-vingt publications toutes d'excellente facture et d'une bonne cinquantaine de notes et rapports, Athanase SACCAS a montré sans relâche ses qualités de phytopathologiste. Il s'est intéressé à de nombreuses maladies cryptogamiques de phanérogames d'intérêt agronomique. Abordant, dans ses débuts, les problèmes des rouilles de Graminées et de Céréales (maïs, riz, sorgho...), il s'est penché ensuite sur les champignons papyriques (*Beauveria heimii*, *Sepedonium chartarum*...). Poursuivant ses travaux sur les rouilles, il s'est orienté également vers les parasites mycologiques de l'arachide, des cacaoyers, caféiers (anthracnose, cercosporiose, fusariose, trachéomycose...), cannes à sucre, colatiers, cotonniers, hévéas, maniocs, palmiers à huile, poivriers, sésames....

Ses recherches lui permirent ainsi de mettre au point des techniques de traitement et de décrire, en même temps, bon nombre de nouvelles espèces. Il a publié plusieurs ouvrages portant sur les maladies cryptogamiques des plantes cultivées, en particulier du café, pour l'Afrique Centrale et l'Afrique Equatoriale.

Homme de terrain très actif, Athanase SACCAS ne s'est pas cantonné au périmètre de la station de recherches de Boukoko. Il a, en effet, effectué de nombreuses tournées et missions en Algérie, Brésil, Cameroun, Congo Belge, Côte d'Ivoire, Gabon, Israël, Mali, Tchad et Zaïre. Au cours de certains déplacements, il a aussi fait profiter de son expérience en Phytopathologie et Cryptogamie, de jeunes chercheurs et étudiants africains en dispensant un enseignement de grande qualité. Il a été le représentant de la République Centrafricaine, de l'IFCC et même de l'UNESCO dans de nombreuses manifestations internationales.

L'importance des travaux d'Athanase SACCAS, tant par leur qualité que leur quantité, et leur impact sur les activités économiques des différents pays africains où il est intervenu, ont été reconnus sans conteste par ses autorités de tutelle. Il a ainsi été lauréat de l'Académie d'Agriculture (1946) puis lauréat du Conseil Supérieur de la Recherche Scientifique et du Progrès Technique (1956). Il a été fait Chevalier (1951) puis Officier

(1963) de l'Ordre du Mérite Agricole et Chevalier (1960) de l'Ordre des Palmes Académiques. Il a aussi été décoré de l'Ordre de la Légion d'Honneur (Chevalier en 1960 et Officier en 1970) et de l'Ordre du Mérite Français d'Outre-Mer.

Les pays africains qui ont bénéficié de ses travaux et compétences lui ont aussi marqué leur reconnaissance de façon éclatante. C'est ainsi qu'il a été fait Grand Chancelier et Grand Croix (1959) de l'Ordre National du Mérite Centrafricain, Commandeur (1963) de l'Ordre du Mérite Agricole du même pays qui l'a également honoré du grade de Commandeur (1964) de l'Ordre des Palmes Académiques de la République Centrafricaine. Il a aussi été élevé au grade de Grand Officier (1965) de l'Ordre de la République Tunisienne et de Commandeur (1969) de l'Ordre du Mérite Tchadien.

Ce palmarès éloquent montre à l'évidence la remarquable activité scientifique menée par Athanase SACCAS. Il a toujours fait preuve d'un sens profond du service public allié à une passion sans limite pour les pays africains auxquels il a consacré toute son énergie.

L'herbier, les notes et les manuscrits de Athanase SACCAS ont été déposés au Laboratoire de Cryptogamie du Muséum National d'Histoire Naturelle (PC).

Publications de A.M. Saccas

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1969

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1971

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1972

- SACCAS A M La rouille «farineuse» des caféiers, due à *Hemileia coffeicola* Malb. et Rog *Bulletin de l'Institut Français du Café et du Cacao* 1972, 11 : 1-68, 32 fig

1975

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1981

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- SACCAS A M Essais comparatifs de désinfection des semences par voie humide et sèche contre les maladies charbonneuses des sorghos à la Station Centrale de Boukoko (A.E.F.) *Congrès de la Protection des végétaux et de leurs produits sous les climats chauds* Marseille, Septembre 1954, 13p. [non vu]

- SACCAS A M Résultats des essais de lutte contre la «pourriture brune» due à *Phytophthora palmifera*, à «pourriture noire» due à *Botryodiplodia theobromae* et la «pourriture farineuse» due à *Trachysphaera fructigena* *Congrès de la Protection des végétaux et de leurs produits sous les climats chauds*, Marseille, Septembre 1954, 7p. [non vu]

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Deux références, données par Saccas dans sa Notice de Travaux demeurent problématiques, pour lesquelles toute information sera la bienvenue :

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RÉPERTOIRE DES DONNÉES UTILES POUR EFFECTUER LES TESTS D'INTERCOMPATIBILITÉ CHEZ LES BASIDIOMYCÈTES.

VII — APHYLLOPHORALES NON PORÉES (DEUXIÈME SUPPLÉMENT)

Jacques BOIDIN* & Paule LANQUETIN

* 17 rue Duguesclin, 69006 Lyon, France

Cette note fait suite aux 6 articles parus dans cette même revue (Boidin & Lanquetin 1984 a, b et c, Capellano, 1985, Lamoure, 1989, Boidin, 1990).

Pour la signification détaillée des signes, nous renvoyons le lecteur à l'introduction, ou aux résumés situés en tête des parties III et V (Boidin & Lanquetin, 1984c, Lamoure, 1989). Rappelons simplement : h, hétérothalle, II b, polaire, IV, tétrapolaire, H, homothalle, H, homothalle présumé, P, parthénogénétique, A, amphithalle, u, uninuclée, d, dicaryotique, p, plurinucléé, m, multinucléé, u/d signifie que la culture monosperme passe spontanément du stade uninuclée au stade dicaryotique car elle est homothalle.

Pour les boucles : b, présentes sans précision, a, absentes, c, constantes, i, inconstantes, r, rares, va, variables selon l'aération, o, parfois opposées, v, verticillées sur les hyphes les plus larges.

Comportements nucléaires : N, normal, SN, subnormal, He, hétérocaryotique, HC, holocenocaryotique, As, astatocenocaryotique, HM, holomonocaryotique, HD, holodicyotique.

Vitesse de croissance : nombre de semaines pour que le rayon de la culture couvre la boîte de Petri de 90 mm de diamètre, 7, plus de 6 semaines.

L'astérisque signifie « avec irrégularité », p. ex. d*, dicaryotique avec quelques articles à 1, ou à 3 ou 4 noyaux.

Nous ne résumerons que les données nouvelles ou complémentaires des index publiés (1984 et 1990). Le *C. sticticum pini-canadense* figure dans l'index de 1990 mais sa polarité n'était pas connue, découverte depuis, elle sera indiquée ici.

ESPÈCES	Thallie	Basidiospores	Monosperme	Poly sperme	Comportement nucléaire	Boucles	Vitesse de croissance	Arthrospores ou conidies	RÉFÉRENCES
<i>acanthophysata</i> Léger <i>Hymenochaete</i>	H	1	p	p	HC	a	7		27
<i>acerina</i> Peck <i>Phlebia</i>	hII				As	va			33
<i>aculeatum</i> Wu <i>Gloeocystidiellum</i>				d		a	7		39
<i>aculeatus</i> Mackawa <i>Kerinomyces</i>	hIV	1				c	7		28
<i>adjacens</i> Boid., Lanq. & Gilles <i>Dendrophora</i>	H	1	u/d*	d*	N	r	2		8
<i>afibulatum</i> (G. H. Cunn.) Boid. et al <i>Megalocystidium</i>	hIV	2	p	d	He	c	7		12
<i>africanum</i> Boid., Lanq. & Gilles <i>Megalocystidium</i>	hII	2	p	d*	He	c	7		12
<i>albicans</i> (Pers.) Nakas. <i>Glyphoderma</i>	hII					c	4		29
<i>albida</i> Post : Fr. <i>Phlebia</i>	hII					va	2-3		29
<i>Alni</i> Boid., Lanq. & Gilles <i>Vuilleminia</i>	hIV	2	p	d	He	c	6-7		11
<i>americanus</i> Nakas. et al <i>Ceraceomyces</i>	hII					c			30
<i>arachnoideum</i> Langer <i>botryobasidium</i>				d		a			22
<i>aspellum</i> Hjortst. <i>Gloeocystidiellum</i>	hII	1	u	d	N	c	7		39
<i>asterospora</i> Boid. & Lanq. <i>Dendrothele</i>	hII	1*	u	d	N	c	7		7
<i>bicornis</i> Hjortst. & Ryv <i>Dendrophora</i>	H	1	u/d	d*	N	r	2-3		8
<i>bispora</i> Burds. & Nakas. "Dendrothele"	A	1*	u	d	N	c	3		7

<i>bisporum</i> Boid., Lanq. & Gilles <i>Gloeocystidiellum</i>	P	1	u	u	HM	a	7		12
<i>borbonica</i> Boid., Lanq. & Gilles <i>Boidinia</i>	h	1	u	d*	N	a	4-6		12
<i>aff. borbonica</i> Boid., Lanq. & Gilles <i>Boidinia</i>	H	1	u	d	N	a	4-5		12
<i>bosei</i> De <i>Epithelopsis</i>	hII								14
<i>botryodeum</i> (Overh.) Parm. <i>Botryobasidium</i>	H		p	p					22
<i>brunnea</i> Wu <i>Phanerochaete</i>		2	m	m	HC	o	2		38
<i>burtii</i> (Romell) Parm. <i>Phanerochaete</i>						v	2-3		29
<i>cana</i> Wu <i>Boidinia</i>	H	1	u, d	d	N	a	4-5		12
<i>canariensis</i> (Manj. & Mor.) Hjortst. <i>Dendrothele</i>	hIV	1	u	d	N	c	7		7
<i>candidans</i> Erikss. <i>Botryobasidium</i>	H		d	d	HD	a			22
<i>carnea</i> (Burt) Parm. <i>Phanerochaete</i>						v	3		29
<i>carpaica</i> Pilat <i>Hymenochaete</i>	H	1	d	d	HD	a	7		27
<i>centrifuga</i> Karst <i>Phlebia</i>	hII	2	m	md	As	va	2		29-23
<i>cervicour</i> (Berk. & Curt.) Massée <i>Asterostroma</i>						a	6-7		29
<i>chrysosporum</i> Burds. <i>Phanerochaete</i>	H?					a	1-2	ar	29
<i>ciliatigerum</i> (Hohn. & F. Sch.) Nakas. <i>Gloeocystidiellum</i>	hIV	1	u	d	N	c	7		12
<i>coffeeana</i> Leg. & Lanq. <i>Hymenochaete</i>	h	1	u*	d	N	a	4		26
<i>columbiensis</i> (Burt) Burds. & Lomb. <i>Gloeodermia</i>	hII ou H								17
<i>compactum</i> Wu <i>Gloeocystidiellum</i>	h	2	u	d	SN	c	5-6		39
<i>conicum</i> (Oberw.) Erikss. & Hjortst. <i>Repetobasidium</i>	hII	1	u	d	N	c	7		16
<i>consersum</i> Erikss. <i>Botryobasidium</i>				d		a			22

l = sensu Hallenberg, voir *vesiculosum*

<i>contiformis</i> G. H. Cunn. <i>Hymenochaete</i>	H		d	d	HD	a			27
<i>crassitunicata</i> Boid., Lanq. & Gilles <i>Peniophora</i>	hIV	1	u	d	N	c	2-3		8
<i>curtisi</i> Hallenb. <i>Botryobasidium</i>				d		a			22
<i>denticulata</i> Lég. & Lanq. <i>Hymenochaete</i>	H	1	d*	d*	HD	a	7		24
<i>duportii</i> Pat. <i>Hydnochaete</i>		1	m	m	HC	a	7		27
<i>elaetidis</i> Boid., Lanq. & Gilles <i>Peniophora</i>	H?	1	u/d*	d*	N	r	2		8
<i>erikssonii</i> Hall. & Hj. <i>Hypochnicium</i>	hIV	1	u	d	n	c	4		18
<i>exilis</i> (Burt) Burds. <i>Phanerochaete</i>						v	2-3		29
<i>fumaceus</i> Boid., Lanq. & Gilles <i>Amuromyces</i>	h	1	u	d	N	c	7		10
<i>fimbriatum</i> Burds. <i>Gloeocystidiellum</i>				d		a	3-4		12
<i>festus</i> Boid., Lanq. & Gilles <i>Peniophora</i>	hIV	1	u	d	N	c	3-4		8
<i>formosanum</i> Wu <i>Gloeocystidiellum</i>	hII	2	u	d	SN	c	7		39
<i>fouquieriae</i> Nakas. & Gubb <i>Hypoderma</i>	hII					b	4-6		31
<i>fuliginea</i> (Pers.) Lev. <i>Hymenochaete</i>	H	1	d	d	HD	a	7		27
<i>gabonensis</i> Boid., Lanq. & Gilles <i>Peniophora</i>	H	1	u/d*	d*	N	r	2		8
<i>globosa</i> Wu <i>Gilothele</i>	h	2	u	d	SN	a	7		12-39
<i>graminicola</i> Wu <i>Gloeomyces</i>				d*		ov	4		39
<i>granulata</i> Wu <i>Boidinia</i>	hII	1	u	d	N	c	6		12-39
<i>grandisporum</i> G. Langer <i>Botryobasidium</i>				d		a			22
<i>griseo-flavescens</i> (Fitsch) Erikss. & Hjortst. <i>Phlebia</i>	hII	1	u	d	N	c	7		16
<i>guadelupensis</i> Boid. & Lanq. <i>Peniophora</i>	h	1	u	d	N	r	2		8
<i>hauserlevii</i> Leger <i>Hymenochaete</i>	H	1	d	d	HD	a	7		27
<i>humilis</i> (Boid.) Boid., Lanq. & Gilles <i>Gilothele</i>	h	?	U	D	SN	a	7	ar	12
<i>incarnatum</i> Ito & Imai <i>Gloeostereum</i>	hII					b	5		34
<i>indica</i> (Thind & Rat.) Rat. <i>Metulodontia</i>	hIV								14
<i>insidiosum</i> (Bourd. & Galz.) Hallenb. <i>Conferticium</i>	H	2	m	m	HC	a	4-5	co2	12

<i>isabellinum</i> (Fr.) Rogers <i>Botryobasidium</i>	H		p	p					22
<i>kuivense</i> Hjortst. <i>Gloeocystidiellum</i>	hIV	1	u	d	N	c	5-7		12
<i>kuehneri</i> (Boid. & Lanq.) Hjortst. <i>Duportella</i>	hIV	1	u	d	N	c	3		8
<i>kuehneroides</i> Boid., Lanq. & Gilles <i>Duportella</i>	hIV	1	u	d	N	c	3		8
<i>laeve</i> (Erikss.) Parm. <i>Botryobasidium</i>	H		p	p		a			22
<i>lamellosa</i> (P. Henn.) Bres. <i>Gloiothele</i>	h	2	u	d	SN	a	7	ar	12
<i>laxa</i> (Wu) Boid., Lanq. & Gilles <i>Gloeopeniophorella</i>		1		d		a	3		12-39
<i>limbospora</i> (Bourd.) Oberw. <i>Athelopsis</i>	h	1	u	d	N	c	4		16
<i>leprosa</i> (Bourd. & Galz.) Boid., Lanq. & Gilles <i>Scopuloides</i>	hII	1	p	p	HC	i	3-4		10
<i>lilascens</i> (Bourd.) Erikss. & Hjortst. <i>Phlebia</i> gr. 1	hIV	1	u	d					
gr. 2	hIV	1	u	d					20
gr. 3	hII		u	dp					
<i>litschaueri</i> (Burt.) Erikss. & Strid <i>Hyphoderma</i>	hII					c	6-7		29
<i>longicystidium</i> (Litsch.) Nakas. <i>Crustoderma</i>	hII					i-v			15
<i>longisporum</i> G. Langer <i>Botryobasidium</i>				d		a			22
<i>lundellii</i> (Bourd.) Erikss. <i>Hypochmicium</i>	hIV					c	3-4		29
<i>luridum</i> (Bres.) Jül. <i>Megalocystidium</i>	H	2	p.d	d	He	c	7		12
<i>luteo-cylindricum</i> (Ta bot) Wu <i>Megalocystidium</i>	H	2	p.d	d	He	c	7		12
<i>macrospora</i> Wu "Boidinia"	H	2	d	d	HD	c	6		39
<i>magnoliae</i> (Berk. & Curt.) Burds. <i>Phanerochaete</i>		1	p	p	HC	v			2
<i>malayensis</i> Boid., Lanq. & Gilles <i>Peniophora</i>	h	1	u	d	N	r	3		8
<i>medioburiense</i> (Burt) Donk <i>Hyphoderma</i>	H								17
<i>melzeri</i> Pouzar <i>Cytidiella</i>	hIV					c	6		29
<i>microspora</i> Jacks. & Lemk. <i>Dendrothele</i>	hIV					c	7		29
<i>microsporella</i> Jül. <i>Grandinia</i>	hII					c	4		29
<i>minuscula</i> G. H. Cunn. <i>Hymenochaete</i>	H	1	d	d	HD	a	7		27

<i>murificum</i> Erikss. <i>Repetobasidium</i>	hII	1	u	d	N	c	7		16
<i>moesta</i> Boid., Lanq. & Gilles <i>Peniophora</i>	hIV	1	u*	d*	N	r	2		8
<i>molle</i> (Fr.) Hjortst. <i>Hypochnicium</i>	hIV	1	u	d/u	N	i			16
<i>moniliforme</i> Wu <i>Gloeocystidiellum</i>	hIV	1	u	d	N	c	6-7		12
<i>cf. montanum</i> Ginns & Fr <i>Megalocystidium</i>	H	2				c	7		12
<i>monticola</i> Boid., Lanq. & Gilles <i>Peniophora</i>	hIV	1	u	d	N	c	4-5		8
<i>mutabilis</i> (Bres.) Jul. <i>Fibulomycetes</i>	hII	1	u	d	N	c	7		16
<i>nanospora</i> Léger <i>Hymenochaete</i>		1		d*		a	4		27
<i>puveo-cremeum</i> (Hohn. & I.) Donk <i>Sistotrema</i>	hII								29
<i>obtusisporum</i> Erikss. <i>Botryobasidium</i>				d		a			22
<i>ochromarginata</i> Talbot <i>Hymenochaete</i>	H	1	d*	d*	HD	a	7		27
<i>olivascens</i> (Bres.) Larss. & Hjortst. <i>Brevicellicium</i>	H								17
<i>ovalispora</i> Boid., Lanq. & Gilles <i>Peniophora</i>	hIV	1	u	d	N	c	3-4		8
<i>parvocystidiata</i> Boid. & L. <i>Peniophora</i>	h	1	u	d*	N	r	2-3		8
<i>peroxydata</i> (Rick) Hjortst. <i>Boidinia</i>		1		d		a	4-5		12
<i>puliferum</i> Boid. & Gilles <i>Botryobasidium</i>				d		a			22
<i>pini canadense</i> (Schwein.) Gilberts <i>Cystostereum</i>	hIV						3-6		29
<i>pinnatifida</i> Burl <i>Hymenochaete</i>	H	1	d*	d*	HD	a	7		27
<i>protrusum</i> (Burl.) Nakas <i>Scytinostroma</i>	hIV								32
subsp. <i>septentrionale</i> Nakas	hIV								32
<i>pruinatum</i> (Bres.) Erikss. <i>Botryobasidium</i>	H		p	p		a			22
<i>pseudo-adusta</i> Lég. & Lanq <i>Hymenochaete</i>	H	1	p	p	HC	a	7		25
<i>pseudo-cystidiata</i> Boid., Lanq. & Gilles <i>Vutilemma</i>	hIV	2	u	d	SN	c	4-5		11
<i>puilverulenum</i> Novob <i>Sporotrichum</i>	hII*					a		col	36
<i>purpureum</i> Wu <i>Gloeocystidiellum</i>	hIV	1	u	d	N	c	4		39
<i>puteana</i> (Schum. : Fr.) Karst. <i>Coniophora</i>	hII								1

<i>queletii</i> (Bourd. & Galz.) Christ <i>Phlebia</i>	hII					va	3-5		29
<i>raduloides</i> (Karst.) Donk <i>Sistotrema</i>	hII								29
<i>ravum</i> (Burt) Ginn & Freeman <i>Conferticium</i>	H	2	m	m	HC	ov	3-4		39
<i>remispora</i> Boid., Lanq. & Gilles <i>Duportella</i>	H?	1	u.d?	d	N	c	7		8
<i>sacratum</i> (G. H. Cunn.) Wu <i>Dextrinocystidium</i>						ov			35
<i>salmonae</i> (Burt) Boid., Lanq. & Gilles <i>Gloeocystidiopsis</i>	H	2	m	m	HC	ov	3		12
<i>scutellans</i> G. H. Cunn. <i>Peniophora</i>	hIV	1	u	d	N	c	3-4		8
<i>setulosa</i> (Berk. & Curt.) Nakas. <i>Phlebia</i>	hII					va	2		29
<i>simile</i> Pouz. & Hol. <i>Botryobasidium</i>	H		d	d		a			22
<i>sinuosum</i> Freem. <i>Gloeocystidiellum</i>	hIV								14
<i>sonorae</i> Nakas. & Galz. <i>Cristina</i>	hII					c	6-7		29
<i>spathulata</i> Léger <i>Hymenochaete</i>		1		d*	a	7			27
<i>stereoides</i> Wu <i>Phanerochaete</i>		2	m	m	HC	o	2		38
<i>subasperispora</i> (Litsch.) Jül. <i>Boidinia</i>	h	1	u	d	N	c	5		12
<i>subcoronatum</i> (Höhn. & Litsch.) Donk <i>Botryobasidium</i>	hIV								22
<i>subcretacea</i> (Litsch.) Christ <i>Phlebia</i>	h	1	u	d	N	c	7		16
<i>subglobispora</i> Hall & Hjortst. <i>Vuilleminia</i>	hIV								19
<i>subsalmonea</i> Boid., Lanq. & Gilles <i>Peniophora</i>	h	1	u	d	N	c	3		8
<i>tamariciphila</i> Boid., Lanq. & Gilles <i>Phanerochaete</i>	H	2	p	p	HC	a	3-4		9
<i>tinctorium</i> (Ell. & Ev.) Ell. & Ev. <i>Echinodontium</i>	hIV	1	u	d	N	c			37
<i>vagum</i> (Berk. & Curt.) Rogers <i>Botryobasidium</i>	H		d	d					22
<i>variosum</i> Boid., Lanq. & Gilles <i>Hyphoderma</i>	H	1	u.d	d	N	c	2	Col	8
<i>vesiculosum</i> (Burt) ² Boid., Lanq. & Gilles <i>Gloeocystidiellum</i>	hIV								17
<i>vile</i> (Bourd. & Galz.) Frass. <i>Repetobasidium</i>	H		d	d	HD	c	7		16
<i>vinosa</i> (Overh.) Nakas. <i>Phlebia</i>	hII					va	2-3		29

2 - appele *Gloeocystidiellum clavigerum* en Amérique du Nord, mais incompatible avec cette espèce Européenne.

<i>makulium</i> (Burds. <i>et al.</i>) W.J. <i>Stereum</i>	H	2	p	p	HC	ov	3-4		12-39
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A ce résumé des données publiées, nous ajouterons, ci après, quelques données obtenues à Lyon et non publiées à ce jour.

<i>africana</i> Boid. <i>Candelabrochaete</i>		1		m	HC	a	5-6		
<i>argillacea</i> Bres. <i>Jaapia</i>	hIV	2	u	d	SN	c	3		
<i>armeniaceum</i> Boid. & Gilles <i>Stereum</i>	H	2	m	m	HC	v	3		
<i>ayresii</i> (Berk.) Boid. & Gilles <i>Hyphoderma</i>	hIV	1	u	d	N	c	3		
<i>cremo-album</i> (Hohn. & Litsch.) Jul. <i>Hyphoderma</i>	hII	1	u	d	N	c	7		
<i>erikssoni</i> Oberw. <i>Repetobasidium</i>	hII	1	u	d	N	c	7		
<i>expallens</i> (Bres.) Domanski <i>Dentocorticium</i>	hIV	1	u	d	N	c	4		
<i>gemmiferum</i> (Bourd. & Galz.) Erikss. & Ryv. <i>Coronicium</i>	hIV	1	u	d	N	c	3		
<i>heimii</i> Malençon <i>Veluticeps</i>	h	2	p	d	He	c	7		
<i>incrustatissimum</i> Boid. & G. <i>Hyphoderma</i>	hII	1	u	d	N	c	4-5		
<i>laetum</i> (Karst.) Erikss. & Hjortst. <i>Ervthricium</i>	H	1	p	p	HC	a	7		
<i>melzeri</i> (Pouzar) ³ Stalpers <i>Auriculariopsis</i>	H	2	d	d	HD	c	6		
<i>pseudotsugae</i> (Burt) Boid. & Gilles <i>Aphanobasidium</i>	hIV	1	u	d	N	c	7		
<i>scaevolae</i> Boid. & Gilles <i>Hyphoderma</i>	h	1	u	d	N	c	3		
<i>septocystidia</i> (Burt) Burds. <i>Candelabrochaete</i>		1		p	HC	a	7		
<i>subcrinate</i> (Pock.) Ryv. <i>Meccherinum</i>			p	d*	He	a	7		
<i>transiens</i> (Bres.) Parm. <i>Hyphoderma</i>	hII	1	u	d	N	c	3		

³ L'homothalle constatée en France contraste avec la tetrapolarité d'une récolte Américaine (Nakasono, 1990)

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THERMOPHILIC FUNGI: BIODIVERSITY and TAXONOMIC STATUS.

Jean MOUCHACCA

Laboratoire de Cryptogamie, Museum National d'Histoire Naturelle,
12, rue Buffon, 75005 Paris, France
e-mail : mouch@mnhn.fr

RÉSUMÉ : Une évaluation critique du statut nomenclatural et, dans certains cas, également du statut taxonomique, a été entreprise pour les champignons thermophiles décrits à ce jour. La distinction entre éléments thermophiles et thermotolerants se base sur les définitions élaborées par Cooney & Emerson. Au total, près de quarante espèces et variétés s'avèrent aptes à réaliser un développement optimal à des températures élevées. Des recherches taxonomiques complémentaires sont toutefois nécessaires pour résoudre les problèmes résiduels : la résultante serait une légère réduction de l'effectif du groupe.

Une croissance optimale à des niveaux thermiques situés au-delà du seuil maximal des espèces mésophiles individualise quelques Mucorales, Eurotiales et Sphaeriales, un nombre limité d'Hyphomycètes et un seul Agonomycète. Aucun Coelomycète ni aucun Basidiomycète n'exprime cette particularité écologique. Les Mucorales recensées sont des éléments du genre *Rhizomucor* et de l'élément générique non valide *Thermomucor*. *Rhizomucor pusillus* (espèce-type) et *Rh. tatei* sont des taxons valides ; *Rh. pak stanskyi* se révèle un synonyme ultérieur de l'espèce-type. La validité taxonomique des *Rhizomucor tauricus* et *Rh. namtalenensis* reste à confirmer.

Parmi les vingt ascomycètes répertoriés, *Chaetomyces thermophilus* et *Chaetomium mesopotamicum* sont des taxons bien définis. Ceci n'est pas le cas des *Chaetomium britannicum* et *C. virginicum*, à statut taxonomique encore imprécis ; il en est de même pour leurs liens respectifs avec *Chaetomium thermophilum* et ses deux variétés. *Dactyliomyces thermophilus* est retenu comme seul élément du genre *Dactyliomyces*, celui-ci n'est plus considéré comme congeneric de *Thermomyces*, pour ce dernier, seule l'espèce-type et une variété sont admises. Le nouveau genre *Coomeremia* est proposé pour rassembler les autres espèces des deux entités précédentes. *Coomeremia crustacea* (= *Dactyliomyces crustacea*) est sélectionnée comme espèce-type ; *C. aegyptiaca* (= *Thermomyces aegyptiacus*) et *C. verrucosus* (= *Th. crustaceus* var. *verrucosus*) et *C. latunguicus* y sont également rattachés. Ces trois genres ont comme particularité commune, les caractères de leurs téléomorphes. Cependant, *Dactyliomyces* révèle une forme imparfaite du genre *Polypaecium*. *Coomeremia* des structures conidiogènes de type *Paeclomyces* alors que les *Thermomyces* ne produisent d'anamorphes à conidies en chaînes.

Cornusculus heterothallicus et *C. thermophilus* sont des ascomycètes à perithecies clos ayant chacun une forme imparfaite distincte, respectivement *Mycelophthora thermophila* et *M. fergusii* en raison du caractère hétérothallique des téléomorphes, les anamorphes peuvent être isolés séparément lors des recherches portant, par exemple, sur des matériaux subissant un processus d'auto-échauffement. *Metamycarpus aibomycetes* développe une forme conidienne arthrosporee dont est dépourvu *M. thermophilus* (= *Thielavia minima* var. *thermophila*). Les trois *Talaromyces* thermophiles sont associés à des anamorphes de type *Paeclomyces* (*T. bissochlamydoides*) ou *Penicillium*.

T. marsonii et *T. thermophilus*. Le genre *Thielavia* recèle également trois thermophiles. *T. terrestris*, espèce type, est un ascomycète cosmopolite dont la forme imparfaite *Acremonium alabamense* se rencontre souvent en l'absence du téléomorphe. *Th. australiensis* pour lequel peu d'informations sont disponibles et le récent *Th. pingtungia* dont certains caractères suggèrent son appartenance au genre *Chaetomidium*.

Le groupe des hyphomycètes rassemble treize espèces. Cependant, *Acremonium alabamense* et deux *Mucella pilularia* sont des anamorphes d'ascomycètes plus ou moins hétérothalliques, ils peuvent par conséquent se développer sans les formes parfaites correspondantes. *Mucellaphthora indica* se révèle un synonyme de *M. thermophila*. Les taxons macedoniens restants sont *Acremonium thermophilum*, seul autre élément thermophile du genre. *Mucellaphthora himalaica* forme parfaite encore inconnue et *Mahranthelia chrysomela*, unique thermophile du genre *Mahranthelia* dont les mesophiles sont associées à des formes parfaites connues. *Thermomyces sp. a tibungera* s'individualise par ses cloisons mycéliennes pourvues de hâcles et la présence d'une forme comédienne de type aleurospore.

Les hyphomycètes dematiés relèvent des genres *Humicola*, *Scytalidium* et *Thermomyces*. Ce dernier se révèle être une entité générique assez homogène et son espèce type représente le premier thermophile avéré. *Humicola grisea* var. *indica* et *H. himalaica* var. *caeruleata* sont proposées comme synonymes additionnels à l'espèce type. Le genre comporte également *Thermomyces ibadensis*, *Th. stellatus* et le mesophile *Th. verrucosus*.

Le statut taxonomique de *Humicola hyalothermophila* se doit d'être reconsidéré, en même temps que les *Humicola grisea* var. *thermophila* et *H. insolens*, récemment placés en synonymes avec *Scytalidium thermophilum*, base sur *Tortula thermophila*. Ces deux *Humicola* sont retenus dans l'immédiat dans le complexe *Scytalidium thermophilum*, dans l'attente d'une redefinition des statuts taxonomiques respectifs. *Scytalidium thermophilum* se situe du concept générique de *Scytalidium* fondé sur l'espèce type *S. nigricans*, ce qui n'est pas le cas pour *S. indonesicum*. *Scytalidium alabastrum* est avéré correspondre au *S. thermophilum* sensu lato. *Humicola nigrescens* var. *thermophila* est un synonyme interneur de *H. grisea* var. *thermophila* alors que *H. fuscoatra* var. *nigra* est identique au *H. insolens*.

Deux taxons révèlent un statut incertain. *Mucor thermohydrosphaera* (*Rhizomucor pastus* ?) et *Strobilactia thermophila* en quête d'un genre plus approprié. D'autre part, divers taxons se sont vu attribuer des épithètes spécifiques pouvant conduire à confusion au regard des aptitudes thermophiles respectives. Les cas les plus simples concernent des champignons dénommés *thermophilum* ou de ses variantes et qui sont loin de représenter des thermophiles sur la base des définitions établies. Exemples : l'ascomycète *Achaetomium thermophilum* (un synonyme ultérieur de *A. macrosporum*, une espèce thermotolerante), les hyphomycètes *Ciliatosporella thermophila*, *Gloianella thermophila* et *Zalerion thermophila* (le statut taxonomique des deux derniers reste à considérer), l'ascomycète *Euglenidium thermophilum*, le zygomycète *Mucor thermophilus* et enfin la levure *Endoblastomyces thermophilus* a statut taxonomique non valant. *Macanomyces thermophila* est un basidiomycète simplement observé dans une localité chaude et humide, des cas similaires ont été également recensés.

Le binôme *Sporotrichum thermophilum* est un exemple d'une source différente de confusion. Ce taxon ne possède de statut taxonomique aucune sorte. Il est cependant fréquemment signalé comme thermophile dans la littérature portant sur les études enzymiques des champignons. Ces binômes fantômes sont relativement fréquents dans ce type de publications. Cette pratique préjudiciable devrait être définitivement abandonnée.

ABSTRACT. A critical reappraisal of nomenclature, status and in some cases also of taxonomic ones was undertaken for known thermophilic fungi. Distinction between thermophilic and thermotolerant follows definitions elaborated by Cooney & Emerson. Altogether less than forty species and varieties are able to achieve best development at high temperatures. Further taxonomic work is however needed to solve residual problems. The outcome will be a slight reduction of the group.

Optimum growth at temperatures above the maximum threshold of mesophiles characterise few Mucorales, Eurotiales and Sphaeriales, a limited number of Hyphomycetes plus one Agonomycete.

cete species. No Coelomycete and no Basidiomycete develop such ecological feature. Recorded *Macrosporales* are species of *Rhizomucor* and the invalid monospecific genus *Thermomucor*. *Rhizomucor* *fishii* is (type species) and *Rh. michei* are valid taxa. *Rh. pakistanicus* proved to be a later name for the former. The validity of *Rhizomucor tauricus* and *Rh. nantlachi* awaits confirmation.

Among the twenty ascomycetes, *Camarosporium thermophilum* and *Chaetomium mesopitatum* are well defined taxa. But the status of *Chaetomium brunnescens* and *C. virgatum* is still unclear. Also relations with *Chaetomium* and its two varieties awaits clarification. *Dactylomyces thermophilus* is retained as the sole species of the genus, no longer regarded as congeneric with *Tarmonia*, for the latter only the type and one variety are accepted. A new genus *Coomenia* is proposed to accommodate remaining taxa of both genera. *Coomenia cristata* (= *Dactylomyces cristatus*) is selected as type species. *C. aegyptiaca*, *Thermomyces aegyptiacus* and *C. verrucosus* (= *Th. chaetacthus* var. *verrucosus*) and *Th. taeniatus* are two other members of this genus. These three genera have in common the characters of their teleomorphs. But *Dactylomyces* has a *Pezizacanthus* type anamorph. *Coomenia* develop conidogenous structures of the *Pezizomyces* type while *Thermomyces* has no anamorph producing chains of conidia.

Crematosphaeria and *Cremophilus* are cleistothecial ascomycetes having a well defined anamorphic state, namely *Mycetophthora thermophila* and *M. fusigera* as the teleomorphs are heterothallic. The anamorphs could be observed a one in states involving for instance self fertilized material. *Mycetophthora albomyces* has a well defined arthroconidial state not developed by *M. thermophila* (= *Thickia minima* var. *thermophila*). The three thermophilic *Lobomyces* have conical states belonging either to *Pezizomyces* (= *Pezizochlamys hirsuta*) or to *Pezizacanthus* (*Lobomyces* and *T. thermophilus*). *Thickia* (also three species) is represented by the widely distributed *Th. terrestris* whose anamorph *Acremonium* and *monose* could equally be isolated separately. The undocumented *Th. australensis* and the recent *Th. pinguicula* with features favouring his relocation in *Chaetomium*.

Recorded hyphomycetes comprise thirteen taxa. But *Acremonium adamsense* and two *Mycetophthora* are anamorphs of a most heterothallic ascomycetes, these could thus develop at high temperatures without producing respective ascocarps. *Mycetophthora indica* is considered a synonym of *M. thermophila*. Other heterothallic taxa are *Acremonium thermophilum*, the second thermophilic of the genus. *Mycetophthora annulata* perfect state is yet unknown and *Malbranchea annulata* is a thermophile of a genus whose mesophilic components are associated with perfect states. *Tricophaema asporibidigera* is unique by its hyphae disclosing septal clamp connections and the formation of an aleuriosporic state.

Dematiaceous thermophiles are members of *Humicola*, *Sclerotium* and *Thermomyces*. The latter is regarded as a homogeneous genus with the type species *Thermomyces lanuginosus* being the first established thermophile. *Humicola grisea* var. *maria* and *H. lamagnyana* var. *celata* are additional later names for the type species. Other known members are *Thermomyces thadensis*, *Th. stellatus* and the mesophilic *Th. verrucosus*.

The taxonomic status of *Humicola halothermophila* awaits to be reconsidered together with *Humicola grisea* var. *thermophila* and *H. insolens*, recently placed in synonymy with *Sclerotium thermophilum*, based on *T. nuda* thermophila. Both *Humicola* are placed here for the moment as synonyms of the complex *S. thermophilum*, pending reassessment. *Sclerotium thermophilum* deviates from the current concept of *Sclerotium* (based on *S. lignicola*) but such is not the case for *S. adamsense*. *Sclerotium allahabadicum* proved to match *S. thermophilum sensu lato*. *Humicola nigrescens* var. *thermophilus* duplicates the description of *Humicola grisea* var. *thermophila* while *H. fuscoatra* var. *nigra* is identical to *H. insolens*.

Two taxa have an uncertain position. *Macrothermomyces* (*Rhizomucor pusillus*) and *Stilbella thermophila* requiring a more appropriate genus. Also several taxa disclose confusing specific epithets with regard to thermophilic abilities. Simple cases refer to fungi with epithets as *thermophilum* or variants of, and that are not thermophilic based on accepted definitions. Examples, the ascomycete *Chaetomium thermophilum* (a later synonym of *T. rosarium*, a thermotolerant), the hyphomycetes *Calcarisporium thermophila*, *Gibberella thermophila* and *Zalerion thermophilum* (the latter two have yet unsettled status), the oomycete *Lagenidium thermophilum*, the zygomycete *Macro-*

thermophilus and finally the invalid yeast *Enkblastomyces thermophilus*. *Melanophthalma thermophila* is a basidiomycete simply observed in a warm humid locality: similar cases could also be traced.

The binomial *Sporodionum cellulosiphum* is an example of a different confusing situation. The taxon has no taxonomic status of any type although being infrequently reported as a thermophile in literature related to fungal enzymic studies. Such ghost binomials are not uncommon in these publications. This practice being a source of serious confusion should be definitely prohibited.

INTRODUCTION

Temperature is one of the extremely important environmental variables that play a decisive role in the survival, growth, distribution and diversity of microorganisms on the surface of the earth. The response of fungi to temperature varies between the two extremes of obligatorily thermophilic through thermotolerance to psychrophilic species. However, by far the majority of known fungi are mesophiles developing in culture between 5 and 37 °C; the psychrophiles extend below that range of temperatures (Dix & Webster, 1995).

Thermophily has been defined variously with reference to different groups of microorganisms and sometimes also within the same group. The response of fungi to high temperatures has been the subject of classificatory schemes successively proposed by Apinis (1963), Cooney & Emerson (1964), Craveri *et al.* (1964), Evans (1971) and Crisan (1973). These schemes are either based on values of minimum and maximum growth temperatures alone or, in addition, integrate the criteria of optimum development.

The commonly accepted definitions of thermophilic and thermotolerant fungi are those of Cooney & Emerson (1964). Thermophilic fungi are those that have a growth temperature maximum at 50 °C or above and a temperature minimum of 20 °C or higher. Thermotolerant species are those that have a growth temperature maximum of about 50 °C and a temperature minimum well below 20 °C. This simple segregative scheme is sometimes difficult to apply since the response of thermophilic taxa at the minimum temperature threshold tends to vary among respective strains.

Serious consideration of fungi able to develop only at high temperatures dates back to 1899 when P. Tsiakinsky first reported on a thermophilic hyphomycete incidently encountered on a potato inoculated with garden soil. The fungus was then grown on bread kept at 52–53 °C and its thermophilic nature assessed. Tsiakinsky named this hyphomycete *Thermomyces lamagnus*. Rapidly, however, this thermophile was successively relocated in other genera as *Acremonium*, *Humicola*, *Monospora* and *Scedonum* before its definite reinstallation in *Thermomyces*. Similar changes also characterize thermophilic moulds described in the early decades of the nineties. The outcome of such changes is the chaotic nomenclatural state of few members of this group in published literature. Absence of homogeneity in binomial citations develop cases of taxonomic confusion coupled with divergence in species concept (Cooney & Emerson, 1964). The final result is a partial or total incomplete identification of encountered taxa or names reported being shadowed.

Although Lindt description of *Rhizomucor pusillus* (as *Mucor pusillus*) dates back to 1886, there is a general agreement that Tsiakinsky (1899) is the first to draw attention to thermophilism among fungi. Very rapidly, Mische's (1905) serious investigation of self heating hay produced the first extensive report on thermophily in fungi. This author isolated and studied a range of thermophiles including *Thermosaurus aurantiacus* and *Malbranchea cinnamomea* (Mische, 1907). Griffon & Maublanc (1911) then introduced the first thermophilic *Penicillium*, *P. daponii*, now *Talaromyces thermophilus*. It is only several decades later that La Touche (1950) reported on the new *Chaetomium thermophile*. Such discovery generated much interest to this group of fungi, substantiated by the cellulolytic nature of the new ascomycete.

Several pioneer publications then followed on thermophilic fungi inhabiting soils of temperate regions (Apinis, 1963; Eggins and Malik, 1969), tropical areas (Hedger 1974; Gochenaur, 1975) and on soils of arid regions (see review in Mouchacca, 1995). Thermophilic fungi of habitats rich in organic materials were also extensively surveyed and data from relevant publications critically reviewed by Tansey & Brock (1978). Reports on less widespread habitats and habitats deserving future investigations were also considered by Tansey & Brock (1978).

The first modern comprehensive account on the taxonomy, biology and economic importance of thermophilic fungi was published by Cooney & Emerson (1964). Eleven thermophiles were reported. Since then the number of taxa developing at high temperatures is expanding rapidly. In 1973, Crisan provided a list of 55 names of thermophilous fungi, i.e. thermophilic and thermotolerant ones, however, only half are thermophiles in the sense of Cooney & Emerson. Crisan reviewed in addition current concepts about thermophilism in microorganisms, he then underlined that our knowledge about the physiological ability of fungi to grow at elevated temperatures was much limited. Later Samson & Tansey (1977) prepared a guide to species able to grow and sporulate at 45 °C, this list concerns eight macorales, around twenty taxa each of ascomycetes and hyphomycetes and two basidiomycetes. The subsequent list prepared by Tansey and Brock (1978) reports 67 species or varieties growing at 50 °C or above, a good proportion of these taxa was however not specified at the species level. A Russian compilation of descriptions and published illustrations of thermophilic fungi was prepared by Bilal & Zakharchenko (1987), 38 species were considered but few are not strict thermophiles. Finally, according to Abdullah & Al Bader (1990), around 70 species detected in various substrates are now reported to be thermophilic or thermotolerant.

Cooney & Emerson (1964) monograph introduced new thermophilic taxa. However, some taxonomic decisions they adopted rapidly proved to be misleading and their descriptions of novel taxa supported not critical analysis. These limitations triggered subsequent studies. Several interesting notes thus appeared in the sixties clarifying pending problems while expanding the group. Apinis & Chester (1964) described *Dactylomyces crustaceus*. Pugh *et al.* (1964) reintroduced *Thermomyces*. Stolk (1965) assessed the taxonomic status of *Penicillium dipontii* and *Thermoascus aurantiacus*. Again Apinis (1967) clarified generic concepts of *Dactylomyces* and *Thermoascus*. However, inspite of the above and later contributions, not all standing problems received attention. Recently, Straatsma and Samson (1993) focused on both *Humicola* proposed by Cooney & Emerson (1964).

The material of this contribution was collected while preparing a lecture for the Microbial Diversity and Ecosystem Function workshop held at Egham, UK, in 1993. The lecture focused on thermophilic fungi of desert soils, an example of a neglected extreme environment (Mouchacca, 1995). A second lecture on the subject was later presented at IMC V, this was entitled "Thermophilic and thermotolerant fungi in the Middle East: Biodiversity and Taxonomic Reappraisal" (Mouchacca, 1994), however, the corresponding note suffered some publication delay. In the meantime, the first draft of the present paper was due to be part of a book in the pipeline somewhere in the Indian subcontinent. Decision was then taken to update and publish the applied last version.

The present document aims to provide a sound reappraisal of the nomenclatural and in some cases of the taxonomic status of known thermophiles. First to overcome a major difficulty encountered while interpreting published data on this ecological group. Second to enhance future taxonomic work on its components and, finally, to stress the attention on taxa other than those commonly studied for eventual industrial applications.

THERMOPHILIC MUCORALES

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basionym: *Mucor pusillus* Lindt — *Archiv für experimentelle Pathologie und Pharmakologie* 21: 272. 1886.

= (?) *Mucor septatus* Bezold in Siebenmann — *Die Schimmelmikosen des menschlichen Ohres*: 97. 1889.

= *Rhizomucor septatus* (Bezold) Lucet & Cost. — *Archives de Parasitologie* 4: 362. 1901.

= *Mucor* (sect. *Rhizomucor*) *parasiticus* Lucet & Cost. — *Comptes rendus hebdomadaires des Séances de l'Académie des Sciences, Paris* 129: 1033. 1899.

= *Rhizopus parasiticus* (Lucet & Cost.) Lendner — *Matériaux pour la Flore Cryptogamique Suisse* 3: 115. 1908.

= (?) *Mucor muriperda* Sacc. & Sinigaglia in Sacc. — *Sydow, Annales Mycologici, Ser. II*, 11: 321. 1913.

= *Tieghemella muriperda* (Sacc. & Sinigaglia) Naumov — *Opredelitel' Mukorovykh (Mucorales)*: 84. 1935.

= *Mucor buntingii* Lendner — *Bulletin de la Société botanique de Genève* 21: 260. 1930.

= *Mucor hagemii* Naumov — *Opredelitel' Mukorovykh (Mucorales)*: 55. 1935 (*nom. inval.*, Art. 36.1).

= *Rhizomucor pakistanicus* Qureshi & Mirza — In Mirza, Khan Begum & Shagufta "Mucorales of Pakistan (Faisalabad)": 100. 1979 (*nom. inval.*, Art. 37.1); Qureshi & Mirza — *Biologia, Lahore* 29: 343. 1983, a superfluous publication.

Descriptions: Cooney & Emerson (1964); Schipper (1978); Domsch *et al.* (1980).

This is the type species of *Rhizomucor* (Lucet & Cost.) Wehmer: Vuill. The genus was reintroduced by Schipper (1978) to segregate three hitherto known thermophilic species of *Mucor* distinguished by the presence of rhizoids at the base of their sporangio-phores.

The early history of *Rhizomucor pusillus* (as *Mucor pusillus*) and its long confusion with the *Mucor* species studied by Miehe (Miehe, 1907; now *Rhizomucor miehei*) was reviewed by Cooney & Emerson (1964). The former had indeed often been misidentified with the equally thermophilic, thinly growing and equally common *Rhizomucor miehei*. This zygomycete is however regularly homothallic while in *Rhizomucor pusillus* homothallic isolates have only exceptionally been found. An excellent account of the morphology, physiological characteristics and distribution is provided by Domsch *et al.* (1980).

Rhizomucor pakistanicus was isolated from several sources in Pakistan: groundnut seeds and lizard droppings collected at the city of Faisalabad, from a potato field at Sialkot and from soil at Rawalpindi (Mirza *et al.*, 1979). It was then correctly assigned to *Rhizomucor* (indicated as *Rhizomucor* Lucet & Cost.) on account of the presence of rhizoids. However no comparison was undertaken with the indicated type species, *Rhizomucor parasiticus* Lucet & Cost. Further, in the second superfluous publication made by the same authors (simply a duplicate of the former), taxonomic decisions concerning this genus introduced by Schipper (1979) were overlooked. For *Rhizomucor pakistanicus*, a number of morphological features forwarded in its description leads to consider the fungus as a synonym of *Rh. pusillus*.

The current concept of *Rhizomucor* was however recently expanded to make provision for mesophilic isolates also producing rhizoids from the base of their sporangiophores. *Rhizomucor variabilis* Zheng & G.-q. Chen var. *variabilis* was obtained from a cutaneous mucormycosis of a human hand in China (Zheng & Chen, 1991). *Rhizomucor variabilis* var. *regularior* Zheng & G.-q. Chen represent another agent of cutaneous human disease (Zheng & Chen, 1993). Optimum, maximum and minimum growth temperatures of the two varieties are the same, i. e. 24-30° C, 37° C and 9° C respectively. The additional mesophilic *Rhizomucor endophyticus* Zheng & H. Jiang (1995) was isolated from wheat collected in China; its optimum, maximum and minimum growth temperatures are even lower, being 18-28° C, 36° C and 5° C respectively.

— *Rhizomucor miehei* (Cooney & Emerson) Schipper — *Studies in Mycology* 17: 58. 1978.

basionym: *Mucor miehei* Cooney & Emerson — *Thermophilic Fungi*: 26. 1964.

Descriptions: Cooney & Emerson (1964); Schipper (1978).

As stressed before, isolates of this zygomycete regularly produces zygospores in cultures. This finding led Cooney & Emerson (1964) to propose a specific rank for this taxon previously considered as identical to the almost morphologically similar type species. Schipper (1978) stressed however that the general morphology could also be used to distinguish both taxa. *Rhizomucor miehei* exhibits a looser sympodial branching pattern with relatively longer side branches while *Rh. pusillus* produces small bunches of short subterminal branches on the main sporangiophores. In addition the sporangia and columellae of the latter are usually larger.

Rhizomucor miehei displays a wide geographic distribution (Domsch *et al.*, 1980). Factors affecting development of zygospores were investigated by Lasure & Ingle (1976) and those regulating germination of sporangiospores by Deploey (1992).

— *Rhizomucor nainitalensis* Joshi — *Sydowia* 35: 100. 1982.

This still Indian based homothallic zygomycete was isolated from a heavily decomposed oak log in the forest of Pungote, Nainital. It differs from *Rhizomucor miehei*, *Rh. pusillus* and *Rh. tauricus* mainly by sporangiospores of varying shapes and sizes: subglobose, ellipsoidal, oblong, reniform, dump-bell shaped, etc., 3-6 µm or more wide. Variation in sporangiospore shape might however be an artefact.

According to Joshi (1982), growth is very rapid at 48° C filling half of a Petri dish in 2 d. At 38° C, "the growth of the mycelium takes place after three days but about one week is required to colonize the culture medium in a petri dish at 25° C". *Rhizomucor nainitalensis* appears very close to *Rh. miehei*.

— *Rhizomucor tauricus* (Milko & Schkurenko) Schipper — *Studies in Mycology* 17: 62. 1978.

basionym: *Mucor tauricus* Milko & Schkurenko — *Novosti sistematiki nizshikh rastenii* 7: 139. 1970.

Description: Schippers (1978).

The fungus is apparently still known only from the original strain isolated from forest soil in the Ukraine. It was maintained as a separate species by Schipper (1978)

pending further informations. *Rhizomucor tauricus* is distinguished from other thermophilic *Rhizomucor* mainly by definitely more swollen sporangiophores. Growth and sporulation occurs between 24-55° C, development is extremely slow at 21° C and nil at 57° C

***Thermomucor indicae-seudaticae* Subrahmanyam, Mehrotra & Thirumalachar (as "Subrahmanyam,...") — Georgia Journal of Botany 35: 2. 1977. (nom. inval., Art. 37.1).**

Descriptions Subrahmanyam, Mehrotra & Thirumalachar (1977), Schipper (1979)

This is the type species of *Thermomucor* Subrahmanyam *et al.* (1977) which is apparently still monospecific. It was established mainly on account of formation by the type species of smooth zygospores and presence of rhizoids. Zygospores are definitely rough-walled in members of *Absidia* van Tieghem, *Mucor* Mich. Fr., *Rhizopus* Ehrenb. Corda and *Rhizomucor*.

The described strain was isolated from municipal compost at Pimpri, Poona, India. Prior to its description, this zygomycete was reported as *Rhizopus* sp. and as such isolated from various habitats in Europe, India, Ghana and Nigeria (Schipper, 1979).

THERMOPHILIC ASCOMYCETES

***Canariomyces thermophila* Guarro & Samson in von Arx, Figueras & Guarro — Beihefte zur Nova Hedwigia 94: 34. 1988.**

Canariomyces von Arx (von Arx, 1984) was established for a mesophilic cleistothecial ascomycete, *C. notabilis* von Arx, having ascoma wall made up of angular dark cells, irregularly disposed asci, aseptate ascospores dextrinoid when young and provided with a single germ pore, an anamorph having conidia of the form genus *Chrysosporium* von Arx (catenate aseptate or septate hyaline conidia) and of *Trichosporiella* Kamyschko (simple lateral conidia) is produced.

Canariomyces thermophila was isolated from Cameroon, Africa, apparently from soil. The original protologue is based on colonies developing at 45° C but no data about minimum and maximum growth temperatures are provided. Ascospores are greenish brown when mature with a subapical germ pore. 14.0-18.0 × 7.5-10.0 µm. However no anamorph developed in cultures of the single available strain.

***Chaetomium britannicum* Ames — A Monograph of the Chaetomiaceae: 16. 1963.**

Descriptions: Ames (1963); von Arx *et al.* (1986).

This *Chaetomium* was described as having ovoid to vase-shaped ascomata. Terminal and lateral hairs are very slender, greyish, straight to undulate. Asci club shaped, 8-spored. Ascospores brown, large, 19-24 × 11-14 µm, irregularly oval, rounded on the ends. In the protologue, it is simply indicated that "perithecia develop when incubated at approximately 47° C. A thermophile" (Ames, 1963). The specific epithet refers to the type locality: southern part of England.

The original material developed on mushroom compost and apparently no living culture was realised. von Arx *et al.* (1986) regard this *Chaetomium* as a doubtful species, only ascospores could be observed in the type specimen at BPI. Millner (1977) attempted but without success to obtain a culture from the dried type material, as no living strain, authentic or representative, was available to him, no growth temperature relationships could thus be established.

Gochenaux (1975) reported having isolated this *Chaetomium* from soil in the Bahamas but Millner *et al.* (1977) stressed the absence of a corresponding culture or herbarium specimen. Further and based on informations communicated by Gochenaux, the micromorphology of the fungus she examined was probably not *Chaetomium britannicum*. Spores of Gochenaux's taxon measured $13 (18) \times 7 (10) \mu\text{m}$ and had subapical germ pores while spores from Ames material measure $19.0-24.8 \times 11-14 \mu\text{m}$ and have single apical germ pores (Millner *et al.*, 1977). *Chaetomium britannicum* remains undocumented in the sense of Cooney and Emerson. Also no additional record either from the type locality (Cannon *et al.*, 1985) or elsewhere has apparently been published.

***Chaetomium mesopotamicum* Abdullah & Zora — Cryptogamic Botany 3: 387, 1993.**

The original locality is a date palm plantation in Basrah, Iraq. This recently described species has a growth temperature range from 30-52° C. It differs from *Chaetomium thermophilum* La Touche and *Chaetomium virgicum* Ames by its clavate asci and long highly branched terminal hairs. Ascospores are globose to ovoid, olive to brown, $5.5-7.8 \times 5.2-6.3 \mu\text{m}$, provided with one apical germ pore.

***Chaetomium thermophilum* La Touche as “*thermophile*” — Transactions of the British mycological Society 33: 94, 1950; Cooney & Emerson — Thermophilic Fungi: 62, 1964.**

— ? *Chaetomium virgicum* Ames — A Monograph of the Chaetomiaceae 43, 1963, *file* von Arx *et al.*, 1986

Descriptions La Touche (1950), Ames (1963), Cooney & Emerson (1964), von Arx *et al.* (1986).

This is the first known thermophilic *Chaetomium*. The species is also distinguished by its long, tapering terminal hairs at times dichotomously branched at wide angles. Ascospores are dark brown, globose to subglobose, $7.9 \times 5-7 \mu\text{m}$, prominently umbonate at one end.

Cooney & Emerson (1964) observed this *Chaetomium* to produce in culture two dissimilar growth patterns, as a result they proposed two new varieties: *coprophile* and *dissitum*.

***Chaetomium thermophilum* La Touche var. *coprophile* Cooney & Emerson — Thermophilic Fungi: 69, 1964.**

The variety is mainly distinguished by the presence of dichotomously branched hairs, which more or less completely covers the entire perithecium.

***Chaetomium thermophilum* LaTouche var. *dissitum* Cooney & Emerson
— Thermophilic Fungi: 68. 1964.**

Differs from the species mainly in the diffuse manner in which perithecia are produced in culture.

***Chaetomium virginicum* Ames — A Monograph of the Chaetomiaceae:
43. 1963.**

The fungus was originally isolated from leaf litter collected under very old trees at White Marsh, North of Old Point Comfort, Virginia (U.S.A.). Perithecia are described as globose, up to 240 μ m wide. Terminal and lateral hairs cover the entire perithecium, indistinguishable, irregularly branched, minutely granular, 2-4 μ m wide. Asci cylindrical, 8-spored, 70 \times 10 μ m. Ascospores yellow-brown to pale brown, almond-shaped, 8-11.5 μ m. A thermophile (Ames, 1963).

Cooney & Emerson (1964) consider this species to approximate *Chaetomium thermophilum* while being identical to its variety *coprophile*; ascomata of the latter are also entirely covered by dichotomously branched perithecial hairs. On the basis of morphological characters and growth-temperature responses, Millner (1977) provided evidence that *Chaetomium virginicum* (culture IA-7 obtained from L. M. Ames collection at BPI) is identical to *Ch. thermophilum* var. *coprophile*. This proposal was later substantiated by Millner *et al.* (1977), among the large number of *Chaetomium* studied by these authors, ascospores with papillate germ pores were found only in *Chaetomium thermophilum*, its two varieties and *Ch. virginicum*.

Chaetomium virginicum is also regarded as a probable synonym of *Ch. thermophilum* by von Arx *et al.* (1986).

— *Coonemeria* Mouchacca gen. nov.

Thermophila. Coloniae lanatae aurantiae-rubrae quando ascoma matura sunt. Innum. ascomatis est convoluta hypha. Ascoma non-ostiolata sphaerica, solitaria vel confluenta. Ascomata confluentes in crusta disposita. Ascomatis paries crassus, e cellulis pseudoparenchymaticis compositus, ascomatis textura angulosa. Ascus ex hamo singulato oriundus, ovoides vel pyriformis, octosporus demum evanescens. Ascospora unicellularia ellipsoidea vel ovoida, flavescentes vel pallida aurantio-brunnea, cum pariete crassa laeves vel verrucosae. Structura conidiogena semper praesens, similis Paecilomyces Bamier forma genericum.

Species typica: *Coonemeria crustacea* (Apinis & Chesters) Mouchacca.

Thermophilic. Colonies lanose, reddish-orange due to mature ascomata. Ascomatal initials a coiled hyphae. Ascomata non-ostiolate, spherical, solitary to confluent, forming a crusty layer. Ascomatal wall thick, made up of pseudoparenchymatous cells, *textura angularis*. Asci arise singly from croziers, ovoid to pyriform, 8-spored, evanescent. Ascospores unicellular, ellipsoid to ovoid, yellowish to pale reddish-brown, thick-walled, smooth to verrucose. Conidiogenous structures always present, belong to form genus *Paecilomyces* Bamier.

Type species: *Coonemeria crustacea* (Apinis & Chesters) Mouchacca.

Etymology: genus name coined from the first four respective letters of Cooney & Emerson's names, authors of the first comprehensive (although somewhat confusing) monograph on thermophilic fungi.

Coonemeria is proposed to accommodate thermophilic cleistothecial ascomycetes having coiled ascogonial initials, pseudoparenchymatous walls of *textura angularis* type and a distinctive *Paecilomyces* anamorph. Asexual reproductive structures generally represent comparatively reduced forms of well elaborated *Paecilomyces* conidial structures developed in several taxa of this form genus (Samson, 1974).

The three species accepted in *Coonemeria* were formerly assigned to *Thermoascus* Mische (Mische, 1907) and *Dactylomyces* Sopp (Sopp, 1912). These genera have morphologically similar perfect states. *Thermoascus* is distinguished by the absence of any accessory state producing chains of conidia while this state belongs to *Polypaecilum* G. Smith in the case of *Dactylomyces* (Apinis, 1967). The proposal of *Coonemeria* definitely clarifies the status of ascomycetes formerly placed in one or the other of these entities.

Anamorphs of the *Paecilomyces* type are also associated with *Talaromyces* Benjamin and *Byssochlamys* Westling (Stolk & Samson, 1972). The former is distinguished by soft white to yellow ascomata having walls composed of loose hyphae and the production of asci in chains. Most *Talaromyces* however are associated with *Penicillia* while only two have a *Paecilomyces* state, the latter were placed by Stolk & Samson (1972) in *Talaromyces* section *Emersonii*: the thermotolerant *T. leycettianus* Evans & Stolk and the thermophilic *T. byssochlamydioides* Stolk & Samson, here considered. The two other taxa of the section have *Penicillia*: the thermophilic *T. emersonii* (treated in this note) and the thermotolerant *T. bacillosporus* Benjamin.

On the other hand, all *Byssochlamys* have a conidial *Paecilomyces* state. This teleomorphic genus is characterized by initials consisting of swollen antheridia and coiled ascogonia producing almost naked ascomata in which globose asci are formed from croziers (Stolk & Samson, 1972).

Coonemeria crustacea (Apinis & Chesters) *comb. nov.*

basionym *Dactylomyces crustaceus* Apinis & Chesters — Transactions of the British mycological Society 47: 428, 1964

Thermoascus crustaceus (Apinis & Chesters) Stolk — Antonie van Leeuwenhoek 31: 272, 1965

anamorph *Paecilomyces crustaceus* Apinis & Chesters — Transactions of the British mycological Society 47: 428, 1964

Misapplied names *Thermoascus aurantiacus* Mische — Die Selbsterhitzung des Heues 70, 1907; *sensu* Cooney & Emerson — Thermophilic Fungi: 39, 1964

Dactylomyces thermophilus Sopp — Skrifter udgivne af videnskabs selskabet i Christiania. Matematisk naturvidenskabelig klasse 11: 35, 1912, *sensu* Cooney & Emerson — Thermophilic Fungi: 39, 1964

Penicillium thermophilus (Sopp) Bourge — La Cellule 33: 106, 1923, *sensu* Cooney & Emerson — Thermophilic Fungi: 39, 1964

— *Penicillium thermophilum* (Sopp) Sacc., *vide* Trotter 1931 — Sylloge Fungorum 25 (Suppl. 10): 671, 1931, *sensu* Cooney & Emerson — Thermophilic Fungi: 39, 1964

Descriptions as *Dactylomyces crustaceus* by Apinis & Chesters (1964) and Apinis (1967), as *Thermoascus aurantiacus* by Cooney & Emerson (1964), as *Thermoascus crustaceus* by Stolk (1965), Awao & Otsuka (1974) and Chen & Chen (1996)

In 1964, Apinis & Chesters reported on an ascomycete isolated at 38° C from grass debris collected in a salt marsh on the Lincolnshire coast. *Dactylomyces* Sopp was then thought to be the correct genus. The fungus also proved to compare with the CBS strain 374 62 (= QM 6798 = NRRL 1563) deposited by Raper & Fennell as *Dactylomyces thermophilus*, the genus type species. At that time, Sopp's fungus was only known from the original description and illustrations (Sopp, 1912).

Apinis & Chesters (1964) compared these two *Dactylomyces* strains with the protologue of the type species and noted several discrepancies. Both isolates were found to deviate by the absence of dactyloid conidiophores bearing small conidia and the presence of relatively small asperulate ascospores. For *Dactylomyces thermophilus*, Sopp had reported ascospores as oval, smooth, $10\text{--}12 \times 6\text{--}7\text{ }\mu\text{m}$. Ascospores of the living strains were reddish brown, globose to oval, rough and smaller, $7\text{--}9 \times 5\text{--}7\text{ }\mu\text{m}$. Further, their respective anamorphs were found to perfectly fit in *Paecilomyces* with conidia being hyaline, cylindrical to oval, smooth, $3\text{--}8 \times 2\text{--}4\text{ }\mu\text{m}$. Based on these deviations, Apinis & Chesters proposed the new *Dactylomyces crustaceus* and selected as type material their strain BDUN 378 (= IMI 102470).

Also in 1964, Cooney & Emerson published their monograph on thermophilic fungi. They provided a latin diagnosis for the type species of *Thermosascus*, *Th. aurantiacus*, based on their strain M 206516. They also regarded *Dactylomyces* as congeneric with the previously described *Thermosascus* and thus listed known synonyms of both states of *D. thermophilus* under *Thermosascus aurantiacus* (see also under *Dactylomyces thermophilus*). Before this monograph, the anamorph of *Dactylomyces thermophilus* was regarded as approximating a *Penicillium* (Biourge, 1923).

A year later, Stolk (1965) re-examined strain CBS 374 62 and concurred with Apinis & Chesters (1964) about its similarity with their *Dactylomyces crustaceus*. Stolk admitted however Cooney & Emerson's (1964) disposition for *Dactylomyces* and accordingly proposed the new combination *Thermosascus crustaceus* (Apinis & Chesters) Stolk. Stolk finally compared the above strain with four other CBS cultures labelled *Thermosascus aurantiacus*: CBS 256 34, 257 34, 415 62 & 398 64. No conidial state matching the anamorph Cooney & Emerson (1964) depicted for this ascomycete developed in any of these cultures. Stolk then concluded "Cooney & Emerson's strain M206516 of *Thermosascus aurantiacus* is most likely identical with CBS 374 62", a suggestion being substantiated by the fact that the relevant iconography is suggestive of *Paecilomyces*.

Finally, in 1967, Apinis re-examined strain M 206516. It proved conspecific with *Dactylomyces crustaceus*. Thus he, first, confirmed Stolk's suggestion about this strain and, second, the description provided by Cooney & Emerson (1964) for *Thermosascus aurantiacus* applies in fact to the former. In addition, the study of fresh isolates matching the protologue of *Dactylomyces thermophilus*, lead Apinis (1967) to conclude *Dactylomyces* should be maintained distinct from *Thermosascus*, a decision largely accepted subsequently (Cannon *et al.*, 1985; Eriksson & Hawksworth, 1993). As underlined before, the development of a distinctive *Paecilomyces* anamorph favours the placement of *Dactylomyces crustaceus* in *Coonemeria*.

On common laboratory agar media, the minimum growth temperature lies between 20–25° C with the maximum being below 60° C. Optimum growth is around 40° C with a standard Petri dish being covered in 4 d, mature colonies are colored orange with orange brown reverse.

Initials a simple ascogonial coil. Cleistothecia scattered or confluent and then forming a crusty layer, spherical, 300–700 μm diam., orange to reddish-brown. Ascomatal wall made of few layers of brown pseudoparenchyma cells with slightly thickened walls.

Asci are produced singly from croziers, irregularly disposed in the ascomatal cavity, 8-spored, subglobose to pyriform, $15\text{--}20 \times 13\text{--}16\text{ }\mu\text{m}$, evanescent. Ascospores oval, pale brown to red-brown, $6\text{--}8\text{--}0 \times 5\text{--}6\text{--}5\text{ }\mu\text{m}$, wall $0\text{--}5\text{ }\mu\text{m}$ thick, provided with fine echinulations.

Asexual reproductive structures develop within 2 d at 40°C , they are evanescent and not affecting overall colony characters. Conidiophores coarse, septate, pale yellow, smooth, up to $900\text{ }\mu\text{m}$ long, tapering to $4\text{--}5\text{ }\mu\text{m}$ wide apical parts. Upper parts of the conidiophores bears irregularly arranged branches, $6\text{--}35 \times 4\text{--}5\text{ }\mu\text{m}$, these are usually rebranched and end with phialides, phialides occur either singly as side branches, or in irregular verticils of 2–3, phialides cylindric, $12\text{--}30 \times 5\text{--}7\text{ }\mu\text{m}$, gradually tapering to a long conidium-bearing tube, slightly bent away, $12 \times 3\text{ }\mu\text{m}$. Conidia produced in conspicuous long diverging chains, smooth, yellow to pale brown, cylindrical when young, ellipsoid when mature, $6\text{--}10 \times 3\text{--}6\text{ }\mu\text{m}$, responsible for the slight ochraceous color of young colonies.

Coonemeria crustacea is distinguished from the two other members of the genus mainly by oval finely echinulated ascospores. It displays a wide geographic distribution being isolated from soil in several localities and from various self-heating material (Cooney & Emerson, 1964; Cannon *et al.*, 1985; Chen & Chen, 1996).

Coonemeria aegyptiaca* (Ueda & Udagawa) *comb. nov.

basionym *Thermascus aegyptiacus* Ueda & Udagawa Transactions of the Mycological Society of Japan 24: 135, 1983.

anamorph *Paeecilomyces aegyptiaca* Ueda & Udagawa Transactions of the Mycological Society of Japan 24: 135, 1983

The fungus was originally isolated from a sample of marine sludge collected along the Suez Canal banks at Port-Said City, Egypt. It develops between 25 to 55°C with the optimum being at 40°C . At this temperature, colonies fill the plate within 4 d with a thin almost submerged basal mycelium producing numerous superficial ascocarps, often forming a crusty mass, vinaceous to reddish brown, conidia fairly abundant, grayish yellow and not affecting colony color.

Cleistothecia superficial, subglobose, orange-brown, $250\text{--}550\text{ }\mu\text{m}$ wide, initials a simple coiled hyphae. Peridium $25\text{--}40\text{ }\mu\text{m}$ thick, pseudoparenchymateous, rather coriaceous, *textura angularis* type. Asci borne singly on croziers, scattered in the ascomatal cavity, 8-spored, ovate, $14\text{--}18 \times 11\text{--}15\text{ }\mu\text{m}$, evanescent. Ascospores 1-celled, ellipsoid to ovoid, yellowish to pale reddish orange, $6\text{--}8\text{--}5 \times 4\text{--}0\text{--}5\text{ }\mu\text{m}$, thick-walled and nearly smooth (slightly verruculose under SEM).

Conidiophores erect arising more commonly from aerial trailing hyphae, hyaline, smooth-walled, $50\text{--}300 \times 5\text{--}7\text{ }\mu\text{m}$, apical parts irregularly branched and bearing terminal verticils of 2–4 phialides usually without any metulae, phialides solitary or irregularly verticillate, cylindric, $12\text{--}30 \times 3\text{--}6\text{ }\mu\text{m}$. Conidia formed in long divergent or tangled chains, continuous, hyaline but fulvous in mass, cylindrical to elliptical, $4\text{--}11 \times 3\text{--}4\text{ }\mu\text{m}$, conida sometimes ovoid to subglobose, $3\text{--}10\text{ }\mu\text{m}$ diam.

Coonemeria aegyptiaca is mainly distinguished by its ellipsoidal almost smooth ascospores. Ueda & Udagawa (1983) indicate the fungus produces two morphological kinds of asexual structures: the typical *Paeecilomyces*-type with cylindrical to doliform conidia are produced at $37\text{--}40^\circ\text{C}$, at higher temperatures, conidia are subglobose to ovoid, borne in shorter chains on phialides having a swollen and thick-walled apex.

Coonemeria aegyptiaca was recently reported by Abdullah & Al-Bader (1990) to inhabit soil in Iraq

***Coonemeria verrucosa* (Yaguchi, Someya et Udagawa) comb. nov.**

bas.onym *Thermascus crustaceus* (Apinis & Chesters) Stolk var. *verrucosus* Yaguchi, Someya et Udagawa Mycoscience 36: 161. 1995.

Thermascus taitungiacus Chen K-Y & Chen Z-C Mycotaxon 50: 226. 1996
anamorph. *Puccilomyces taitungiacus* Chen K-Y & Chen Z-C Mycotaxon 50: 226. 1996

= *Puccilomyces crustaceus* Apinis & Chesters pro parte fide Yaguchi, Someya et Udagawa Mycoscience 36: 161. 1995.

Descriptions: Yaguchi *et al.* (1995); Chen & Chen (1996).

Colonies in agar: malto addito dispositae post 7 diebus et temperatura 40° C cum 9.0 cm diametro lanatae. Ascوماتes superficiales intermixta cum paucis conidiogenis structuris. Mycelium ex hyphis hyalinis, septatis, ramosis, laevibus, 2-8 µm crassis.

Ascoma non-ostiolatum solitarium ad confluentem deinde crustaceum sphaericum aurantiocanicum cum 300-600 µm diametro. Peridium crassum pseudoparenchymatum textura angulare. Asci dispersi in ascomatis cavitate subglobosi vel pyriformes, 12-16 × 11-14 µm octospori evanescentes. Ascospores unicellulares ellipsoideae rari subglobosae hyalinae ad pallidae aurantiacae, 6-8 × 5-6 µm, cum crasso verrucoso pariete.

Conidiophorum septum, laeve hyalinum ad brunnescum 100-300 × 6-10 µm. Apicalis regio irregulariter ramosa, terminales rami cum solitariis phialidibus vel verticillatis per 2-4. Phialis cylindrica 16-30 × 4-6 µm. Conidia disposita in catenis non coalescentibus, conidia cylindrica, flavida laeva, 4-10 × 2-4 µm, conidia elliptica aliquando 5-8 × 4-6 µm.

Holotypus PF 1160 cultura exsiccata ex soli isolata a I. Yaguchi, Guangzhou in Sma. 4 XI 1993. In herbario Musci et Instituti Historiae Naturalis Chiba. CBM deposita

On common laboratory agar medium, colonies filling the plate in 7 d at 40° C, lanate with superficial ascomata intermixed with sparse conidiophores and conidia, rosy buff to orange, reverse reddish-brown, conidiogenesis inconspicuous not affecting colony appearance. Optimal growth between 30 and 40° C, the minimum lies between 20 and 25° C and the maximum somewhat above 55° C.

Ascomatal initials a coiled hyphae. Cleistothecia solitary or confluent and then forming a crusty layer, orange, spherical, 300-600 µm diam., peridium pseudoparenchymatous of *textura angularis* type, outer layer consisting of thick-walled yellow brown angular cells, 4-8 × 2-6 µm, inner layer of hyaline, angular or rounded cells, 10-20 µm wide. Asci irregularly disposed, 8-spored, globose to pyriform, 12-16 × 11-14 µm, evanescent. Ascospores unicellular, hyaline to pale-orange, ellipsoidal, rarely subglobose, 6-8 × 5-6 µm, thick-walled, verrucose.

Conidiophores arise from the basal mycelium or from aerial hyphae, stipes hyaline to brownish, septate smooth, 100-300 × 6-10 µm, apical parts not uniformly branched giving rise to irregular verticils of terminal and subterminal secondary branches, these bear phialides either singly or in verticils of 2-4, cylindrical to slightly swollen, 16-30 × 4-6 µm. Conidia produced in long disordered chains, unicellular, cylindrical, truncate at both ends, yellowish, smooth, 4-10 × 2-4 µm, few wider elliptical conidia sometimes produced, 5-8 × 4-6 µm.

Holotype PF 1160, a dried culture of a soil isolate from Guanghou, China, 4. XI 1993, deposited at the Natural History Museum and Institute, Chiba, Japan (CBM) and at T. Yaguchi collection (described as *Thermoascus crustaceus* var. *verrucosus*)

While describing *Thermoascus taitungiacus*, Chen & Chen (1996) were probably unaware of the verrucose variety of *Th. crustaceus* established a year before by the Japanese mycologists Yaguchi *et al.* (1995). Authentic material of both taxa have in common a soil origin and not widely separated original locations. Indeed the former derives from a weed soil located at Taitung in Taiwan, while the latter was isolated from a soil sample taken from the Chinese locality of Guanghou.

Ascospores of the Taiwanese fungus were described as being yellowish green (although overall colony color tends to orange tones), oval to elliptical, rarely subglobose, $6.3-7.5 \times 4.5-5.6 \mu\text{m}$, thick-walled and predominantly echinulate when viewed under light microscope but irregularly verrucose under SEM. A comparison of given SEM pictures for both taxa clearly stress ascospore ornamentation is similar being represented by large well individualised warts of heterogeneous shape.

Coonemeria verrucosa is mainly distinguished by its definitely verrucose ellipsoidal ascospores. These are smooth in *C. aegyptiaca* and finely echinulated in *C. crustacea*. The ascomycete might have been previously mislead with *Coonemeria crustacea*. Nevertheless further comparative studies are required to ascertain differences in growth temperatures relations and other minor morphological characters.

***Corynascus heterothallicus* (van Klopotek) von Arx — Sydowia 34: 25. 1981(1982).**

basionym *Thielavia heterothallica* van Klopotek Archives of Microbiology 107: 223-224, 1976.

anamorph *Myceliophthora thermophila* (Apinis) van Oorschot Persoonia 9: 403, 1977
basionym *Sporotrichum thermophilum* Apinis as 'thermophile' Nova Hedwigia 5: 74, 1963.

Chrysosporium thermophilum (Apinis) van Klopotek Archives of Microbiology 98: 366, 1974.

Descriptions von Arx (1981(1982)), van Klopotek (1976), Domsch *et al.* (1980, as *Thielavia heterothallica*).

Apinis (1963) isolated several strains of a new thermophilic "*Sporotrichum*" anamorph from soil and plant debris in Nottingham, UK, he was not aware of the heterothallic nature of the corresponding teleomorph. Few years later, von Arx (1971) provided a modern definition of *Sporotrichum* Link based on a freshly isolated strain matching the type species *S. aureum* Link. S. F. Gray. This form genus was then restricted to hyphomycetes having basidiomycetous affinities as evidenced by the regular presence of clamp connections at the cross walls and production of simple types of aleuriospores. On account of this new definition, van Klopotek (1974) transferred Apinis fungus to *Chrysosporium* Corda.

Myceliophthora Costantin was reintroduced by van Oorschot (1977) as suggested earlier by von Arx (1973) in his treatment of *Sporotrichum* and related genera. This disposition aimed to accommodate the type species *Myceliophthora lutea* Costantin and the *Chrysosporium* (*Sporotrichum*) anamorphs of two hitherto described ascomycetes. *M. lutea* has not yet developed a corresponding teleomorph. The fungus described by

Apinis is now widely accepted as belonging to *Myceliophthora*. It differs from known species by its dark colored colonies and smaller mostly obovate conidia, $4.5-11.0 \times 3.0-4.5 \mu\text{m}$; conidia are hyaline, smooth and thick-walled.

The teleomorph was later discovered by van Klopotek (1976) after mating appropriate strains. Developed cleistothecia produced ascospores ellipsoidal, dark brown, $7.5-11.0 \times 5.0-7.0 \mu\text{m}$, provided with one distinctive germ pore. The teleomorph was described as *Thielavia heterothallica*. Few years later, it was relocated in *Corynascus* von Arx which groups ascomycetes having anamorphs of the *Myceliophthora* type (von Arx *et al.*, 1986).

An excellent account of the cultural and physiological characteristics and the distribution of this fungus is provided by Domsch *et al.* (1980).

***Corynascus thermophilus* (Fergus & Sinden) van Klopotek — Archives of Microbiology 98: 366, 1974.**

basionym *Thielavia thermophila* Fergus & Sinden Canadian Journal of Botany 47: 1635, 1969

= *Chaetomium thermophilum* (Fergus & Sinden) B. Lodha In Taxonomy of Fungi: Proceedings of the International Symposium Madras 1973, Part I, 248, 1978.

anamorph *Myceliophthora fergusi* (van Klopotek) van Oorschot Persoonia 9: 406, 1977.

basionym *Chrysosporium fergusi* van Klopotek Archives of Microbiology 98: 366, 1974.

Descriptions: Fergus and Sinden (1969), Hedger and Hudson (1970), van Klopotek (1974), von Arx (1975)

Mating of several pure strains of another thermophilic "*Sporotrichum*" species isolated from mushroom compost in Pennsylvania (USA) developed black ascocarps of a new heterothallic cleistothecial ascomycete. This was described as *Thielavia thermophila* by Fergus & Sinden (1969). No provision was however then made for the corresponding anamorph. This was simply regarded as deviating from *Sporotrichum thermophilum* Apinis by some cultural characteristics. Both hyphomycetes were later compared by Hedger & Hudson (1970) following isolates obtained in Britain. Distinctive growth and morphological features were also simply underlined by Hedger and Hudson.

In 1974, van Klopotek ascribed the anamorph of *Thielavia thermophila* to *Chrysosporium* Corda while dedicating the hyphomycete to Fergus, she also transferred the teleomorph to the recently established *Corynascus* von Arx. *Chrysosporium fergusi* was later on relocated in *Myceliophthora* by van Oorschot (1977) together with the anamorph of a second *Corynascus* species. However it is unfortunate the specific epithet *thermophila* was not selected for the anamorph of *Corynascus thermophilus*. Such would have prevented any form of confusion with the anamorph of *Corynascus heterothallicus* (van Klopotek) von Arx named *Myceliophthora thermophila* (Apinis) van Oorschot.

Corynascus thermophilus being a heterothallic ascomycete is usually only represented by its ascospores in isolation studies. These are ellipsoidal or obovate, nearly hyaline and measure $7-12 \times 5-8 \mu\text{m}$. Mating of appropriate strains would produce cleistothecia black, globose, usually smooth, up to $300 \mu\text{m}$ diam. Asci are irregularly disposed, each having four ascospores being ellipsoidal, dark brown, $22-32 \times 17-22 \mu\text{m}$, provided with two prominent germ pores.

Corynascus novoguineensis (Udagawa & Horie) von Arx also produces a *Myceliophthora* anamorph but yet unnamed. However this fungus has its maximum growth temperature at 40° C as compared to 55° C for *Myceliophthora fergusii* (van Oorschot, 1980).

***Dactylomyces thermophilus* Sopp — Skrifter udgivne af videnskabs-selskabet i Christiania. Matematisk-naturvidenskabelig klasse 11: 35. 1912.**

= *Thermoascus thermophilus* (Sopp) von Arx — The Genera of Fungi Sporulating in Pure Culture: 84. 1970

anamorph: *Polypaectium* sp.; *fide* Apinis, 1967.

Misapplied names: *Thermoascus aurantiacus* Miele 1907, *sensu* Cooney & Emerson Thermophilic Fungi. 39. 1964.

Penicillium thermophilus (Sopp) Biourge 1923, *sensu* Cooney & Emerson — Thermophilic Fungi. 39. 1964.

= *P. thermophilum* (Sopp) Sacc. *fide* Trotter 1931 — *Sylloge Fungorum* 25 (Suppl. 10) 671.1931; *sensu* Cooney & Emerson — Thermophilic Fungi: 39. 1964.

Description: Apinis (1967).

This is the type species of *Dactylomyces* Sopp (Sopp, 1912). The original material developed in Norway on the wooden casing of a bath thermometer. In the protologue, Sopp suggested the new genus might be identical with *Thermoascus* Miele, despite his awareness of marked differences between respective type species, for example, the presence of a penicillioid anamorph in his fungus and the absence of a distinctive conidial state in the type species *Thermoascus aurantiacus*. Absence of authentic material for effective comparison made a considerable impact on subsequent interpretations of both genera. Such resulted in much confusion in the identity of respective type species.

The presence of a penicillioid anamorph in the description of *Dactylomyces thermophilus* lead Biourge (1923) to list this name among the hitherto known *Penicillia*. However Biourge did not include the corresponding *Penicillium thermophilus* in his group of accepted species. Trotter (1931) published a short description of *Penicillium thermophilum* (Sopp) Sacc. Later and for their treatment of *Penicillia*, Raper & Thom (1949: 20) examined a culture obtained by Prof. Ralph Emerson from retting guayule at Salinas, California, believed to represent Sopp's fungus. In addition to developed ascospores, conidial structures were found by Raper & Thom to be very large and coarse, evanescent, somewhat penicillate and thus not characteristic of their concept of *Penicillium* Link. These authors then regarded *Thermoascus* as a possible synonym of Sopp's genus.

Few years after the publication of Raper & Thom's Manual of the *Penicillia* (1949), Raper & Fennell deposited at the CBS the strain NRRL 1563 as *Dactylomyces thermophilus* Sopp (List of Cultures, Supplement 1. 1962), this number originally referred to a strain of *Cephalophora tropica* but from about 1950, it was discovered that it has been replaced by a strain of this ascomycete.

In 1964, Cooney & Emerson provided a latin diagnosis for *Thermoascus aurantiacus* based on their strain M 206516. This isolate was made from retting guayule in June 1945, at Salinas, California, but there is no clear indication whether it is the same strain earlier examined by Raper & Thom (1949). For this ascomycete, Cooney & Emerson depicted a distinctive conidial stage. Their illustration approximate figures produced by

Sopp (1912) for his *Dactylomyces thermophilus* and which are rather reminiscent of *Paeclomyces* Bain. Cooney & Emerson also admitted Sopp's suggestion his fungus being identical to *Thermoascus aurantiacus*. They substantiated their conclusion by the assumption that isolates of the latter examined by Miehé (1907) might represent "some naturally occurring strains of *Thermoascus* (*Dactylomyces*) incapable of producing conidia". As *Thermoascus* predates *Dactylomyces*, the former was thus retained. Earlier synonyms of the type species of the latter were then disposed under *Thermoascus aurantiacus*. At the date of publication of Cooney & Emerson's book on thermophilic fungi, *Dactylomyces* was still a monospecific genus.

In 1964 however, Apinis & Chesters (1964) added a second species to *Dactylomyces*, *D. crustaceus*, developing a conspicuous *Paeclomyces* anamorph designated *P. crustaceus*. For this work, they re-examined Raper & Fennell's *Dactylomyces thermophilus* strain (NRRL 1563, CBS 374.62), this proved to match the fungus they were proposing. One year later, Stolk (1965) also studied this isolate, she concurred with Apinis & Chesters's decision. Stolk then suggested Cooney & Emerson's neotype of *Thermoascus aurantiacus* (M 206516) is most likely identical with *Dactylomyces crustaceus*.

Stolk (1965) also examined all cultures maintained at the CBS as *Thermoascus aurantiacus* CBS 256.34, CBS 257.34, CBS 415.62 and CBS 398.64. These strains were characterized by the presence of ascospores being elliptical and finely echinulated, $5.0-7.0 \times 3.5-5.0 \mu\text{m}$, and the general absence of an associated anamorph producing chains of conidia. Miehé (1907) had already stressed the absence of any conidial state producing spore chains in his type species and such was confirmed few years later by Noack (1912). Stolk then stressed these features should distinguish Miehé's fungus from *Dactylomyces crustaceus* Apinis & Chesters (having a *Paeclomyces* anamorph) and also from the yet unclear *D. thermophilus* Sopp. Unfortunately Stolk (1965) transferred *Dactylomyces crustaceus* to *Thermoascus*.

In 1967, Apinis re-examined *Thermoascus aurantiacus* neotype strain M 206516. He found it to rather correspond to *Dactylomyces crustaceus* Apinis & Chesters having a well defined *Paeclomyces* anamorph. This observation enabled Apinis to definitely refute the similarity of these two teleomorphic genera as stated by Cooney & Emerson (1964). At that time, Apinis was already familiar with the ascomycete described by Miehé, from pasture soils, he had isolated (Apinis, 1963) several strains matching the original description.

Based on several fresh isolates originating from Sweden and England, Apinis (1967) then provided an updated description of *Dactylomyces* type species, *D. thermophilus*. The fungus has hyaline, unicellular ascospores, more or less oval and smooth, $5.5-8.0 \times 3.5-6.0 \mu\text{m}$. Conidia are also produced, these are continuous, cylindrical to ovoid, subhyaline, smooth, $3.0-11.0 \times 2.5-5.5 \mu\text{m}$. Such ascospores and conidial dimensions are somewhat smaller than those reported by Sopp (1912), however there is a general agreement that measurements given by this author are unreliable. Apinis (1967) selected as neotype strain BDUN 394 (=IMI 123298) obtained by T. Nilsson in Sweden. No provision was established for the anamorph. He simply indicated chains of conidia are produced by branched annellophores as in the recently described form genus *Polypaeclium* G. Smith (Smith, 1961). Apinis proposed two new subgenera to consider distinctiveness in anamorphs of both *Dactylomyces*. Subgenus *Dactylomyces* based on the type and subgenus *Paeclomyces* based on *D. crustaceus*. These subgeneric divisions were rapidly regarded as superfluous.

In spite of clarifications introduced by Apinis (1967), von Arx (1970) listed *Dactylomyces* as congeneric with *Thermoascus*, a taxonomic disposition he maintained for

several years (von Arx, 1987), apparently he was following Stolk (1965) who did transfer *Dactylomyces crustaceus* to *Thermoscypha*. However this generic synonymy did not gain general acceptance (Malloch & Cain, 1972; Cannon *et al.*, 1985; Eriksson & Hawksworth, 1993). *Dactylomyces* Sopp is actually considered a valid distinctive generic entity.

***Melanocarpus albomyces* (Cooney & Emerson) von Arx — Studies in Mycology 8: 17. 1975.**

basionym *Myriococcum albomyces* Cooney & Emerson — Thermophilic Fungi 60: 1964
Thielavia albomyces (Cooney & Emerson) Malloch & Cain — Canadian Journal of Botany 50: 65. 1972

Descriptions: Cooney & Emerson (1964), von Arx (1975), von Arx *et al.* (1988), Guarro *et al.* (1996)

Cooney & Emerson ascribed this fungus to the sterile form-genus *Myriococcum* Fr. based on Corda's interpretation of its type species, *M. praecox* (Corda, 1842). They based their decision on the account both fungi have in common "the dark, shiny, astomous fruiting bodies, associated with a white, mucedineous subiculum". Such an addition was made in spite of their awareness no asci were ever reported in *Myriococcum praecox* and also that what was considered as "spores" by Corda was apparently only the inner cells of immature ascocarps.

The genus *Melanocarpus* von Arx (von Arx, 1975) was later proposed to accommodate this widespread ascomycete known to produce in culture a characteristic arthroconidial state. This anamorph is not developed by taxa of the two related genera *Thielavia* Zopf and *Chaetomidium* (Fuekel) Zopf. Smooth ascomata and obovate-oblate darker ascospores provided with a prominent germ pore ($13.16 \times 11.14 \times 9.11 \mu\text{m}$) were also then considered additional distinguishing features. Further, the presence of a pseudoparenchymatous wall in *Melanocarpus albomyces* precludes any confusion with the hitherto known species of *Thielavia* (von Arx, 1975).

The original concept of *Melanocarpus* was however subsequently partly expanded by von Arx *et al.* (1988) to allow provision for the mesophilous *M. oblatulus* Guarro & van der Aa described in the meantime, arthroconidia are produced by this species. This tendency was also recently substantiated by Guarro *et al.* (1996) with their description of the mesophilic *Melanocarpus coprophilus* Guarro & Vallados, and the transfer of *Thielavia minuta* var. *thermophila* Abdullah & Al-Bader. However both latter taxa are not known to have an associated arthroconidial state or any other state, a feature that should have favoured their inclusion rather in *Chaetomidium*.

***Melanocarpus thermophilus* (Abdullah & Al-Bader) Guarro, Abdullah & Al-Bader — Mycological Research 100: 75. 1996.**

basionym *Thielavia minuta* (Cain) Malloch & Cain var. *thermophila* Abdullah & Al-Bader — Basrah Journal of Agricultural Science 5: 116. 1992

Descriptions: Abdullah & Al-Bader (1992); Guarro *et al.* (1996).

Living strains of the thermophilic variety of *Thielavia minuta* (Cain) Malloch & Cain (a mesophile) originate from a forest soil in the north of Iraq. Re-examination of authentic material led Guarro *et al.* (1996) to relocate the fungus in *Melanocarpus* von Arx.

as *M. thermophilus*. However this ascomycete produces not the arthroconidial anamorph characteristic of the type species. Asci are 8-spored with ascospores being ovoid, dark brown, $7.5-9.0 \times 6.0-7.5 \mu\text{m}$, each provided with a single germ pore.

***Talaromyces byssochlamydioides* Stolk & Samson — Studies in Mycology 2: 45. 1972.**

anamorph *Paecilomyces byssochlamydioides* Stolk & Samson — Studies in Mycology 2 45. 1972

Descriptions. Stolk & Samson (1972); Awao & Otsuka (1974).

This species of *Talaromyces* Benjamin is definitely less reported than its close relative *T. emersonii* Stolk. It is mainly distinguished by its conspicuous *Paecilomyces* anamorph as compared to the *Penicillium* state of the latter. Ascomata always develops in culture concomitantly with the anamorph and such prevents its confusion with the similar imperfect taxon *Paecilomyces variotii* Bamber. Ascospores are globose to subglobose, $3.7-4.5 \times 3.5-4.0 \mu\text{m}$, thick-walled smooth or nearly so, often partially covered by material which may represent the remnants of a gelatinous covering.

Dactylomyces crustaceus Apinis & Chesters also has a *Paecilomyces* state but its conidia are ellipsoidal as compared to the cylindrical conidia of *Talaromyces byssochlamydioides*. The latter was apparently only reported from soil in Japan (Awao & Otsuka, 1974) and Egypt (Mouchacca, 1995).

***Talaromyces emersonii* Stolk — Antonie van Leeuwenhoek 31: 262. 1965; Stolk & Samson — Studies in Mycology 2: 48. 1972.**

— *Byssochlamys* sp. fide Cooney & Emerson — Thermophilic Fungi 155. 1964
anamorph *Penicillium emersonii* Stolk — Antonie van Leeuwenhoek 31: 262. 1965

Geosmithia emersonii (Stolk) Pitt — Canadian Journal of Botany 57: 2027. 1979
Misapplied names: *Talaromyces dupontii* (Griffon & Maublanc) Apinis, *sensu* Apinis
Nova Hedwigia 5: 72. 1963; as *comb. nov. (nom. inval., Art. 36.1)*.

• *Penicillium dupontii* Griffon & Maublanc 1911, *sensu* Apinis — Nova Hedwigia 5: 72. 1963

Descriptions. Stolk (1965), Stolk & Samson (1972), Domsch *et al.* (1980)

Talaromyces emersonii was described inclusive of its distinctive anamorph developing *Penicillia* of the *Asymmetrica* type. The selected representative strain was obtained by Mrs. A. J. van der Plaats Niterink from Italian compost but other isolates were also then available at the CBS. Dedicated to R. Emerson for his contribution to our knowledge of thermophilic fungi.

Apinis (1963) based his taxonomic decision on strain BDUN 272 (CBS 397.64) isolated from soil near Nottingham (UK). Stolk (1965) re-examined this isolate which proved to represent *Talaromyces emersonii* rather than the teleomorph of *Penicillium dupontii* as concluded by Apinis. The same observation also applies to strain CBS 394.64 labelled *Byssochlamys* sp. by Cooney & Emerson (Stolk, 1965).

The anamorphic genus *Geosmithia* Pitt (Pitt, 1979) was erected to accommodate *Penicillia* formerly placed in the *Penicillium pallidum* series. These are mainly distinguished by the formation of cylindrical conidia borne from cylindroidal, rough walled

phialides and not colored green en masse. Stolk & Samson (1985) emitted doubts as to the necessity of such a generic distinction based on slight morphological differences. In *Penicillium* such differences are instead appropriately used to delimit generic sections. The anamorph of *Talaromyces emersonii* is thus better referred to as a *Penicillium*.

Talaromyces emersonii was subsequently reported from various habitats (Domsch *et al.*, 1980). It produces globose, reddish to orange brown ascomata, up to 300 µm diam., ascospores are thick-walled, smooth, subglobose to ovoidal, $3.5-4.0 \times 2.7-3.5$ µm, ascospores may be covered by material, representing remnants of a gelatinous coating.

***Talaromyces thermophilus* Stolk — Antonie van Leeuwenhoek 31: 268. 1965; Stolk & Samson — Studies in Mycology 2: 55. 1972.**

Penicillium dupontii Griffon & Maublanc emend. Emerson in Raper & Thom A Manual of the Penicillia, 573-577. 1949.

— *Talaromyces dupontii* (Griffon & Maublanc) Emerson, incidently mentioned by Fergus Mycologia 56: 277. 1964 (*nom. inval.*, Arts. 36.1 & 37.1)

anamorph *Penicillium dupontii* Griffon & Maublanc — Bulletin trimestriel de la Societe mycologique de France 27: 73. 1911

— ? *Citromyces sphagnicola* Mal'chevskaya — Trudy Pushkin nauchno-issled. Lab. Rasv. Sel'skokhoz. Zhivot. Inst. 13: 23. 1939.

Misapplied names: *Talaromyces dupontii* (Griffon & Maublanc) Apinis, *sensu* Apinis Nova Hedwigia 5: 72. 1963, as *comb. nov.* (*nom. invalid*, Art. 36.1).

Talaromyces *Penicillium dupontii* (Griffon & Maublanc) emend. Emerson in Raper & Thom 573. 1949, l.c. Cooney & Emerson — Thermophilic Fungi 28. 1964 (*nom. inval.*, Art. 36.1).

Descriptions : Stolk (1965); Stolk & Samson (1972); Pitt (1979).

The original publication of Griffon & Maublanc (1911) dealt only with the *Penicillium* anamorph. No corresponding teleomorph was reported from cultures of the two strains then available for study. These were obtained in France from manure and damp hay by Mr. Dupont, Chief Chemist at the Ecole Nationale d'Agriculture de Grignon, and to whom the fungus was dedicated. Unfortunately, the two original isolates are definitely lost.

The fungus was later on and in 1945 isolated by Emerson from rotting guayule shrub at Salinas, California, for the first time the perfect state developed in culture. Emerson then prepared an emended description of both states of *Penicillium dupontii* based upon his strain No. 26 (NRRL 2155) to be incorporated by Raper & Thom in their first Manual of the Penicillia (1949: 573).

In 1963, some confusion about this taxon was introduced by Apinis. Under the binomial *Talaromyces dupontii* (Griffon & Maublanc) Apinis, he provided a description of a teleomorphic fungus thought to "correspond in general to the original strain described from France". As representative material, Apinis selected his strain BDUN 272 originating from a water-logged pasture in Nottingham. The above binomial was however not validly published as no latin diagnosis provided and no new type material specified.

Further confusion but of the nomenclatural type was also simultaneously introduced by Fergus (1964) following his study of an isolate from compost in Pennsylvania (USA). His observations were published under the name *Talaromyces dupontii* (Griffon & Maublanc) Emerson, a combination not proposed as such by Emerson himself.

Unfortunately, this designation was subsequently reproduced in several studies of thermophilic fungi.

Two years later Stolk (1965) re-examined Apinis strain BDUN 272. She found it to largely deviate from Emerson's isolate No. 26 (NRRL 2155, CBS 236.58). Moreover, the former proved to perfectly match her newly described *Talaromyces emersonii* Stolk. To eliminate the state of confusion prevailing around the name *Talaromyces dupontii*, Stolk then proposed the new name *Talaromyces thermophilus* for this teleomorph and provided a latin diagnosis. Emerson strain was then selected as holotype (Stolk, 1965; Pitt, 1979). However, the original accession number of this holotype was erroneously cited by Cooney & Emerson (1964: 28) under the diagnosis provided for this fungus is indicated specimen M 206516 (our culture No. 26). In fact strain M 206516 was selected by Cooney & Emerson as representing their interpretation of *Thermoascus aurantiacus* Miehé (1964: 50) and this corresponds to their annotation: our culture No. 2.

Pitt (1979) stressed the reasons why the corresponding perfect state should be maintained in *Talaromyces* and to continue considering its simple reduced anamorph as a *Penicillium*. *Talaromyces thermophilus* is the only thermophile with a *Penicillium* anamorphic state producing green conidia. This character should prevent any misidentification since ascocarps do not always readily develop in cultures of freshly isolated strains. The fungus grows fairly rapidly and optimally at 45-50°C, no growth develops at 25°C and 60°C respectively. Ascospores are ellipsoidal, 3.5-4.5 × 2.2-3.5 µm, ornamented by 2-6 somewhat jagged, irregular, usually longitudinal ridges.

***Thermoascus aurantiacus* Miehé — Die Selbsterhitzung des Heues: 70. 1907.**

— ? *Thermoascus tsatschenkoii* Mal'chevskaya. Trudy Pushkin Nauchno issled. Lab. Razv. sel'khoz. Zhivot. 13: 26. 1939, *vide* Cooney & Emerson. Thermophilic Fungi: 39. 1964.

Misapplied names: *Thermoascus aurantiacus* Miehé 1907, *sensu* Cooney & Emerson. Thermophilic Fungi: 39. 1964.

Dactylomyces thermophilus Sopp 1912, *sensu* Cooney & Emerson. Thermophilic Fungi: 39. 1964.

Penicillium thermophilus (Sopp) Bourge 1923, *sensu* Cooney & Emerson. Thermophilic Fungi: 39. 1964.

Penicillium thermophilum (Sopp) Sacc., *vide* Trotter 1931; *sensu* Cooney & Emerson. Thermophilic Fungi: 39. 1964.

Descriptions Stolk (1965), Apinis (1967), Awao & Otsuka (1973), Domsch *et al.* (1980), Chen & Chen (1996).

The type species of *Thermoascus* Miehé (Miehé, 1907), *T. aurantiacus*, was isolated from self-heating hay and carefully described by the author. Few years later, Sopp (1912) reported a second thermophilic ascomycete having a well developed conidial state and for which he proposed the new genus *Dactylomyces*. Sopp then considered *Thermoascus aurantiacus* as approximating his *Dactylomyces thermophilus* n. sp. *ad interim*. This suggestion coupled with the lack of any authentic material for either ascomycetes resulted in much confusion about the exact nature of Miehé's fungus.

In 1963, Apinis isolated from soil near Nottingham several strains he referred to *Thermoascus aurantiacus*. These isolates exhibited no morphological deviations from the original description. They also proved to match a strain maintained at the CBS under this

binomial and isolated by Noack (1912). Apinis noted the structure of the cleistothecium be related to certain species of the *Gymnoascaceae* with the presence of large clavate conidia reminiscent of "clasterospores" of some *Trichophyton* species.

In 1964, Cooney & Emerson provided a detailed description and a latin diagnosis of *Thermoascus aurantiacus* based on their "strain M 206516 (our culture No. 2)" thought to match Miehe's fungus. This was also then regarded as an earlier name of *Dactylomyces thermophilus*. In the established description, a distinctive conidial state is depicted. This anamorph approximate figures produced by Sopp (1912) and which were later regarded (Stolk, 1965) as rather reminiscent of *Puccinomyces* Bain (see also comments under *Dactylomyces thermophilus*).

However, in the same year, Apinis & Chesters (1964) introduced *Dactylomyces crustaceus* (anamorph *Puccinomyces crustaceus*) and reported to it the CBS strain 374 62 (= NRRL 1563) labelled *Dactylomyces thermophilus* by Raper & Fennell. In 1965, Stolk re-examined this strain and concurred with Apinis & Chester's decision. She then suggested Cooney & Emerson's neotype of *Thermoascus aurantiacus* (M 206516) is most likely identical with *Dactylomyces crustaceus*.

Stolk (1965) then examined all cultures maintained at the CBS as *Thermoascus aurantiacus*: CBS 256 34, CBS 257 34, CBS 415 62 and CBS 398 64. These isolates produced elliptical finely echinulated ascospores, measuring $5.0-7.0 \times 3.5-5.0 \mu\text{m}$. No associated anamorph producing chains of conidia was developed by any. The absence of an anamorph producing spore chains was already stressed by Miehe (1907) and such was confirmed few years later by Noack (1912). Stolk then underlined these features support the distinctiveness of *Thermoascus aurantiacus* from *Dactylomyces thermophilus* (having a yet undefined anamorph) and also from the well described *Dactylomyces crustaceus* and its *Puccinomyces* anamorph. Unfortunately Stolk then transferred the latter to *Thermoascus*.

In 1967, Apinis published a comparative study of *Thermoascus* and *Dactylomyces* based on freshly isolated strains. He re-examined Cooney & Emerson strain M 206516 and confirmed Stolk (1965) suggestion about its similarity with *Dactylomyces crustaceus*. This led to a definite rejection of Cooney & Emerson's taxonomic considerations about *Thermoascus* type species and to its identity with *Dactylomyces thermophilus*. Second, the morphology of Apinis fresh isolates of *Thermoascus aurantiacus* was in line with Stolk (1965) observations.

Apinis also noted the presence in the aerial mycelium of "conidia" of *Aphanascus* or *Microsporium* type—developing terminally on long or short hyphal branches singly and being clavate or somewhat spindle-shaped, smooth, 0-3 septate, $12-35 \times 5-10 \mu\text{m}$. Miehe (1907) did mention such aleuriospores in the type species. Strain BDL N 343 (= IMI 91787) isolated from alluvial grassland soil was designated neotype for *Thermoascus aurantiacus*.

Recent reports confirm *Thermoascus aurantiacus* have a wide distribution (Domsch *et al.*, 1980; Chen & Chen, 1996). This ascomycete proved to be a strong thermophile with growth starting at 30° C and up to 62° C, growth optimum around 45° C with formed colonies being bright orange to orange brown. Ascospores are definitely elliptical and slightly roughened. Presence of terminal aleuriospores apparently depends on examined strains.

Thermoascus was placed in the family *Omygenaceae* (order *Onygenales*) by Benny & Kimbrough (1980); they placed *Dactylomyces* in the *Trichocomaceae* (order *Eurotiales*). It was maintained in this family by von Arx (1987) but with *Dactylomyces* being a synonym. *Thermoascus* was however excluded by Currah (1985) from the *Omygenaceae*.

because "there is no evidence of keratinolytic abilities nor does it have strictly rheolytically dehiscing conidia". Currah mentions not *Dactylomyces*.

Thermoascus tsatschenkoi is regarded as a doubtful species of which no satisfactory description exists and no material is available for comparison (Cooney & Emerson, 1964, Apinis, 1967).

***Thermoascus aurantiacus* Miehe var. *levisporus* Upadhyay, Farmelo, Goetz & Melan — Mycopathologia 87: 73. 1984.**

The original isolate was obtained from a top layer soil at La Ceiba, Republic of Honduras. Minimum and maximum growth temperatures are 31 and 61 °C respectively with the optimum being at 49–50 °C. The variety differs mainly by ellipsoidal smooth rather than "echinulate ascospores", $3.7-7.1 \times 2.2-5.5 \mu\text{m}$ ($5.0-7.0 \times 3.5-5.0 \mu\text{m}$ for the species). Conidial anamorph of the aleurospore type matching those of the species were infrequent, borne terminally, clavate, thick walled, smooth, $15.25 \times 7-17 \mu\text{m}$. All other characters duplicate the species. Production of protease enzymes was also assessed (Marcy *et al.*, 1984).

***Thielavia australiensis* Tansey & Jack — Canadian Journal of Botany 53: 82. 1975.**

Descriptions Tansey & Jack (1975), von Arx (1975), von Arx *et al.* (1988)

The protologue was based on strains isolated from nesting material of an incubator bird, the mallee fowl *Leipoa ocellata* Gould in New South Wales, Australia. Optimum growth recorded at 35–40 °C, maximum at 50 °C, minimum not defined.

This *Thielavia* is distinguished by small pyriform brown ascospores, $6.8 \times 5-6 \mu\text{m}$, having a germ pore at the attenuated end. Simple aleuriocomidia are produced in culture according to the protologue, these are continuous, lateral, sessile, colorless, ovoid, $5-8 \times 3-5 \mu\text{m}$. The fungus has apparently not been reported after its description (von Arx *et al.*, 1988).

***Thielavia pingtungia* Chen K-Y. & Chen Z-C. — Mycotaxon 60: 242. 1996.**

The fungus was isolated from a sugar-cane field in Taiwan. The specific epithet refers to the original locality Pingtung. No growth developed between 25 and 30 °C with the optimum being around 40 °C and the maximum fairly above 50 °C.

The species is characterized by dark globose cleistothecia covered with brown thick-walled hairy appendages, ascomatal hairs of the *Chaetomium* type, $2.5-4.0 \mu\text{m}$ wide and up to $350 \mu\text{m}$ long, ascomatal wall pseudoparenchymatous. Asci cylindric, $40-52 \times 7-9 \mu\text{m}$, stipitate, fasciculate, 8 spored. Ascospores usually uniseriate, globose to subglobose dark brown smooth, thick walled, $8.5-10.0 \times 6.5-8.5 \mu\text{m}$. No anamorph developed in examined cultures.

Thielavia pingtungia have several features in common with species assigned to *Chaetomium*, for the moment, the latter groups only mesophilic ascomycetes (Silva & Hanlin, 1996).

***Thielavia terrestris* (Apinis) Malloch & Cain — Canadian Journal of Botany 50: 66. 1973.**

basionym: *Allescheria terrestris* Apinis — Nova Hedwigia 5: 68. 1963.
anamorph: *Acremonium alabamense* Morgan Jones as '*alabamensis*' — Canadian Journal of Botany 52: 429. 1974

Descriptions. Apinis (1963); Malloch & Cain (1973); von Arx (1975)

The original material was observed by Apinis (1963) in the course of his work on thermophilous fungi inhabiting alluvial soils in Great Britain. He described the fungus as *Allescheria terrestris* without providing any argument favouring such a decision, he also assigned the anamorph to *Cephalosporium* (now *Acremonium* Link Fr.). *Allescheria terrestris* was then transferred to *Thielavia* Zopf. Following its description, the fungus was reported from various habitats and is now known to display a wide geographic distribution. Ascospores are ovate or pyriform, brown, thick-walled, provided with a distinct germ pore at the attenuated end, $5.0-7.5 \times 4.0-5.5 \mu\text{m}$.

The hyphomycete *Acremonium alabamense* was described exclusive of a teleomorph, it was isolated from needles of *Pinus taeda*. Later on, Samson *et al.* (1977) found it to match the anamorph of *Thielavia terrestris*. For this ascomycete, sometimes only the anamorph is observed during isolation studies and appropriate matings are required for the development of the teleomorph. These authors conducted extensive mating experiments with several strains of *T. terrestris* and *A. alabamense*, they came to the conclusion that the mating behaviour of *Thielavia terrestris* could best be interpreted as indicating homothallism with cross-feeding.

Some species of *Chaetomium* also produce in culture a phialidic state approximating *Acremonium alabamense*. The latter was recently selected as the type of the new section *Chaetomionides* of *Acremonium* established to also accommodate phialidic states of some *Chaetomium* species (Morgan-Jones & Gams, 1982).

THERMOPHILIC HYPHOMYCETES

***Acremonium alabamense* Morgan-Jones as "*alabamensis*" — Canadian Journal of Botany 52: 429. 1974.**

teleomorph: *Thielavia terrestris* (Apinis) Malloch & Cain — Canadian Journal of Botany 50: 66. 1973

Descriptions. Apinis (1963), Morgan-Jones (1974), Morgan-Jones & Gams (1982)

As underlined before, this hyphomycete was described exclusive of the teleomorph being isolated from needles of *Pinus taeda* collected in the state of Alabama (USA). The teleomorph was described before from alluvial soils in Nottingham (UK) with the anamorph being indicated as simply representing a *Cephalosporium* sp.

The repeated isolation of an *Acremonium* sp. from heated habitats led Samson *et al.* (1977) to compare it with the fungus described by Morgan-Jones and the anamorph of

Thielavia terrestris. All three hyphomycetes were found to represent the same fungus. This finding rose few questions concerning the developmental behaviour of the teleomorph. Extensive mating studies were then undertaken with isolates of *Thielavia terrestris*, *Acremonium* sp. and of *A. alabamense*. Although results allowed not a definite conclusion as to the heterothallic nature of the teleomorph, these support the hypothesis indicating the mating behaviour of *Thielavia terrestris* is a case of homothallism with cross-feeding (Samson *et al.*, 1977).

Acremonium alabamense could thus be observed alone in studies involving high temperatures incubation. It was recently selected as type of the new section *Chaetomioides* of the genus established to also accommodate the morphologically similar phialidic states of some *Chaetomium* species (Morgan-Jones & Gams, 1982). The fungus produces comparatively fast growing colonies, velvety, whitish, with yellowish to brownish runner hyphae, 3-4.5 µm wide. Conidiophores are simple, short, 8-25 × 1-1.5 µm. Conidia are obovoid to pyriform, smooth, with a truncated base, 3-6 × 2-3 µm (Morgan Jones, 1974).

***Acremonium thermophilum* W. Gams & Lacey — Transactions of the British mycological Society 59: 520. 1972.**

The described material developed on self heated sugar cane bagasse in Trinidad. The fungus is regarded as unique among known *Acremonium* Link. Fr. species on account of its thermophilic habit and production of submerged hyphae partly having pigmented walls. The species was assigned in *Acremonium* sect. *Nectria* due to the development of thick-walled conidiophores with basitonsous ramification. Growth is strong but slow at 20° C, very good between 25 and 40° C and very weak at 47° C. Conidia are ellipsoidal, 3.0-4.0 × 1.3-1.7 µm.

***Humicola hyalothermophila* Moubasher, Mazen & Abdel-Hafez — Transactions of the British mycological Society 72: 509. 1979.**

Descriptions: Moubasher *et al.* (1979); Moubasher (1993).

This soil-borne hyphomycete was originally isolated from several localities in Jordan. No growth develops either at 28 or 55° C with good development being at 45° C, growth optimum value is not specified. This thermophile was distinguished from the mesophilic *Humicola fuscoatra* Traaen mainly by its slightly larger hyaline conidia (not colored light brown as in *H. fuscoatra*) and intercalary chlamydospores. It was later on observed in Saudi Arabian soils (Bokhary, 1986).

The taxonomic position of this fungus needs to be re-assessed.

***Malbranchea cinnamomea* (Libert) van Oorschot & de Hoog — Mycotaxon 20: 129. 1984.**

basionym *Trichothecium cinnamomeum* Libert — *Plantae cryptogamae Arduenna*, Coll. I, Nr. 1013. 1830

— *Geotrichum cinnamomeum* (Libert) Sacc — *Revue Mycologique* (Toulouse) 11: 55. 1881; *Michelia* 2: 636. 1882

— *Thermodeum sulfureum* Mieh — *Deutsche Botanische Gesellschaft* 25: 515. 1907

— *Malbranchea pulchella* Sacc — *Sydow, Annals Mycologici*, Ser II, 6: 557. 1908, Sacc & Traverso — *Sylloge Fungorum* 20: 11. 1911.

Malbranchea pulchella Sacc & Penzig var. *sulfurea* (Mieh) Cooney & Emerson — *Thermophilic Fungi*: 102. 1964.

- *Malbranchea sulfurea* (Miehe) Pidoplichko — In "Fungus Flora of Coarse Fodders (in Russian)", 170, 1953.
- *Malbranchea sulfurea* (Miehe) Sigler & Carmichael — Mycotaxon 4: 441, 1976.

Descriptions: Cooney & Emerson (1964), Sigler & Carmichael (1976)

Miehe (1907) erected *Thermothecium*, type species *T. sulfureum*, for a hyphomycete he encountered during his pioneer investigation of the self-heating process of hay. He studied the fungus in culture and stressed its thermophilic nature. Saccardo (1908) however immediately considered this type species as matching the morphologically close mesophilic type species of his genus *Malbranchea*, *M. pulchella* Sacc.

In 1964, Cooney & Emerson provided an excellent account of a strain matching Miehe's description. For this fungus, they simply proposed the varietal name *sulfurea* pending a comprehensive comparison with the almost identical mesophile *Malbranchea pulchella*. The comparison was later undertaken by Sigler & Carmichael (1976); they concluded high temperature requirements are sufficient to warrant a specific status and provided the binomial *Malbranchea sulfurea* (Miehe) Sigler & Carmichael.

The combination *Malbranchea cinnamomea* was based on *Trichothecium cinnamomeum* Libert. It was established by van Oorschot & de Hoog (1984) after examining dried authentic material of the latter. However the possible similarity with *Malbranchea sulfurea* was not considered. Such was established later on by Sigler (1987) after a study of appropriate authentic material.

Malbranchea cinnamomea is an easily recognizable thermophilic hyphomycete being recorded on a variety of substrates under different conditions (Sigler & Carmichael, 1976).

***Myceliophthora fergusii* (van Klopotek) van Oorschot — Persoonia 9: 406, 1977.**

basonym *Chrysosporium fergusii* van Klopotek — Archives of Microbiology 98: 366, 1974.

Teleomorph *Corynascus thermophilus* (Fergus & Sinden) van Klopotek — Archives of Microbiology 98: 366, 1974.

Descriptions: van Klopotek (1974); van Oorschot (1977, 1980).

As underlined under the teleomorph, provision for the anamorph of this ascomycete was made several years after the discovery of the heterothallic nature of the perfect state. The anamorph was simply stated as being distinct from the close previously described hyphomycete now renamed *Myceliophthora thermophila*. Both anamorphs can thus be observed separately from their respective teleomorphs in mycological analyses conducted at high incubation temperatures.

Myceliophthora fergusii produces pinkish-cream floccose colonies, aleuriospores are pyriform to clavate, smooth and thick-walled, nearly hyaline and with narrow basal attachments, 5-12 × 3-5 µm.

***Myceliophthora hinnulea* Awao & Udagawa — Mycotaxon 16: 438. 1983.**

The type locality is cultivated soil in Japan. Fungal growth is extremely reduced at 20° C, optimal growth is at 40-45° C and maximum somewhat above 50° C. No connection with a teleomorph yet established.

Myceliophthora hinnulea differs from the five previously described species (van Oorschot, 1980) mainly by dull to greyish brown colonies and brownish conidia conspicuously verrucose to spinulose, 8.0-10.0 × 6.0-7.5 µm. Almost all known members of this genus are thermotolerant or thermophilic with sporulation often being good between 30-40° C.

***Myceliophthora thermophila* (Apinis) van Oorschot — Persoonia 9: 403. 1977.**

basionym *Sporotricum thermophilum* Apinis as '*thermophile*' Nova Hedwigia 5: 74. 1963.

— *Chrysosporium thermophilum* (Apinis) van Klopotek Archives of Microbiology 98: 366. 1974.

— *Myceliophthora indica* Basu Nova Hedwigia 40: 85. 1984 (nom. inval., Art. 37.1).

Teleomorph *Corynascus heterothallicus* (van Klopotek) von Arx Sydowia 34: 25. 1981.

Descriptions: van Oorschot (1977, 1980), van Klopotek (1974)

As stressed before *Myceliophthora thermophila* was described exclusive of its corresponding teleomorph. Since the latter is heterothallic, the anamorph could thus be observed alone in studies involving thermophilic fungi. The species differs from other members of the genus by its dark colored colonies, occasionally greenish and by smaller mostly obovate conidia, 4.5-11.0 × 3.0-4.5 µm, conidia are hyaline, thick-walled and rough. Fresh isolates always have some rough conidia but older cultures tend to produce only smooth ones. The species displays a wide geographic distribution being a common component of decaying manure, silage, wood chips and pulp, etc. (Cannon, 1990).

Myceliophthora indica was isolated from garden soil and from decomposed leaves of *Clitoria* sp. Attempts to locate original material were unsuccessful although Basu (1984) underlined her intention to deposit both available strains at the CBS. The fungus was compared with the type culture of *Myceliophthora thermophila* considered by Basu as being thermotolerant. The "strongly thermophilic" Indian strain was found to deviate mainly by smaller definitely roughened conidia. No mating attempts were undertaken and the existence of a known teleomorph not stressed in the publication. Analysis of the protologue clearly indicates the Indian strain do represent *Myceliophthora thermophila*.

***Scytalidium indonesicum* Hedger, Samson & Basuki — Transactions of the British mycological Society 78: 365. 1982.**

The original material was isolated from soil of the Bogor Botanic Garden, West Java. The fungus was also recovered from *Dipterocarp* forest soils in South Sumatra. It was reported as being simply "thermophilous" able to grow rapidly at 45° C. 8.5 cm at 36 h. Later Straatsma & Samson (1993) stated it is thermophilic.

The Indonesian taxon is distinguished by the production of conidia (intercalary chlamydospores) thick-walled brown, ellipsoid to barrel shaped, often with irregular

outgrowths and also often constricted at the middle of the cell. $15.25 \times 7.12 \mu\text{m}$, on maturity these conidia secede rather easily and appear irregular in shape. Dark brown and thick-walled similar but less wider conidia (termed arthroconidia) also develop in chains. $13.32 \times 5.8 \mu\text{m}$, these do not secede easily. The presence of terminal conidia (or lateral) is not underlined.

Scytalidium indonesicum approximates *S. thermophilum* which mostly produces spherical to subspherical dark brown smooth conidia $9.14 \mu\text{m}$ wide, oblong or ellipsoidal ones measure $8.18 \times 7.14 \mu\text{m}$. However, neither these *Scytalidium* develop the second hyaline arthroconidial state characteristic of the type species, *S. lignicola* (Ellis M. B., 1976). The description of *S. indonesicum* is however in line with the introduction in *Scytalidium* of taxa only developing dematiaceous arthroconidia (Sigler & Wang, 1990). Such additions makes *Scytalidium* a heterogeneous entity.

***Scytalidium thermophilum* (Cooney & Emerson) Austwick — New Zealand Journal of Agricultural Research 19: 29, 1976; emend. Straatsma & Samson — Mycological Research 97: 327, 1993.**

basonym. *Torula thermophila* Cooney & Emerson — Thermophilic Fungi 92, 1964

Humicola insolens Cooney & Emerson — Thermophilic Fungi: 79, 1964

— *Humicola fuscoatra* var. *longispora* forma *insolens* (Cooney & Emerson) Fassatiava Ceska Mykologie 21: 80, 1967.

— *Humicola grisea* Traaen var. *thermoidea* Cooney & Emerson — Thermophilic Fungi 79, 1964

Humicola insolens Cooney & Emerson var. *thermoidea* D. H. Ellis — Transactions of the British mycological Society 78: 133, 1982.

Humicola fuscoatra Traaen var. *nigra* Subrahmanyam — Hindustan Antibiotics Bulletin 24: 41, 1982 (nom. inval., Art. 36.1 description only), *Ibid* 25: 62, 1983 (latin diagnosis, nom. inval., Art. 37.1)

— *Humicola nigrescens* Omvik var. *thermorongcura* Subrahmanyam — Hindustan Antibiotics Bulletin 24: 45, 1982 (nom. inval., Art. 36.1 description only), *Ibid* 25: 62, 1983 (latin diagnosis; nom. inval., Art. 37.1).

= *Scytalidium allahabadum* Naram, Srivastava & Mehrotra — Zentralblatt für Mikrobiologie 138: 570, 1983.

Descriptions Cooney & Emerson (1964), Ellis M. B. (1976), Straatsma & Samson (1993)

Cooney & Emerson (1964) while describing *Humicola insolens* and *H. grisea* var. *thermoidea* indicated "the problems concerned with the *Monotospora* *Humicola*-*Torula* can only be resolved when all forms, both thermophilic and mesophilic, can be studied and compared in detail". The genus *Monotospora* was cited in relation to Mason (1941) who had then concluded that *M. dulae* Mason predates *Humicola fuscoatra* Traaen. *Torula thermophila* was apparently not concerned by this statement since its description is found some twenty pages later. *Humicola grisea* var. *thermoidea* was considered as a variety (although not producing phialospores as the species) "chiefly because of the uncommon occurrence of intercalary chlamydospores". The abundance of these structures was then used to distinguish *Humicola insolens* Cooney & Emerson.

Later, Emerson (1968) stressed "*Humicola grisea* var. *thermoidea* has smooth-walled chlamydospores (aleuriospores) borne singly on short lateral branches with the almost absence of any intercalary chlamydospores, on the other hand, isolates of *Humicola insolens* regularly produces intercalary chlamydospores singly, in pairs or in short

chains in addition to solitary terminal spores on short lateral branches, in *Torula thermophila* chlamydospores are again smooth and brown, all formed in longer or short, intercalary chains and rarely in a terminal position”

The taxonomic status of this *Humicola-Torula* complex remained unchanged until Austwick (1976) transferred *Torula thermophila* to *Scytalidium* Pesante sensu Ellis M. B. (1971), the latter had emphasized the dark pigmented arthroconidia of the type species *S. lignicola*. However Austwick did not provide any argument in favour of such transfer. Later Sigler & Carmichael (1976) in the course of their study of hyphomycetes with arthroconidia accepted *Scytalidium* as delimited by Ellis M. B., seven species which then retained with some developing only the dematiaceous chlamydosporic state. These additions introduced much heterogeneity in the genus (Sigler & Wang, 1990).

Ellis D. H. (1982) conducted ultrastructural studies of the conidial ontogeny in both *Humicola* proposed by Cooney & Emerson (1964). After examining type strains and other isolates, he concluded *H. grisea* var. *thermoidea* is a separate entity exhibiting considerable genetic variation among strains, further, it should rather be considered a variety of *Humicola solis*. This proposal was not in line with the suggestion earlier by Awao & Otsuka (1974) stating that both Cooney & Emerson's *Humicola* represent the same fungus.

The respective status of these three hyphomycetes remained as such until a recent publication made by Straatsma & Samson (1993). They compared a large number of strains labeled *Torula thermophila*, *Humicola grisea* var. *thermoidea* or *Humicola thermophila* including corresponding authentic material. Their conclusion was that all such strains represent one single variable species or a morphologically indistinguishable species complex for which the binomial *Scytalidium thermophilum* (Cooney & Emerson) Straatsma should continue to be applied pending further studies. Such a limitation according to the fact the type species of *Scytalidium* is a dimorphic fungus having in addition to arthroconidia a hyaline anamorphic form, the use of the binomial *Scytalidium thermophilum* as is questioned.

Humicola solis var. *neera* Subrahmanyam was isolated from soil at K. India. The protologue almost duplicates that of *Humicola insolens* sensu Cooney & Emerson except that aetia spores of the Indian strain are indicated as being smaller. Aleutospores are unicellular, rarely bicellular, single, rarely in chains of 2-3, smooth, spherical and 10-20 µm diam., ovoid and 13.0-16.5 × 10.2-16.5 µm or pyriform and 6.5-19.5 × 4.0-14.0 µm. Chlamydospores intercalary with dimensions and position identical to those of the aleutospores.

Humicola grisea var. *thermoidea* Subrahmanyam was isolated from *Rattus* sp. at Madras, India. The provided protologue stress the presence of aleutospores produced singly or in chains, globose (8.5-14.5 µm) or ovoid (11.0-14.0 × 9-12.0 µm) with similar intercalary chlamydospores. This description matches *cola grisea* var. *thermoidea* sensu Cooney & Emerson.

Scytalidium albidum Naram *et al.* developed while examining material from the Allahabad region, India. It was first identified by P. M. Kirk (1973) as *Scytalidium thermophilum* (Naram *et al.*, 1983). However the Indian authors still refer to it as *Scytalidium thermophilum* on account of its colonies colored greyish-black coupled with the production of larger spores with shapes commonly other than globose. In *Scytalidium thermophilum*, globose spores vary from 4.5-12.0 µm while those of other shapes measure 4.5-13.3-11.0 µm (10.0-2.5 × 7.5-10.0 µm for an Indian strain of *Scytalidium thermophilum*). Re-examination of the type material (IMI 243118) confirm it represents a *Scytalidium thermophilum*.

***Thermophymatospora fibuligera* Udagawa, Awao & Abdullah — Mycotaxon 37: 100-101. 1986.**

Thermophymatospora Udagawa *et al.* (1986) was proposed for an unusual soil-borne hyphomycete assignable to a basidiomycete anamorph. The original strain of the type species *T. fibuligera* derives from an Iraqi date palm plantation. It is characterized by aloblastic unicellular conidia being terminal or lateral, large, brownish, globose, thick-walled and tuberculate, 20-25 µm wide. Such conidia are superficially reminiscent of some *Mechophthora* species. However the hyphae of this hyphomycete are regularly provided with simple clamp connections at the transverse septa. No link with a particular teleomorph has yet been established.

The fungus growth and sporulation are optimal around 35-40° C, almost nil at 20° C, with maximum being at 45° C.

***Thermomyces ibadensis* Apinis & Egginis — Transactions of the British mycological Society 49: 631. 1966.**

This hyphomycete was first recorded during studies of micro-organisms responsible for the biodeterioration of palm kernels in Nigeria. The minimum temperature for growth is between 31-35° C, optimum lies around 42-47° C and maximum at 60-61° C. This *Thermomyces* differs from the type species *T. lanuginosus* by its smaller unicellular, spherical, smooth, brown conidia, 4.0-8.0 µm wide, and by its slender and more frequently branched conidiophores.

***Thermomyces lanuginosus* P. Tsiklinsky (*sensu* Miehe 1907) — Annales de l'Institut Pasteur, Paris 13: 500-505. 1899.**

Sepedonnum lanuginosum ('Miehe') Griffon & Maublanc — Bulletin de la Société Mycologique de France 27: 70. 1911.

Monotospora lanuginosa (Griffon & Maublanc) Mason — Mycological Papers 3: 59. 1933.

Humicola lanuginosa (Griffon & Maublanc) Bance as '*lanuginosus*' — Transactions of the British mycological Society 44: 375. 1961.

Acremonium sp. Rege — Annales of Applied Biology 14: 28. 1927, *vide* Mason, 1933.

A. thermophila Carzi — Atti dell'Istituto botanico dell'Università di Pavia, Ser. 4: 154. 1929, *vide* Mason, 1933.

Humicola grisea Traaen var. *indica* Subrahmanyam — Current Science 49: 30. 1980 (*nom. inval.*, Art. 36.1).

Humicola lanuginosa (Griffon & Maublanc) Bance var. *catenulata* Morinaga in Morinaga, Kanda & Nomi — Journal of Fermentation Technology 64: 452. 1986.

Descriptions Cooney & Emerson (1964), Barron (1968), Ellis M. B. (1971), Domsch *et al.* (1980).

Thermomyces was introduced by Tsiklinsky (1899) for one species, *T. lanuginosus*, isolated from garden soil, the original isolate was however not maintained. Miehe (1907) retained this binomial for an isolate from composted hay. Griffon & Maublanc (1911) studied a culture identical with the strain figured by Miehe, but expressed doubts as to whether the corresponding hyphomycete would be conspecific with the fungus proposed by Tsiklinsky. They argued the protologue was insufficient for a definite conclusion.

since from published informations, the fungus examined by Tsklinsky would have conidia definitely smaller than indicated by Miehé". Griffon & Maublanc then assigned their isolate to *Sepedonium* Link. on account of the slightly verrucose nature of the conidial wall.

In 1933, Mason examined a culture of *Acremonella thermophila* Curzi, "kindly supplied by Mr Curzi", he noticed the similarity with *Sepedonium lanuginosum* and also with *Acremonella* sp. Rege. As Mason had concluded before that *Acremonella* Sacc. was a synonym of the earlier *Monotospora* Corda (non *Monotospora* Vuill.), he proposed to rename the fungus of Griffon & Maublanc *Monotospora lanuginosa*. No mention of the binomial *Thermomyces lanuginosus* was made by Mason. In the meantime, Curzi (1930) published an extensive cultural study of his *Acremonella thermophila*, a fungus he had previously submitted to Griffon & Maublanc for examination (vide Mason, 1933).

While describing *Thermomyces stellatus* (= *Humicola stellata*), Bunce (1961) questioned the maintenance of the Griffon & Maublanc fungus in *Monotospora* Corda since [and as also stressed by Mason (1933, 1941)], the concept of this genus was still under debate. Bunce rather favoured the transfer of *Monotospora lanuginosa* to *Humicola* Traenck. established for mesophilic hyphomycetes sharing the same type of aleurospores. Cooney & Emerson (1964) followed Bunce proposal. On the other hand La Touche (1950) who had isolated this fungus from compost, considered the binomial *Thermomyces lanuginosus*. The latter name was also retained by Apinis (1963) on the basis all his isolates from alluvial soils agreed with the original description provided by Tsklinsky (1899).

The status of *Thermomyces* was finally definitively settled by Pugh *et al.* (1964) while describing the mesophilic *T. verrucosus*, in the latter a transverse septum is present just below the conidophore apex delimiting a small apical cell. According to these authors, such a feature is evident from Tsklinsky's photomicrographs and this provides arguments that her isolate is identical to the fungus now known as *Thermomyces lanuginosus*. Pugh *et al.* then provided a description and drawings for the latter based on the neotype strain (IMI 84460 (= ATCC 22070) isolated by Bunce from mouldy hay at the Rothamsted Experimental Station in 1959).

In the same year 1964, Cooney & Emerson in their treatment of thermophilic fungi underlined their first isolate of *Humicola* *Thermomyces lanuginosus* was strain No. 20 obtained in 1945 by D. G. Cooney from retting guayule shrub. This strain was later on numbered M 206522 at the University of California Herbarium, Berkeley (= ATCC 16455 (= CBS 632.91)). It provided the material for the description and drawings reported in their monograph. The isolate selected by Pugh *et al.* (1964) was explicitly designated "neotype", it has to be regarded as such against M 206522.

Subsequent taxonomic treatments of hyphomycetes uniformly accepted *Thermomyces* (Carmichael *et al.*, 1980). An ultrastructural study of the conidial ontogeny of its type species was conducted by Ellis D. H. (1981). Further, recently Straatsma and Samson (1993) re-examined isolate CBS 153.75 (= ATCC 28402) belonging to the unpublished *Humicola brevis* (Gillman & Abbott) Gillman var. *thermidea* Subrahmanyam, this was re-identified as *Thermomyces lanuginosus*. They also concluded the same for the similar unpublished taxon *Humicola brevisporea* Subrahmanyam & Thirumalachar based on CBS 152.75 (= ATCC 28403).

Humicola grisea var. *indica* (Subrahmanyam, 1983) was obtained as a laboratory contaminant at Pimpri, Poona. Trials to locate the representative strain were unsuccessful. According to the author, a critical study revealed "it belonged" to *Humicola grisea* (which produces intercalary chlamydospores) and approximates its var. *thermidea* (which also produces intercalary chlamydospores). However, the general lay-out of the drawings and

features underlined in the description clearly stress the proposed variety represents *Thermomyces lanuginosus*. The only deviation is the smooth character of the conidia in the proposed variety against the wrinkled conidial surface of *Thermomyces lanuginosus*.

Humicola lanuginosa var. *catenulata* (Morinaga *et al.*, 1986) was obtained in the course of a survey of soil borne thermophiles for high producer strains of lipase enzymes. Morphological details underlined in the publication clearly indicate it represents a deviant strain of *Thermomyces lanuginosus*.

The mesophilic *Thermomyces verrucosus* Pugh, Blakeman & Morgan-Jones (1964) displays no growth above 37 °C. It has globose, dark brown conidia with conspicuously warted surfaces, 10-17 µm wide. These structures are definitely larger than aleuriospores of the type species.

***Thermomyces stellatus* (Bunce) Apinis — Nova Hedwigia 5: 75. 1963.**

basionym *Humicola stellata* Bunce as '*stellatus*' Transactions of the British mycological Society 44: 372. 1961

Descriptions Bunce (1961); Apinis (1963); Ellis M. B. (1971).

The original material was isolated from mouldy hay in England and Wales. The fungus develops optimally at 40 °C with growth being very slow at 24 °C and not extending above 50 °C. Conidia of the aleuriospore type, angular, lobed, smooth, pale to mid brown or greyish brown, 5-10 × 5-9 µm. The transfer to *Thermomyces* is based on account of the absence of phialospores in culture and similarity in conidiogenesis.

THERMOPHILIC MYCELIA STERILIA

***Myriococcum thermophilum* (Fergus) van der Aa — Verhandelingen Koninklijke Nederlandse Akademie van Wetenschappen Afd. Natuurkunde, series II, 61: 60. 1973.**

basionym: *Papulaspora thermophila* Fergus — Mycologia 63: 426. 1971.

Descriptions: Fergus (1971); van der Aa (1973)

This 'bulbil-producing fungus' was described from mushroom compost in Switzerland (Fergus, 1971). *In vitro*, such structures appear very rapidly at 45 °C in the aerial and submerged mycelium. They are white at first, then yellow and finally orange at maturity, in mature bulbils, cells of the outer layers are narrower and more elongate than corresponding internal more intensely colored globose cells. The fungus shows no growth at 28 °C and 53 °C with the optimum being at 45 °C. No connection with a perfect stage yet established.

From seedlings of a *Begonia* species heavily infected with small sclerotia, van der Aa (1973) isolated a fungus matching the description of the type of *Myriococcum* Fr., *M. praecox* Fr. Subsequent comparison with representative strains of *Papulaspora byssina* Hotson confirmed similarity of both taxa. Also examination of the type of *Papulaspora thermophila* Fergus proved it to be congeneric with *Myriococcum praecox* except for

its thermophilic character. As *Miriococcum* predates *Papulaspora* Preuss, van der Aa transferred Fergus fungus to the former generic entity.

The term *bulbil* is now restricted to homogeneous pseudoparenchymatous bodies occurring only in the basidiomycetous genera *Burgoa* Goidanich and *Munimedusa* Weresub & LeClair. The term *papulaspore* is applied to thalloid propagules differentiated from the inception into central and sheathing cells (Weresub & LeClair, 1971). Such thalloid propagules occur amongst the mycelia of some species of *Melanospora* Corda and few probably related ascomycetes.

THERMOTOLERANT BASIDIOMYCETE

Phanerochaete chrysosporium Burdsall *apud* Burdsall & Eslyn — Mycotaxon 1: 124. 1974.

anamorph *Sporotrichum prunosum* Gilman & Abbott — Iowa State College Journal of Science 1: 306. 1927.

— *Chrysosporium prunosum* (Gilman & Abbott) Carmichael — Canadian Journal of Botany 40: 1166. 1962.

Emmensia brasiliensis Batista *et al* — Revista Facultad Medicina, Universidad de Ceara (Brazil) 3: 52. 1963.

— *Sporotrichum dehradunense* Sarbhoy & Saksena — Sydowia, Annales Mycologici, Ser II, 19: 198. 1966 ("1965").

= *Chrysosporium lignorum* Bergman & Nilsson — Department of Forestry, Proceedings of the Royal College of Forestry, Stockholm, Research Notes, 53: 28. 1966, (*nom. inval.*, Art. 36.1).

Sporotrichum pulverulentum Novobranova — Novosti sistematiki nizshikh rastenii 9: 184. 1972.

Description: Stalpers (1984)

Sporotrichum prunosum (also as *S. pulverulentum* and *Phanerochaete chrysosporium*) is a thermotolerant hyphomycete that has become the subject of many recent physiological studies. The fungus is known to produce three types of hydrolytic enzymes active in the degradation of cellulose. It is actually used as a model for the biodegradation of lignin and the production of protein from lignocellulosic waste material, a process designated single-cell protein (Stalpers, 1984). For these reasons, it is included in this study.

The protologue of the anamorph is based on a strain isolated from soil. The fungus was later on transferred to *Chrysosporium* Corda on account of the confusion surrounding the generic concept of *Sporotrichum* Link (Stalpers, 1978). As for the teleomorph, the first specimen was collected in the Sonoran Desert, Arizona (USA), when cultured, it produced a *Chrysosporium* state matching *Sporotrichum prunosum*. Later on the teleomorph could be obtained *in vitro* under particular cultural conditions. Reported cardinal temperatures are: minimum 7° C, optimum 36-40° C, maximum 46-49° C.

In culture *Sporotrichum prunosum* is the most variable species of the genus and such accounts for the several published synonymies. The similarity with *S. pulverulentum* has been the matter of a long debate (Burdsall, 1981) but recent studies provided

arguments in favour of such a synonymy (Stalpers, 1984). Citation in applied work of the teleomorphic name is favoured against the two commonly cited anamorphic binomials since several nomenclatural problems are still connected with the latter. The fungus has also been reported as a human pathogen being isolated from lungs and this explains its inclusion in the genus *Emmonsia* Ciferri & Montemartini.

TAXA OF UNCERTAIN POSITION

— *Mucor thermo-hyalospora* Subrahmanyam — *Bibliotheca Mycologica* 91: 421, 1983. (*nom. inval.*, Art. 37.1).

The examined strain was isolated from contaminated curd collected in the local market at Pimpri, Poona, India. The fungus is clearly thermophilic with growth starting at 24° C, being optimum at 45° C and maximum at 55° C. According to the author "careful study of monosporic cultures showed that it closely resembled *Mucor tauricus* Milko & Schkur but differs from it in being homothallic. Therefore it is described here as a new species".

The presence of weakly developed rhizoids in *Mucor tauricus*, accounted for its transfer to *Rhizomucor* by Schipper (1978). This information was however overlooked by Subrahmanyam (1983) who also provides no details concerning the presence or the absence of corresponding structures in his zygomycete. This taxon might simply represent a deviant strain of *Rhizomucor pusillus*.

Stilbella thermophila Fergus — *Mycologia* 56: 277, 1964.

This synnematus hyphomycete was first isolated from mushroom compost in Switzerland. Optimum growth is between 35-50° C, at 55° C, slight development still occurs but such is not the case below 25° C. In culture, the fungus produces white synnemata, up to 300 µm high bearing whitish glistening mucoid conidial heads, conidia are hyaline, continuous, oblong-ellipsoid, 15-17 × 6-10 µm.

Seifert (1985) in his monographic treatment of *Stilbella* Lindau re-examined authentic material. Conidia were observed to develop from percurrently proliferating conidiogenous cells, i.e. annellophores, a feature enhancing its exclusion from the genus. Additional work is undertaken to establish the correct taxonomic position of this species (Seifert, pers. comm.).

CONFUSING BINOMIALS

— *Achaetomium thermophilum* Basu — *Current Science* 51: 524, 1982.

The original living strain was isolated from leaf litter at Bhattni, Uttar Pradesh, India. It was described as being similar to *Achaetomium macrosporum* Rai, Wadham, & Tewari but differ by being "thermophilic" in nature, although no minimum growth temperature had been indicated.

Cannon (1986) examined a culture (IMI 292262) derived from the holotype. Growth and sporulation proved to be satisfactorily at 25° C indicating the fungus is rather thermotolerant. Ascospores produced were also found to be uniporate rather than biporate as stressed in the protologue and thus matching those of *Achaetomium macrosporum*. Based on these observations, Cannon concluded the ascomycete proposed by Basu is conspecific with *Achaetomium macrosporum*.

Species of *Achaetomium* Rai & Tewari are known to be good thermotolerants (von Arx *et al.*, 1988). The concept of the genus is however still under debate. Thus von Arx *et al.* (1988) excluded *A. macrosporum*, they also suggested the latter is rather similar to *Chaetomium vitellinum* Carter or *Ch. megasporum* Sörgel.

***Calcarisporiella thermophila* (Evans) de Hoog — Studies in Mycology 7: 68. 1974.**

basonym *Calcarisporium thermophile* Evans Transactions of the British mycological Society 57: 247. 1971.

This is the type species of the mucedinaceous genus *Calcarisporiella* de Hoog (de Hoog, 1974). The original living culture was isolated from coal spoil tips at Staffordshire, England. According to Evans (1971 a & b), the minimum growth temperature value is 16° C, optimum at 40° C and maximum at 50° C. The fungus should thus be considered a thermotolerant.

***Endoblastomyces thermophilus* Odinzowa — Microbiology, Moscow 16: 273. 1947 (description only); Die Systematik der Hefen: ?. 1960 (latin diagnosis but no type designated); (nom. inval., Arts. 36.1 & 37.1).**

This is the type species of *Endoblastomyces* Odinzowa. The original protologue was not accompanied by a latin diagnosis provided later on by Odinzowa in Kudryavtzev's book "Die Systematik der Hefen", the german translation of which was published in Berlin in 1960, however, Odinzowa then omitted to designate a holotype.

This yeast was isolated from brewing wort inoculated with baker's yeast in a bread factory in Central Asia, USSR, it was considered to represent a new thermophilic taxon. Carmo-Sousa (1970) was unable to locate the corresponding living strain and according to him, the original description strongly suggests similarity with *Trichosporon capitatum* Diddens & Lodder. This was substantiated by the arrangement of the pseudomycelium, endoblastospores formation and maximum temperature of growth being identical in both taxa.

Trichosporon capitatum is not thermophilic in the sense of Cooney & Emerson being able to develop below 20° C with a maximum at 44-46° C (Carmo-Sousa, 1970). The fungus was later on relocated in *Geotrichum* Link. Fr. and its perfect state discovered by de Hoog *et al.* (1980).

***Geotrichum candidum* Link var. *thermoideum* Qureschi & Mirza — Biologia, Lahore 27: 144. 1981.**

The original material was isolated from camel dung in Pakistan. The fungus was regarded by van Oorschot & de Hoog (1984) as a possible synonym of *Arthrographis sulfurea* (Grev. Fr.) Stalpers & van Oorschot, a mesophilic hyphomycete.

***Gilmaniella thermophila* Qureschi & Mirza — *Biologia*, Lahore 29: 341. 1983.**

The original material developed on goat dung collected in Pakistan. The species was overlooked by Sivanesan and Sutton (1985) while describing *Gilmaniella punctiformis* and also by Moustafa and Ezz-Eldin (1989) during their recent addition of *G. multiporosa*, isolated from Egyptian soils in North Sinai. These additions bring to five the number of known species.

Gilmaniella thermophila might be a later name of *G. macrospora* Moustafa, the final decision awaits comparison of authentic material. The latter was first encountered while investigating the mycoflora of salt-marsh soils of Kuwait. It was also subsequently recovered, although infrequently, from Iraqi soils analysed by Abdullah & Al-Bader (1990). The Iraqi strains developed optimal growth at 40°C with a maximum between 45-50°C thus confirming the thermotolerance abilities of *Gilmaniella macrospora*. The specific epithet refers to globose conidia being larger than conidia of *Gilmaniella humicola*, the type species, 14-18 µm versus 7-10 µm for the latter.

***Lagenidium thermophilum* Nakamura, M. Nakamura, Hatai & Zafran — *Mycoscience* 36: 400. 1996.**

The specific epithet coined for this newly described Oomycete is misleading. The fungus was found to infect the eggs and larvae of the mangrove crab, *Scylla serrata* Forsskal, in Bali, Indonesia. Isolated strains proved to represent a new species of *Lagenidium* Schenk having a unique discharge process. Growth range is from 15-45°C with the optimum being between 30-40°C. This taxon is thus a fast growing thermotolerant fungus.

***Melanomphalia thermophila* (Singer) Singer — *Atas*, Instituto de Micologia, Universidade de Recife 5: 482. 1963.**

basonym *Tubaria thermophila* Singer — *Papers of the Michigan Academy of Sciences, Arts and Letters* 32: 145. 1948

The type specimen of this basidiomycete was collected by the author in the state of Florida (USA) at Highland Hammock State Park (Singer F 20, F 20a, FH). The habitat in which the carpophore developed was specified as "In dumetis subtropicis humidissimis in terra humosa sabulosa vel nonnunquam nivalibus. Caryae megacarpae affixa vel e ligno mucido ecrecentes, aestate". The reason underlying the selection of the epithet *thermophila* seems to have simply been suggested by the very warm to hot humid weather prevailing in this southern state of the United States. The fungus is not a thermophile in the Cooney & Emerson's sense.

Similar cases concern *Russula roseipes* (Sacc.) Sacc. subsp. *thermophila* Singer, collected under *Pinus taeda* in North Florida, and *Stellus hirtellus* (Peck) Kuntze var. *thermophilus* (Singer) Smith & Thiers (Singer, 1975).

***Mucor thermophilus* Prakash & Sarbhoy — Zentralblatt für Mikrobiologie 148: 531. 1993.**

The specific epithet coined for this recently described zygomycete is misleading since "the species is able to grow and sporulate at 30° C and above 30° C" (Prakash & Sarbhoy, 1993), however, the minimum and maximum growth temperature values were not ascertained. Further the statement that "the specific epithet has been given on thermotolerant nature of the species" clearly indicates the fungus is not a thermophile in the Cooney & Emerson's sense.

***Paecilomyces puntonii* (Vuillemin) Nannfeldt *sensu* Eicker (1972):**

The correct binomial for this hyphomycete is *Paecilomyces puntonii* (Vuillemin) Nannizi (Samson, 1974).

Eicker (1972) isolated a hyphomycete from the faeces of domestic fowls in South Africa strain UP 71 T (University of Pretoria) he identified as *Paecilomyces puntonii*. This isolate "did not grow at 20° C, neither at 30° C but good growth took place at 50° C. No perfect state developed on any of the cultures media at the various temperatures of incubation used". *Paecilomyces puntonii* is a mesophilic fungus with optimum growth being at 25° C (Samson, 1974).

***Sporotrichum cellulophilum*:**

Durand *et al.* (1984) clearly specify this binomial correspond to a thermophilic fungus. Its ability to produce interesting enzymes of the cellulases and hemicellulases types were largely investigated by several workers (Kinoshita *et al.*, 1986). However, Stalpers in his 1984 revision of *Sporotrichum* makes no mention of this binomial in the check-list of epithets used in combination with the genus. Also publication of this binomial after this date following standard taxonomic rules could not be traced. It is thus evident that *Sporotrichum cellulophilum* has no taxonomic status.

With regard to literature on thermophilic fungi, the generic epithet *Sporotrichum* was first introduced by Apinis (1963) for a hyphomycete which ultimately will be renamed *Mycophthora thermophila*, this will also prove to be the anamorph of *Coryascus heterothallicus*. A second *Sporotrichum* made its appearance in the last decades in papers dealing with biotechnological work. *Sporotrichum prunosum*, anamorph of the basidiomycete *Phanerochaete chrysosporium*. This thermotolerant hyphomycete was described also under several *Sporotrichum* names (see comment under *Phanerochaete chrysosporium*). It is sometimes erroneously indicated as being thermophilic (Deshpande *et al.*, 1978).

The common use of "ghost binomials" in publications dealing with applied studies involving fungi is a source of serious confusion. A similar case is forwarded by the binomial *Acremonium cellulophilum* (Satyanarayana *et al.*, 1992). Such a practice should be definitely prohibited.

***Sordaria thermophila* Fields — Mycologia 60: 1117. 1968.**

The original strain of this ascomycete developed on cow dung collected in Texas (USA) and incubated in a moist chamber. According to the protologue "the specific epithet refers to a high temperature requirement for ascospore germination. Ascospores of the new species germinated less than 1% on media containing sodium acetate. With an additional treatment of 40-45° C for a period of 8-12 h, germination was increased to 40%."

In the published description no data is reported on the *in vitro* linear variation of growth with temperature. Also the conditions at which moist chambers were incubated are not specified (Fields, 1968). The thermophilic nature of this taxon thus cannot be ascertained. The selected specific epithet seems to relate to the heat treatment applied to enhance ascospore germination.

Guarro & von Arx (1987) regarded this heterothallic relative of *Sordaria fimicola* (Rob.) Ces. & de Not. as representing a good species. Further investigations are however required to underline the biological and taxonomical characteristics of this ascomycete which apparently has not been reported since its description.

***Zalerion thermophylli* Udaiyan — Journal of Economic and Taxonomic Botany 15: 664. 1991 (1992); (nom. inval., Art. 37.1).**

The original material developed on beech wood test blocks immersed in the cooling tower and the collecting lagoon of a hydroelectric plant at Tamil Nadu, holotype was not indicated.

The dematiaceous hyphomycete genus *Zalerion* Moore & Meyers was established for a widely distributed mesophilic fungus trapped on wood blocks immersed in sea-water, *Z. maritima* (Linder) Anastasiou, described before under several names (Ellis M. B., 1976). *Zalerion thermophylli* is most probably identical to the type species. The epithet *thermophylli* must have been suggested by the high temperature of the water circulating in the cooling tower.

DISCUSSION

Thermophilic fungi dealt with in this contribution are found to form a small group of less than forty species and varieties. Growth at high temperatures is thus definitely a rare feature among fungi. Also several of these taxa were described in recent years. This ecological group is thus expected to expand rapidly in the near future, in particular if some credit is awarded to the estimate amounting the number of existing species to one million and half. A major emphasis for this trend is also embodied by the outcome of taxonomic work conducted in the last decades. Such achievements have provided adequate answers for long standing problems. A limited additional work of this type is still necessary to solve remaining minor ones.

Taxa treated here are considered as strict thermophiles based on the definition of thermophilism provided by Cooney & Emerson (1964). However the use of this simple classificatory system to segregate between thermophilic and thermotolerants is sometimes difficult to apply, this is particularly critical at the lower temperature threshold of 20 °C. Thus following Bokhary *et al.* (1984), the well established thermophile *Melanocarpus albomyces* should be regarded a thermotolerant being able to grow below 20 °C. It is possible the response of different strains of the same taxon accounts for such deviations. Further difficulties in defining true thermophiles results from the absence of reliable growth curves covering a wide range of temperatures for most taxa proposed as such. This basic simple type of data is needed to ascertain the true nature of few members of this group.

Based on available informations, the ability to only develop at high temperatures is disclosed by few Mucorales, Eurotiales and Sphaerales (*sensu* von Arx, 1988) and by

several Hyphomycetes. No coelomycete and no basidiomycete was found to be thermophilic. Further the teleomorph of the sole thermophilic ascomycete, *Mycococcium thermophilum*, is hypothesized not to belong to Eurotiales on the assumption it should have made its appearance since the fungus was described. However observations relating to mating experiments of this sterile fungus are uncommon in the literature.

The group of live thermophilic Mucorales comprise the still monospecific *Thermomucor* and several Rhizomucors including the type, the former differs mainly by having smooth zygospores, a character uncommon in the *Mucoraceae*. Also although regular production of zygospores by *Rhizomucor mitchellii* should prevent confusion with *Rh. pusillus*, the type species, the ecology of each taxon is not yet clearly understood. Further, the validity of both *Rhizomucor tauricus* and *Rh. namitatusensis* is questioned.

The group of ascomycetous fungi brings together twenty species and three varieties, these relate to only nine genera. Following von Arx (1987-1988), *Dactylomyces* (inclusive of *Thermosascus*) and *Talaromyces* belong to Family *Onygenaceae*, Order Eurotiales, as *Chaetomium* was established for taxa previously assigned in *Dactylomyces* and *Thermosascus*, then the new genus should also be accommodated in this family. The remaining six genera are representatives of Families *Chaetomiaceae* (*Chaetomium*), *Microascusaceae* (*Canariomyces*) and *Thielaviaceae* (*Corynascus*, *Melanocarpus* and *Thielavia*).

Dactylomyces appears monospecific. *Canariomyces* and *Thermosascus* have one species each plus one variety for the latter, new informations about the variety might lead to the proposal of a specific rank. *Corynascus* and *Melanocarpus* have two species each, this number becomes three in the case of *Chaetomium*, *Talaromyces* and *Thielavia*. *Chaetomium* is represented by four species and two varieties but *Ch. britannicum* might prove not to be a true thermophile when a living culture becomes available. Also definite taxonomic decisions about *Chaetomium thermophilum*, its varieties and *Ch. virginicum* might reduce this group to only two accepted species, the genus would thus comprise only three thermophiles.

Not all thermophilic ascomycetes have an associated anamorphic state, also among these conidial states some do not develop concomitantly with the corresponding teleomorph. *Canariomyces thermophilus* has no anamorph although the type was described with a catenate conidial state. Thermophilic *Chaetomium* do not develop conidia of any kind. *Thermosascus* can be regarded as not having an anamorph producing catenate conidia. *Dactylomyces* has a distinctive but yet unnamed *Polypaecium* anamorph, the fungus remains not satisfactorily documented probably due to the taxonomic confusion with *Thermosascus* that prevailed. *Polypaecium* anamorphs also characterise *Dichotomomyces* Saito Saito having 2-3 described species (von Arx, 1981). The genus also has cleistothecia with a wall of *textura angularis* type but asci are produced in chains, it also belong to Family *Larotaceae* sensu von Arx (1987). *Melanocarpus* approximates *Canariomyces* since respective type species have distinctive conidial states but that of the latter is a mesophile. The former genus has now four taxa with the second thermophile, *M. thermophilus* not developing the characteristic arthroconidial state of the type, the same situation is disclosed by the two other members of the genus which do not develop at high temperatures.

Regarding the genus *Thielavia*, *Th. pingtungia* has no conidial state, a feature characteristic of all known Chaetomidia. *Th. australiensis* was reported with an anamorph of the *Trichosporiella* type, this is developed by other members of the genus (Mouchacca, 1973). The fungus is however badly documented being known only from the protologue. *Thielavia terrestris* is associated with a distinctive anamorphic state described exclusive of the teleomorph, *Acremonium alabamense*, due to the "homothallic with

cross-feeding' nature of the perfect state, the anamorph could be encountered alone in studies involving high incubation temperatures. *Thielavia* is still admitted to represent a heterogeneous entity due to lack of informations about the behaviour in culture of *Thielavia basicola* (type species) and production of hyaline and dark coloured colonies by known members.

On the other hand, thermophilic *Talaromyces* all develop a conidial state, these belong either to *Paeclomyces* (*T. hyssoclumydonides*) or to *Penicillium* (*T. emersoni* and *T. thermophilus*). Regarding *Paeclomyces*, von Arx (1987) suggested it be expanded to include *Penicillia* of Sections *Biverticillata* and *Sagenomella* known as anamorphs of genera he grouped in Family *Onygenaceae*, such a proposal was made to increase the degree of homogeneity among genera. A similar situation is disclosed by *Coenomeria* and *Corynascus*, all three taxa of the former have a well developed *Paeclomyces* state while both species of the latter have anamorphs now correctly assigned to *Myceliophthora*. Taxa of *Coenomeria* and *Corynascus* had very complicated respective taxonomic histories either due to cases of misidentification (species now placed in *Coenomeria*) or to the heterothallic nature of the *Corynascus* perfect states. For the latter, it follows that either *Myceliophthora* could develop singly in studies conducted on thermophile habitats with only appropriate mating leading to ascospore formation. The anamorph of *Corynascus heterothallicus* was proposed prior to the discovery of the teleomorph while the reverse is true for *C. thermophilus*.

The group of thermophilic hyphomycetes comprises thirteen species although for *Scytalidium thermophilum*, the term species appears inadequate in the present situation. These fungi belong to seven genera. *Acremonium*, *Malbranchea*, *Myceliophthora* and *Thermophymatospora* are mucedinaceous entities, while dematiaceous thermophiles belong to *Humicola*, *Scytalidium* and *Thermomyces*.

Among mucedinaceous taxa, three are established anamorphs of "almost heterothallic" ascomycetes and thus could be observed alone in studies involving a self-heating process. *Acremonium alabamense* is the conidial state of *Thielavia terrestris* whose mating behaviour is not yet clearly understood. This is not a typical *Acremonium* species and such accounts for its inclusion in a new section with phialidic states of some *Chaetomia*. Such is not the case for *Acremonium thermophilum*, a not yet well documented thermophile. The perfect state of *Myceliophthora fergusii* is *Corynascus thermophilus* and care should be taken to avoid confusion with the teleomorph of *M. thermophila*. *C. heterothallicus*, the third member *M. humildea* has not yet developed a perfect state, a situation analogous to the mesophilic type species of the genus, *Malbranchea cinnamomea* is a very distinctive colored arthroconidial fungus actually displaying a wide distribution, it is the sole thermophile of a genus known to comprise mesophiles associated with well defined teleomorphs (von Arx, 1987). *Thermophymatospora fibuligera* is unique with its septal clamp connections and an aleurosporic state, this peculiar fungus is apparently still known only from the type locality.

The remaining dematiaceous thermophiles were assigned to *Humicola*, *Scytalidium* and *Thermomyces*. But only the taxonomic status of the latter is now the subject of large consensus. *Thermomyces lanuginosus* is the first assessed thermophilic fungus. His complex nomenclatural history has involved genera as *Acremonella*, *Humicola*, *Monotospora* and *Scpedoniam*. The definite re-instatement of *Thermomyces* by Pugh *et al* (1964) clarified its links with thermophilic species of *Humicola*. The genus now also comprises *Th. ibadensis*, *Th. stellatus* and the mesophilic *Th. verrucosus*. However, only the type species is by far the most reported one.

The status of *Humicola hyalothermophila* needs to be re-assessed in conjunction with that of both *Humicola* recently proposed as synonyms of *Scytalidium thermophilum* (Straatsma & Samson, 1993). The introduction of *Scytalidium* in an attempt to relocate *Torula thermophila* added much confusion as the transfer was not substantiated by valid taxonomic arguments. This combination was however immediately reported by Ellis M. B. (1976). *Scytalidium lignicola*, type species of the genus, is a mesophile producing cultures with scanty aerial mycelium. The fungus develops conidogenous cells of two kinds: hyaline fertile hyphae become septate, later producing thin-walled arthroconidia by fragmentation; brown fertile hyphae forming chains of brown arthroconidia, also chains of brown aleuriospores could be observed (Ellis M. B., 1976). These intercalary conidia develop by transformation of pre-existing normal hyphal cells. The presence of solitary conidia terminal or lateral was never reported. In *Scytalidium indonesicum* chlamydospore formation and disarticulation follow the same pattern but no hyaline arthroconidia develops. Here too, terminal or lateral solitary conidia were not reported (Hedger *et al.*, 1982).

In species of *Humicola* solitary terminal and more commonly lateral aleuriospores usually develop in addition to intercalary morphologically similar ones. Single terminal aleuriospores (and less often lateral ones) may become intercalary by hyphal extension of their tip. Also chains of aerial or immersed "aleuriospores" do not disarticulate to liberate individual elements but such is achieved by lysis of sustaining hyphal cells. Further, no hyaline arthroconidia are produced by any described member of the genus but mesophile taxa rather produces hyaline phialospores. Straatsma and Samson (1993) compared a large number of strains assigned to *Scytalidium thermophilum*, *Torula thermophila* or to both *Humicola* now regarded as synonyms of the former. They underlined two extreme types could be recognized, the first having simple very dark spores borne on short lateral branches matching the description of *Humicola grisea* var. *thermidea*, the second type develop intercalary slightly pigmented spores in chains, representatives of *Scytalidium thermophilum* or more appropriately of its basonym *Torula thermophila*. Within the two types however, some isolates also develop short terminal chains of conidia making them intermediate between types 1 and 2.

Straatsma and Samson (1993) stress such intermediate isolates favours not the segregation of taxa "on the basis of the single character of conidia in the aerial mycelium": these rather support grouping of all types under one binomial whose placement in *Scytalidium* is to be reconsidered. As the particular mode of chlamydospore formation in this genus deviates from the pattern depicting species of *Humicola*, the exclusion of the above complex from the former is more than justified. *Scytalidium* is now regarded a heterogeneous entity due to addition of species only developing dematiaceous "arthroconidia". Nevertheless, extension of *Scytalidium* characteristics for the understanding of *Humicola* species has shadowed features proper to the latter preventing sound taxonomic separation among its members.

From an ecological point of view, the equivocal application of the now widely accepted (inspite of its limitations) definitions of Cooney & Emerson (1964) lead to consider well established thermotolerants as thermophiles. Ellis D. H. (1981) regards all *Rhizopus* able to grow at 45°C as thermophilic although they display growth below 20°C. These zygomycetes and some other true mesophilic fungi are also currently considered as thermophiles in publications focusing on biotechnological problems (Satyanarayana *et al.*, 1992). Several authors also classify as thermophile all fungi developing in isolation plates incubated at 45°C (Abdel Fattah *et al.*, 1977; Moubasher *et al.*, 1988).

Another type of misleading situations relates to epithets selected while describing a new taxon found to develop at elevated temperatures. The recent *Mucor thermophilus* is a good example among others here considered, from the protologue it is evident this *Mucor* should be regarded as a thermotolerant. A definitely critical situation is exemplified by the frequent use in studies involving fungal enzymes of ghost binomials having no taxonomic status of any kind as *Sporotrichum cellulophilum*, such a practice needs to be totally banished for the confusion it introduces, in particular while attempting to analyse published data (Satyanarayana *et al.*, 1992, Schekkar & Johri, 1992).

Strict restriction to nomenclatural rules governing citations of fungal binomes is fundamental. Authors of applied research dealing with thermophiles should necessarily follow such regulations in order to stabilize names used in produced articles. This would bring an end to the chaotic state prevailing especially in publications relating to fungal ecology and biotechnology. The taxonomic and nomenclatural reappraisal of known thermophilic taxa here undertaken will definitely unravel informations already available. This should enable a sound synthesis of published data and foster the discovery of new elements of this interesting physiological group of fungi.

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PHÉNOMÈNES D'ADAPTATION PARASITAIRE DES CHAMPIGNONS KÉRATINOPHILES TELLURIQUES ET CONSÉQUENCES EN PATHOLOGIE HUMAINE ET ANIMALE

Dominique CHABASSE

Laboratoire de Parasitologie — Mycologie
CHU — 4 rue Larrey 49100 ANGERS — FRANCE

RÉSUMÉ Les champignons keratinophiles constituent une flore importante et variée caractérisée par sa capacité à se développer au dépens de la kératine, protéine complexe entrant dans la composition de la peau et des phanères de l'homme et des animaux. Ils appartiennent pour la plupart à la classe des Ascomycètes, et sont positionnés depuis 1985 dans l'ordre des Onygenales au sein de deux familles distinctes les *Arthrodermataceae* et les *Onygenaceae*. Si beaucoup d'entre eux restent des saprophytes du sol, un certain nombre comme les Dermatophytes et des espèces assimilées (*Aphanogmus*) évoluent vers un parasitisme d'abord facultatif puis quasi obligatoire chez l'homme et ou l'animal. Le cheminement soit animal-homme semble être l'évolution phylogénétique habituelle de ces champignons. Les conséquences pratiques de cette adaptation au parasitisme sont : Sur le plan biologique, une perte progressive de leur mode de reproduction saprophytique (stade téléomorphe puis anamorphe), avec en revanche un développement des structures de diffusion propres au parasitisme (parasitisme pileux et arthrospores virulentes dans les cornéocytes) qui assurent la dissémination du champignon. Sur le plan épidémiologique, il est intéressant de constater que les espèces géophiles ou telluriques (*Microsporum gypseum*, *Aphanogmus fulvescens*) et parfois zoophiles (*Trichophyton verrucosum*) sont à l'origine d'une intense réaction inflammatoire chez l'homme. À l'opposé, les espèces anthropophiles (*Microsporum audouinii*, *Trichophyton violaceum*, *Trichophyton rubrum*) mieux adaptées à l'homme évoluent sur un mode chronique avec des réactions de réjets limitées ou nulles. Les champignons keratinophiles représentent un mode d'étude intéressant d'adaptation au parasitisme en mycologie médicale.

ABSTRACT Keratinophilic fungi form an important and varied flora distinguished by the ability to develop on an ideal substrate, keratin, a complex protein forming part of the composition of the skin, nails and hair of men and animals. These fungi belong to the Ascomycetes and are placed in the order Onygenales at the center of two distinct families, the *Arthrodermataceae* and the *Onygenaceae*. If most of these species remain saprophytes of the soil, a certain number among them as the Dermatophytes and related fungi evolve in the direction of an optional parasitic state on animals while others go resolutely toward an almost obligatory parasitism. The pathway from soil to animal to man is the usual phylogenetic evolution of these fungi. The practical consequence of the parasitic adaptation implies at least two observations. A progressive loss of the saprophytic reproduction of these species. This is shown by the impossibility of producing the teleomorphic state in the anthropophilic fungi and by the difficulty of obtaining a good conidiogenesis (anamorphic state). Geophilic species known as dermatophytes like *Microsporum gypseum* or closely related like *Apha-*

naevius fulvescens and the zoophilic dermatophytes like *Trichophyton verrucosum* produce highly inflammatory lesions with a tendency to spontaneous cure. On the other hand anthropophilic dermatophytes like *Trichophyton mentagrophytes* or *Microporum audouinii* produce less inflammatory and more chronic lesions. Keratinophilic fungi represent the ideal model of parasitic adaptation in medical mycology.

MOTS CLES Champignons keratinophiles telluriques, Dermatophytes, *Chrysosporium*, Saprophytisme, Parasitisme, Evolution phylogénique.

KEY WORDS Keratinophilic fungi, Dermatophytes, *Chrysosporium*, Saprophytism, Parasitic adaptation, Phylogenetic evolution

INTRODUCTION

Les micromycètes keratinophiles telluriques sont des champignons isolés du sol (Chabasse *et al.*, 1985, 1987, 1988, 1989) et souvent sur des animaux (petits mammifères sauvages (rongeurs, insectivores) et autres fouisseurs (Chabasse *et al.*, 1987, Houin *et al.*, 1972, Otčenasek & Dvorak, 1962), ainsi que de certains oiseaux (Chabasse & Guiguen, 1986).

Leur caractéristique commune est d'être keratinophiles, c'est-à-dire qu'ils privilégient un substrat nutritif particulier : la kératine. Cette kératine tellurique appelée « kératine morte » est issue des mammifères (poils, fragments de cornes ou de sabots...), des oiseaux (plumes...) et des carapaces d'insectes mélangées au sol. L'origine tellurique de ces espèces est connue depuis les travaux de Sabouraud (1910) à la fin du siècle dernier. Plus tard le hongrois Szathmary (1940) isola un dermatophyte géophile appelé *Trichophyton terrestre gyratum*, puis *Trichophyton fluviatile*, vraisemblablement le *Trichophyton mentagrophytes* d'aujourd'hui. Le comportement keratinophile de ces champignons fut rapidement utilisé pour les isoler. C'est ainsi qu'à partir des constatations de Toma (1929) sur le parasitisme des cheveux par les agents de teignes et les observations de Karling (1946) qui utilisait des cheveux humains pour piéger dans le sol d'un cimetière un keratinophile appelé *Rhizophydium keratinophilum*, Vanbreuseghem (1952, 1960) mit au point une technique originale pour extraire ces champignons du sol.

Cette méthode consiste à déposer des fragments de cheveux stériles sur de la terre placée dans des boîtes de Petri. Elle fut appelée par Benedek (1962) « the To-Ka Va » hair baiting méthode, ou technique de piégeage sur cheveux de Toma-Karling. Vanbreuseghem, rappelant le rôle respectif de chacun des auteurs dans cette découverte. Cette technique communément appelée technique de Vanbreuseghem fut à l'origine de très nombreux travaux réalisés dans le monde entier confirmant l'hypothèse de départ c'est-à-dire la vie saprophytique des dermatophytes et leur origine tellurique (De Vroey, 1968, 1970, Heliou *et al.*, 1965). C'est ainsi que *Trichophyton ajelloi* (= *Keratinomyces ajelloi*) a été découvert et décrit pour la première fois par Vanbreuseghem en 1952. De même Ajello (1953) isola à son tour *Microporum gypseum* puis *Microporum cooki* et Durie et Frey (1950) mirent en évidence : *Trichophyton terrestre*.

Cette technique a permis aussi de révéler la reproduction sexuée de ces dermatophytes géophiles et de décrire des espèces proches mais appartenant à des genres différents : *Chrysosporium*, *Geomyces* et *Malbranchea* (Carmichael, 1962, Currah, 1985, van Oorschot, 1980).

ÉVOLUTION DE LA TAXIMONIE

La plupart des micromycètes kératinophiles telluriques appartiennent à la classe des Ascomycètes, à la sous classe des Euascomycètes, comprenant elle-même 2 grandes familles : les *Gymnascaceae* et les *Onygenaceae*.

En 1973, Fennel inclut ces deux familles dans l'ordre des Eurotiales, mais Benny et Kimbrough en 1980 proposèrent une nouvelle distribution créant et plaçant ces espèces dans l'ordre des Onygenales. Currah proposa à son tour en 1985 une révision complète de cet ordre ou l'on retrouve les espèces les plus adaptées au parasitisme humain.

Currah ne retient plus comme élément déterminant la morphologie des ascocarpes (gymnothèce, cleistothèce, perithèce...) mais l'aspect des ascospores (lisses ou ornementées) associé à des critères physiologiques, c'est-à-dire l'aptitude à dégrader ou non la kératine. Ainsi il définit 4 familles dont deux ayant une affinité pour la kératine, les *Arthrodermataceae* et les *Onygenaceae*. Les deux autres familles ont soit un substrat nutritif plus varié (*Gymnascaceae*), soit une préférence pour la cellulose (*Microtrichaceae*) (fig. 1).

La comparaison des séquences nucléodiques de la sous-unité 18S ribosomiale de nombreuses espèces appartenant à l'ordre des Onygenales a permis de confirmer la plupart du temps la cohésion de cette classification (Bowman *et al.*, 1992, Leclerc *et al.*, 1994). Un grand nombre d'espèces adaptées au parasitisme appartiennent à l'ordre des Onygenales : *Trichophyton*, *Microsporium*, *Histoplasma*, *Blastomyces*, *Coccidioides*.

DU SAPROPHYTISME AU PARASITISME

Microsporium gypseum est un dermatophyte géophile menant essentiellement une vie saprophytique à partir de substrats kératiniques (kératine morte) comme les poils d'animaux, des fragments de cornes de bœuf ou de sabots de chevaux (Ajello, 1953, Badillet, 1982, 1991, Chmel & Vollekova, 1981). Il possède cependant de réelles capacités au parasitisme si l'on en juge d'après les lésions qu'il engendre chez l'homme et l'animal (Chabasse, 1991). Il en est de même pour un *Chrysosporium*, dont le téléomorphe, *Aphanoascus fulvescens*, est incriminé, lui aussi, dans des lésions « dermatophytic like » (Albala *et al.*, 1982, Chabasse *et al.*, 1989, Gueho *et al.*, 1985, Rippon *et al.*, 1970, Todaro *et al.*, 1984 ; Vanbeuseghem & Devroey, 1980).

Le cheminement du sol à l'animal puis de l'animal à l'homme serait l'évolution classique de ces micromycètes, selon Novac & Galoczy, (1966), Chmel & Vollekova (1981, et Dei Cas & Vernes (1986). Le kératinophile tellurique, saprophyte du sol, végéterait d'abord sur la kératine du sol (kératine morte), puis prépare à ce substrat sélectif, il passerait aisément sur le poil de l'animal (kératine vivante) ou directement chez l'homme. L'évolution de ces espèces géophiles en zoophiles puis anthropophile, serait due à la fois à des phénomènes extrinsèques (rencontre avec l'hôte) et intrinsèques (facteurs de reconnaissance, de virulence...), propres au champignon lui-même.


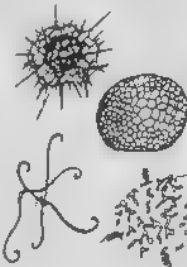





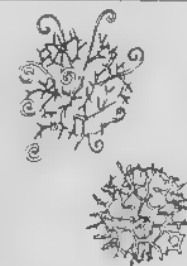




TABLE 3 FAMILIES OF THE ONYGENALES				
Family	Aeciospores	Peridia and Appendages	Anamorphs	Substrate and Habitats
ONYGENACEAE	 <p>pitted; spherical, oblate, obovoid.</p>		 <p>CHRYSOPORIUM MALBRANCHEA SPORENDONEMA</p>	<p>composts dung soil enriched with keratin or dung</p> <p>KERATIN*</p>
ARTHRODERMATACEAE	 <p>smooth, oblate to obovoid-discoid or obovoid-conical.</p>		 <p>CHRYSOPORIUM MICROSPORIUM TRICHOPHYTON</p>	<p>decaying hoof, horn, feathers, hair and skin some are parasitic on animals</p>
MYXOTRICHACEAE	 <p>smooth or striate; fusiform, ellipsoidal.</p>		 <p>GEOMYCES MALBRANCHEA DIDIODENDRON</p>	<p>processed or decaying plant materials</p> <p>CELLULOSE*</p> <p>paper, straw soil around the roots of plants</p>
GYMNOASCACEAE	 <p>smooth or slightly irregular ("lumpy"); often with polar and/or equatorial thickenings.</p>		 <p>absent or of un-named arthroconidia.</p>	<p>decaying vegetation soil rich in organic matter</p> <p>VARIABLE*</p> <p>various types of dung</p>

Figure — Les Onygenales
d'après Currah, *Mycotaxon*, 1985

CONSÉQUENCES ÉPIDÉMIOLOGIQUES, CLINIQUES ET BIOLOGIQUES

Épidémiologiques :

Les géophiles contaminent accidentellement l'homme soit directement (souillure traumatisme) soit par l'intermédiaire d'un animal relais (chien de chasse). Il n'y a pas de contamination inter humaine (Chermette, 1991 ; Garg *et al.*, 1985).

Les zoophiles (et geo zoophiles) passent aussi chez l'homme soit directement par contact avec un animal contaminé (le plus souvent chien, chat...) soit à partir de ses poils contenant la forme virulente du champignon disséminée dans l'environnement humain (tapis, fauteuil, couverture...) (Caretta *et al.*, 1989 ; Chabasse *et al.*, 1991 ; Chermette, 1991 ; Woodyer, 1977).

La diffusion de ces espèces à l'homme dépendra de leur spécificité vis à vis de leur hôte. Elle sera étroite avec *Trichophyton equinum*, *T. gallinae* respectivement adaptés au cheval et aux oiseaux de basse-cour, intermédiaire avec *Trichophyton verrucosum* parasite privilégié des bovins et des ovins et lâche avec *Microsporum canis* qui affectionne de nombreux animaux familiers et d'élevage. La contamination dépendra également des possibilités de contact avec l'animal-hôte. C'est ainsi que le chat, encore plus souvent que le chien, porteur asymptomatique de spores virulentes est à l'origine de bien des contaminations humaines. À l'inverse si un animal peut contaminer plusieurs personnes d'une même fratrie le passage inter humain est très difficile (Badillet, 1991).

Les anthropophiles adaptés au parasitisme humain diffusent bien dans la population. Les teignes dues à *Trichophyton violaceum*, *Trichophyton soudanense*, et *Microsporum langeroni* sont de ce fait très contagieuses (Badillet, 1991).

De même les squames issues de la peau, ou les fragments d'ongles des nageurs ou des judokas qui souillent les tapis de sport, les carrelages des douches, des vestiaires du pourtour des bassins de natation, sont à l'origine de nombreuses dermatophytoses du pied dues à *Trichophyton rubrum* et à *T. mentagrophytes* variété *interdigitale* (Detandt & Nolard, 1988 ; Chabasse *et al.*, 1995).

Cliniques :

Il est habituel de dire que plus une espèce fongique est adaptée à son hôte, mieux elle est supportée par ce dernier. Les dermatophytes anthropophiles bien que très engagés dans la voie du parasitisme sont à l'origine de lésions habituellement sèches, squameuses et évoluant sur un mode chronique (*Trichophyton rubrum*). À l'opposé les espèces géophiles ou geo-zoophiles sont génératrices de réactions bruyantes et inflammatoires. Elles peuvent d'ailleurs guérir spontanément tellement le rejet par l'hôte peut être important. C'est le cas de *Trichophyton verrucosum* infestant les bovidés qui provoque chez l'homme des sycosis de la barbe et de la moustache voire un kérion de cuir chevelu. Le traitement à base de corticoïdes peut parfois suffire à guérir ces patients.

D'autres zoophiles, plus largement répandus dans le monde animal, comme *Microsporum canis* sont en revanche à l'origine de lésions plus ou moins inflammatoires. En général les réactions de l'hôte vis à vis des micromycètes keratinophiles sont assez bien corrélées avec le degré d'adaptation au parasitisme.

Biologiques :

Les micromycètes keratinophiles issus du sol sont à l'origine en primo culture (milieu de Sabouraud) d'une grande abondance de spores. La conidiogenèse très importante explique l'aspect de ces cultures volontiers poudreuses. À l'opposé les espèces anthropophiles (*T. violaceum*, *T. soudanense*, *T. schoenleini*) ont une conidiogenèse de mauvaise qualité. Il n'est pas toujours aisé de retrouver des macro- ou microconidies caractéristiques du genre. Cependant cette règle n'est pas toujours vérifiée chez les anthropophiles où certaines souches de *T. rubrum* et *T. tonsurans* par exemple sont parfois riches en spores.

Chez les zoophiles, la morphologie microscopique est variable, certaines espèces très inféodées à un hôte comme *T. verrucosum* sont habituellement pauvres en spores sur les milieux usités en mycologie. À l'inverse *M. canis*, de diffusion large au niveau des animaux d'élevages et de rente, est plus riche en organes de fructifications.

Ce que l'on constate avec les stades anamorphes se vérifie encore mieux avec les stades téléomorphes ou sexués.

La plupart des keratinophiles issus du sol sont hétérothalliques avec un équilibre entre les souches plus (+) et moins (-).

La méthode de Vanbreuseghem très utilisée pour visualiser la reproduction sexuée des *Arthrodermataceae* et des *Ongophoraceae* illustre bien ce phénomène. Les espèces qui ont choisi le parasitisme vont perdre leur potentiel sexué. Quand on les fait réagir avec *Arthroderma* *vinii* on s'aperçoit qu'il y a sélection d'un seul type sexué, soit de signe (+) (cas du *T. montagnoli*, les var. *interdigitale*), soit de signe (-) (cas du *T. rubrum*). Déjà chez de nombreuses espèces zoophiles il existe un net déséquilibre entre les souches (-) et les souches (+). On ne connaît le stade sexué de *M. canis* (*Nannizzia otus*) que grâce à l'isolement d'une souche négative dans un élevage polonais, toutes les autres sont désignées (+). Les agents de teignes anthropophiles confrontés avec des souches d'*Arthroderma* sont de signe (+) et de signes (-) ne montrent pas de gymnothécées fertiles contenant des ascospores, mais tout au plus quelques ébauches de réaction sexuée.

La perte de la reproduction sexuée lors de l'adaptation parasitaire est-elle la conséquence du parasitisme ou est-ce cette perte qui a provoqué le parasitisme ?

On pourrait imaginer que le champignon face à un environnement hostile non favorable à sa reproduction sexuée s'adapte au parasitisme pour assurer sa survie. Au contraire la vie parasitaire a bien pu avoir comme conséquence une perte de certains de ses métabolismes et voies enzymatiques conduisant à appauvrir et à stériliser sa reproduction de type sexuée d'abord, asexuée ensuite. Il est vraisemblable que les espèces adaptées au parasitisme orientent leur potentiel enzymatique aux exigences de leur nouvel état (Chabasse & Contect-Audonnet, 1994).

CONCLUSION

Ainsi, si l'on retient comme hypothèse une évolution phylogénique des keratinophiles telluriques vers le parasitisme animal ou humain, il convient de rester attentif vis-à-vis de micromycètes isolés aujourd'hui encore de façon exceptionnelle sur l'homme (*Chrysosporium* sp.) mais pouvant devenir demain de véritables agents de mycoses humaines et animales et ceci en dehors d'un contexte d'immunosuppression.

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NUEVOS DATOS SOBRE *TRAMETES JUNIPERICOLA* MANJÓN, MORENO & RYVARDEN

M. N. BLANCO, J. CHECA y G. MORENO

Dpto. de Biología Vegetal, Universidad de Alcalá, Alcalá de Henares, 28871 Madrid, España

RESUMEN Se estudia el comportamiento en cultivo de *Trametes junpericola* y se compara con *Fomitopsis iberica*, especie muy parecida macro y microscópicamente, resultando ambas especies diferentes. *Trametes junpericola* posee podredumbre blanca, diferente de la de *Fomitopsis iberica*, que posee podredumbre parda.

Hasta el momento actual *Trametes junpericola* se comporta como una especie endémica de los sabinares de Guadalajara (España).

RESUME Le comportement en culture du *Trametes junpericola* a été étudié et comparé à *Fomitopsis iberica*, espèce très proche macro- et microscopiquement. Il en résulte qu'il s'agit de deux espèces différentes, *Trametes junpericola* possédant un mycélium blanc, différent de celui du *Fomitopsis iberica*, qui est brun.

Jusqu'à aujourd'hui *Trametes junpericola* est considérée comme espèce endémique des "sabinars" de Guadalajara (Espagne).

ABSTRACT We study the conduct of *Trametes junpericola* in culture and we compare it to *Fomitopsis iberica*, which is a species macro and microscopically very similar. Both species tending to be different species. *Trametes junpericola* has got white rot, which is different from the rot of *Fomitopsis iberica*, which has brown rot.

Trametes junpericola is still not known outside the type locality and must be looked upon as a relict of a previously larger distribution.

KEY WORDS . *Trametes junpericola*, culture, chorology, taxonomy

INTRODUCCIÓN

La provincia de Guadalajara en España tiene un gran interés botánico y micológico, porque conserva grandes extensiones de bosques muy bien conservados de *Juniperus thurifera* L., tanto en terreno ácido, como en terreno básico donde son más abundantes, en este caso a veces mezclados con *Quercus ilex* subsp. *ballota* (Desf.) Samp.

Desde el punto de vista micológico, y aunque no aparecen elementos ectomicorizógenos, hay una alta selectividad de especies parásitas y saprofitas características y de distribución ibero-norteafricana. A este respecto señalamos *Lenzites oxycedri* Malençon & Bertault (Manjón & Moreno, 1981), *Mycenella murganifera* (Maire in Kuhnert)



Foto 1. — *Trametes junipericola*, cuerpos fructíferos.

Maas G. (Moreno & Manjón, 1986), *Trametes junipericola* Manjón, Moreno & Ryvarden (Manjón, Moreno & Ryvarden, 1984) y *Acronephatium junipericola* G. Moreno & Heykoop, (Moreno & Heykoop, 1996)

La descripción de *Fomitopsis thurica* (Melo & Ryvarden, 1989) y la posterior publicación de una fotografía en color de dicha especie (Bernicchia, 1990), nos llamó la atención por su semejanza con *Trametes junipericola*, ambas especies poseen un gran parecido macro y microscópico y fructifican en coníferas mediterráneas. Se diferencian por el tipo de podredumbre que es blanca en el primero y parda en el segundo, característica que separa principalmente los dos géneros.

Posteriormente Ryvarden & Gilbertson (1993) recopilan e iconografían estos taxones, indicando la posibilidad de que *Trametes junipericola* presente podredumbre blanca.

Estos motivos nos impulsaron a realizar pruebas enzimáticas en cultivo puro de *Trametes junipericola* y comparar estas con la de *Fomitopsis thurica*, para asegurar la posición adecuada de estas especies en los géneros en los que se habían descrito. Se ha hecho especial hincapié en la prueba de la lacasa, ya que esta actividad es la que pone de manifiesto el tipo de podredumbre.

***Trametes junipericola* Manjón, Moreno & Ryvarden, Bol. Soc. Micol. Castellana 8: 47. 1984.**

Indicamos a continuación la diagnosis latina de *Trametes junipericola* porque esta especie se publicó en una revista, en aquella época con escasa difusión internacional, y que actualmente está agotada y es por lo tanto difícil de conseguir.

Diagnosis: *Fructificatio sessilis 4-10 cm, pileus cinereus et luridus, zonatus, velutinus et hispidus, pori facie cinerei et ochracei angulati, 2-3 per mm contextus albus et cinereus. Systema hypharum trimiticum hyphae generativae fibulatae hyphae skeletales et hyphae skeletico connectivae hyalinae, sporae cylindricae hyaline non-amyloideae 8-10 x 2,5-3 µm.*

Holotypus España, Guadalajara, Ermita de los Enebrales de Tamajón 30I VL7941, 5-V-1983, en troncos de *Juniperus thurifera* leg. G. Moreno, AH 5739, *isotypus* en MA-Fungi, en herbario 0 y en el herbario privado de H. Jahn.

Material estudiado *Trametes junipericola* España, Guadalajara, Tamajón, Ermita de los Enebrales 30I VL7941 Tronco de *Juniperus thurifera*, leg. R. Galán & G. Moreno 23-II-1982, AH 5237, *Ibidem* leg. J. Checa, R. Galán & G. Moreno 18-II-1983, AH 5740 *Ibidem*, leg. F. Horák, G. Moreno & M. Lizarra, 11 XII 1995, AH 18382

Fomitopsis ibérica Portugal, Ribatejo, Coruche, *Pinus pinaster*, leg. I. Melo, M. Correia & J. Cardoso, 27-XI-1986 LISU 3145 *Paratypus* Ribatejo, Salvaterra de Magos, Foros de Salvaterra, *Pinus pinaster*, em soca queimada, leg. I. Melo & J. Cardoso, 15-XII-1992, O

CULTIVOS

Se ha realizado el cultivo de *Trametes junipericola* en MEA (2° extracto de agar-malta), a partir de cuerpos fructíferos obtenidos de material fresco

Resultados obtenidos según los Códigos de Stalpers (1978):

1, 3, 7, (11), 13, (14), 16, (17), (18), 19, 22, 30, 36, 37, 39, 45, 46, 53, 60, (61), 75, (78), (84), 85, 90.

Actividades enzimáticas: La prueba de actividad lacasa se ha llevado a cabo con siringaldazina, alfa-Naftol, guayacol y benzidina, la actividad tirosinasa, con p-cresol y tirosina, la actividad peroxidasa con pirogalol, la actividad fosfatasa, con alfa-naftol fosfato y la actividad esterasa, con alfa-naftol acetato, observando los cultivos a las 0, 3 y 24 h. Los resultados se ven reflejados en la tabla 1

	0 h	3 h	24 h
LACASA			
Siringaldazina	+	-	-
alfa-Naftol	-	+	+
Guayacol	-	+	+
Benzidina	-	+	+
TIROSINASA			
p-Cresol	-	-	-
Tirosina	-	-	-
PEROXIDASA	-	+	+
FOSFATASA	-	+	+
ESTERASA	-	+	+

Tabla 1 · Actividades enzimáticas de *Trametes junipericola*

Asimismo, se han realizado cultivos y la actividad lacasa de *Fomitopsis iberica* a partir de material de herbario, siguiendo la misma metodología con los resultados reflejados en la tabla 2

	0 h	3 h	24 h
LACASA	-	-	-
Siringaldazina	-	-	-
alfa-Naftol	-	-	-
Benzidina	-	-	-
Guayacol	-	-	-

Tabla 2. Actividades enzimáticas de *Fomitopsis iberica*

Observaciones. Las diferencias entre *Trametes junipericola* y otras especies de *Trametes* (*T. hirsuta*, *T. pubescens*, *T. suaveolens*, *T. trogi* Berk., *T. hubarskyi*, *T. tephroleuca* y *T. palustris*) han sido puestas de manifiesto por Manjón, Moreno & Ryvarden (1984).

Una especie próxima es *Pilatoporus maroccanus* Kotlaba & Pouzar, descrita recientemente por Kotlaba & Pouzar (1993), del Atlas medio (Marruecos) sobre *Cupressus sempervirens*, comparte numerosos caracteres macro y microscópicos con *Trametes junipericola*, el basidioma con la superficie velutina, los poros grisáceos de 1-3 por mm, las medidas esporales ($7-9,3 \times 2,5-3,5 \mu\text{m}$) y el habitat sobre tronco de *Cupressaceae*. La principal diferencia es el tipo de podredumbre que Kotlaba & Pouzar (1993), indican es parda.

Vampola (1996), indica respecto a *P. maroccanus* Kotlaba & Pouzar que "Further specimens from Morocco or other countries where *Cupressus sempervirens* occurs are badly needed to prove that *P. maroccanus* is really an independent species and not only a synonym of *Trametes suaveolens*".

El género *Pilatoporus* fue creado por Kotlaba & Pouzar (1990) para aquellas especies del género *Fomitopsis* que poseen esporas de pared delgada y contexto blanco, estos autores también describen el género *Rhodoformis* segregado de *Fomitopsis* para aquellas especies con esporas de pared delgada y contexto rosado. Ryvarden & Gilbertson (1993) no reconocen estos dos géneros, considerando únicamente el género *Fomitopsis*, opinión que seguimos en este trabajo.

Vampola (1996) aporta nuevas localidades para *Pilatoporus ibericus* (Melo & Ryvarden) Kotlaba & Pouzar en Europa y Asia (República Checa, Eslovaquia, Croacia e Iran), y concluye su comportamiento como un hongo heterotálico y bipolar.

Finalmente podemos concluir después de los estudios enzimáticos realizados que *Trametes junipericola* está bien encuadrado en el género *Trametes*, porque posee podredumbre blanca, y es diferente de *Fomitopsis iberica*, que produce podredumbre parda.

Hasta el momento *Trametes junipericola* es un hongo endémico de los bosques de *Juniperus thurifera* L. de la provincia de Guadalajara, no siendo raro su recolección en los otoños lluviosos.

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***DIDYMIUM CLAVODECUS* (MYXOMYCETES) UNA ESPECIE AMERICANA NUEVA PARA EUROPA.**

M. LIZÁRRAGA, C. ILLANA, G. MORENO & A. CASTILLO

Dpto. Biología Vegetal. Facultad de Ciencias. Universidad de Alcalá,
28871 Alcalá de Henares, Madrid, España

ABSTRACT *Didymium clavodecus*, a species described from USA, has been found in Spain. *Didymium clavodecus* is described and illustrated by I. M. and S. E. M. photographs.

RÉSUMÉ *Didymium clavodecus*, espèce décrite aux U.S.A., a été observée en Espagne. *Didymium clavodecus* est décrite et illustrée par des photographies de microscopies optique et électronique à balayage.

KEY WORDS : *Didymium clavodecus*, Myxomycetes, Chorology, Taxonomy

En la actualidad estamos realizando diferentes estudios, que nos permitan comparar la diversidad existente en diferentes grupos de hongos en áreas mediterráneas de Europa (Extremadura, España) y América (Baja California, México), para ello hemos seleccionado áreas ocupadas por especies esclerófilas del género *Quercus*. *Didymium clavodecus* es un elemento folícola común entre ambos continentes, su aparición en España y por tanto en Europa es de importancia corológica para dicha especie originariamente descrita de California (USA).

Las microfotografías fueron realizadas con un microscopio electrónico de barrido (M.E.B.) Zeiss-DSM 950, y las muestras fueron preparadas con la técnica del punto crítico. Para describir la ornamentación esporal con M.E.B. hemos seguido la terminología propuesta por Rammeloo (1974, 1975). El material estudiado se encuentra depositado en el herbario de la Universidad de Alcalá (AH).

Didymium clavodecus Whitney, *Mycologia* 71: 1257, 1979. Figs. 1-10

Material estudiado ESPAÑA: En hojas de *Quercus ilex* L., Pozanco, Sigüenza, Guadalajara, leg. J. Alvarez, 10-III-1990, AH 12321. MÉXICO: En hojas y madera de *Quercus agrifolia* Née, ctra. Tecate-Ensenada km. 65 (Ejido Ignacio-Zaragoza), Baja California, 6-II-1993, leg. M. Lizárraga, G. Moreno y C. Illana, AH 15883, 15895 y 18627. En hojas de *Quercus agrifolia* Née, ctra. Tecate-Mexicali (Cañada Verde), Baja California, 6-II-1993, leg. M. Lizárraga, G. Moreno y C. Illana, AH 15824, 15827, 15828, 15923, 15924, 15926, 15927, 15935, 15943, 15950 y 18620.

Fructificaciones formando esporocarpos de 0,4-3 mm de diámetro cortos plasmódio-carpos, de 3-5 × 1,5-2 mm, sesiles, de pulvinados a anulados (figs. 1-2). Peridio indiscente, membranoso, cubierto por cristales calcáreos blancos, (figs. 3-4), dehiscencia irregular. Columela central blanca, formada por depósitos calcáreos. Hipotalo membranoso. Capilicio de 1,5-2,5 µm de diámetro abundante, de filamentos gruesos y rígidos, pardo violáceo oscuro, muy anastomosados, formando una red que conecta el centro de la fructificación con el peridio (figs. 5-6). Esporas de 12-14 µm de diámetro, poligonales, negras en masa, pardo-violáceo oscuras al M.O., con un retículo que recuerda a las esporas de *Phyvarium stramineum* ex Lister, (figs. 7-9), ornamentadas con espinas capitadas de 1-2 µm de largo, que al M.E.B. forman características "pila" (fig. 10).

Macroscópicamente *Didymium clavodecus* puede ser confundido macroscópicamente con otras especies sesiles del género *Didymium*, especialmente con formas sentadas de *D. squamulosum* (Alb. & Schwein.) Fr. Sin embargo *D. clavodecus* posee un carácter único en el género, como es la posesión de una espora de forma poligonal, ornamentada con espinas capitadas en forma de clavo, que son perceptibles al M.E.B. como "pila", y que varían en número y densidad (figs. 8-9).

No se conocen citas de *Didymium clavodecus* en Europa (Neubert & al., 1995). Únicamente se conoce esta especie de California, USA (Whitney, 1979), y más recientemente de Baja California, México (Lizarraga & al., 1997), siempre sobre hojas de *Quercus* sp.

El material español ha sido comparado con abundantes muestras recogidas por nosotros, en áreas mediterráneas de Baja California. Un estudio completo de *Didymium clavodecus* de Baja California ha sido realizado por Lizarraga & al. (1997).

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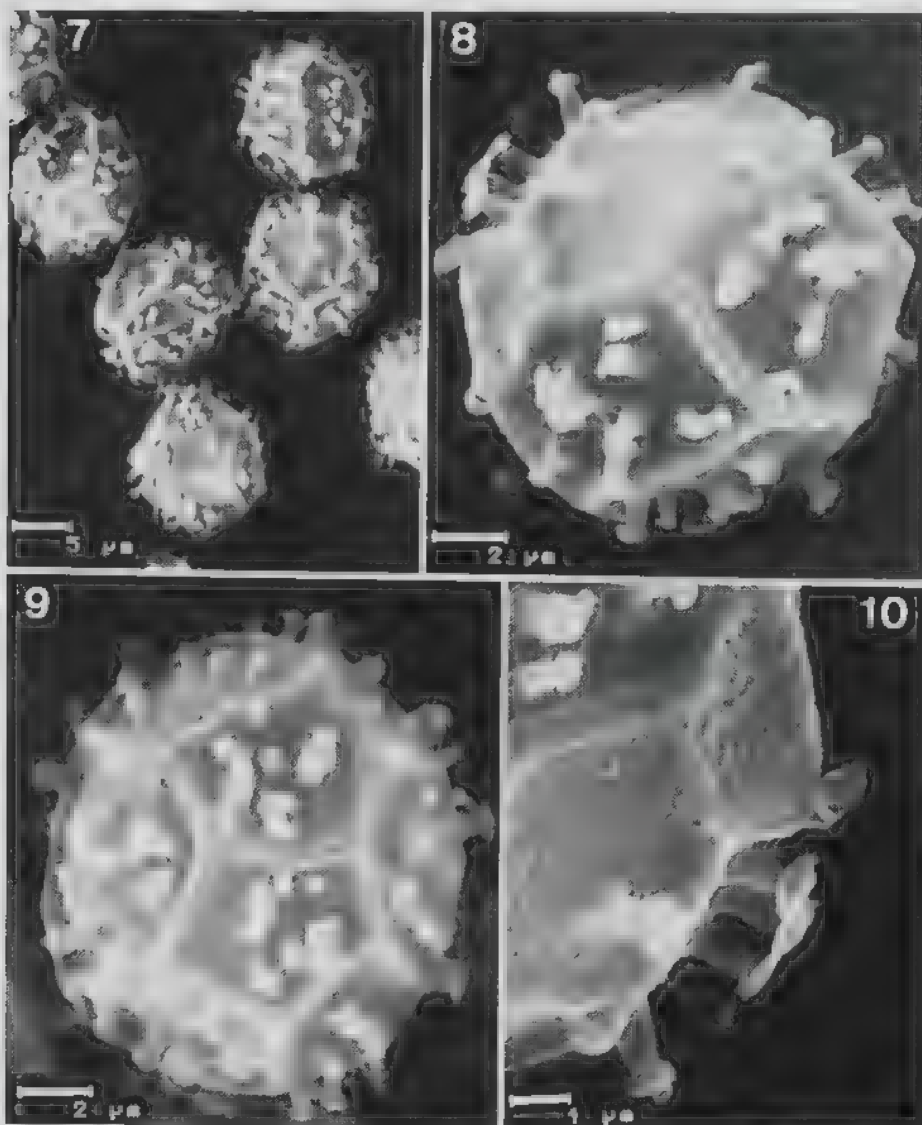
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Figs. 1-6. — *Didymium clavodecus* (AH 12321). 1 fructificaciones 2 esporocarpio sésil con cristales calcáreos sobre el peridio. 3: peridio cubierto con cristales (parte superior), con un filamento del capacio ácido (M E B) 4 cristales calcareos (M E B) 5 filamentos del capacio que forman una red (M E B) 6 detalle del capilicio (M E B)



Figs. 7-10. - *Didymium clavodecus* (AH 12321). 7 grupos de esporas (M.E.B.) 8-9 esporas (M.E.B.). 10: detalle de la ornamentación esporal formada por "pila" (M.E.B.)

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Cryptogamie, Mycologie - Rédacteur : Bruno Denetière

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